

**Feeding habits of New Zealand fishes: a literature review and  
summary of research trawl database records 1960 to 2000**

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## **EXECUTIVE SUMMARY**

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This report provides a comprehensive summary of dietary information available for New Zealand fish species up to and including the year 2000. The report does not review publications or research data held 2001–2011 as comprehensive feeding studies are already published elsewhere. This report does not provide any statistical analyses of sampling levels, dietary composition, variability, or potential error or bias in diet descriptions.

This is the fourth report in a series of data summaries about New Zealand fish species. The first three reports are atlases of the spatial distribution of a wide range of New Zealand marine fish and cephalopod species (Anderson et al. 1998, Bagley et al. 2000, Hurst et al. 2000).

This report includes a literature review of published information for all fish species included in the first atlas (Anderson et al. 1998), and summarises dietary records of 25 key species collected from research trawls (bottom, midwater and prawn) held in the Ministry of Fisheries research trawl database. The distribution of the feeding location and size of the fish sampled by area, and major dietary categories by area and fish size (where appropriate) are presented. These summaries exclude species for which there were fewer than 20 tows or 200 fish sampled.

## 1. INTRODUCTION

Understanding the structure and dynamics of marine ecosystems is critical to develop an ecosystem approach to fisheries management. It allows assessing the impacts of fishing activities and environmental factors not only on target stocks but also on all other species including by-catch and protected species belonging to the ecosystem. The structure of the ecosystem is based on prey-predator relationships among species. Studying trophic dynamics leads to the development of trophic models which can provide a tool to determine the effect of fishing on trophic structure. The first step in the development of trophic models is to quantify the dietary preferences of fish. Until recently, trophic studies on the scale of fisheries were not undertaken in New Zealand. As part of a series of data summaries on New Zealand fish that were collated and published in the late 1990s, a literature review of published feeding studies and a summary of dietary records held in the Ministry of Fisheries research trawl database was also conducted to undertake a stocktake on what was known about the dietary preferences of fish in New Zealand seas at that time.

This is the fourth report in a series of data summaries. The first three reports (Anderson et al. 1998, Bagley et al. 2000, Hurst et al. 2000) were atlases of the spatial distribution of a wide range of New Zealand marine fish and cephalopod species. These atlases summarised data from research and commercial sources, included bottom trawls, midwater trawls, aerial sightings and tuna longlines, and also presented adult and juvenile distributions. They were produced as background information to the analysis of fish community structure in the New Zealand region (*see* Bull et al. 2001, Beentjes et al. 2002, Francis et al. 2002).

The fourth report on feeding studies and dietary records remained unpublished, however. As interest in ecotrophic information has increased, the data held in the report has been increasingly sought. The current material presented here has largely drawn from the original unpublished report and makes the information summarised in the report publicly available.

Apart from several taxonomic updates to prey species, no other changes or updates to the dietary information contained in the original report have been made.

The report therefore provides a comprehensive summary of feeding information available in 2000 for New Zealand fish species that were included in the first atlas (i.e., those caught in bottom trawls, Anderson et al. 1998). No attempt is made in this document to further analyse feeding patterns (e.g. with respect to season), or determine feeding interactions between species. The report is organised into a summary of results and three Appendices:

Appendix A provides a summary of published and some unpublished dietary information for New Zealand fish species listed in Anderson et al. (1998). Relevant data from northern hemisphere studies were included if information about fishes found in New Zealand was minimal or lacking. Species are grouped by the four main depth assemblages identified by Francis et al. (2002), to facilitate comparison of species diets within a similar depth range.

Appendices B and C provide summaries of data on stomach contents of fish collected from research (bottom, midwater and prawn) trawls held on the Ministry of Fisheries research trawl database. Some additional non-digitised data is also included – *see* Table 2). These summaries comprise 25 key species for which there were sufficient feeding data (i.e., over 20 tows or 200 fish sampled in total).

Appendix B provides the location of feeding records collected from research trawls. It comprises length frequencies of the fish sampled by area, and major dietary categories by area and fish size (where possible). Data were not summarised by season as there were not sufficient data to do this for more than a few species.

Appendix C provides tables, by species, of all prey items recorded, by area from research trawls. Within each section, the species are ordered alphabetically by scientific name.

Readers should be aware that there have been a number of substantial dietary studies since 2000 that build considerable understanding of the trophic relationships among New Zealand's marine fish fauna. In particular, a comprehensive trophic study of the Chatham Rise has recently been conducted on a number of deepwater species, including species covered in this review (Dunn 2009, Dunn et al. 2010 a, b, c, Horn et al. 2010, Stevens and Dunn 2011).

## **2. METHODS**

### **2.1 Literature review (Appendix A)**

#### **2.1.1 General overview**

The main aim of the literature review was to supplement feeding information held on the Ministry of Fisheries research trawl database about New Zealand fish species included in the first fish distribution atlas (Anderson et al. 1998).

#### **2.1.2 Literature Sources**

For New Zealand fish dietary records we have cited the primary source, which may be from published papers, unpublished theses or reports. The only book used was 'A Treasury of New Zealand fishes' (Graham 1953) which provided useful data on the range of prey types consumed by a large number of fish species. A number of other popular books on New Zealand fishes have been published which provide general information on feeding (Parrott 1957, 1960, Moreland 1963, Heath & Moreland 1967, Moreland & Heath 1983, Paul & Heath 1985, Paul 2000, Ayling & Cox 1987, Doak 1984, 1991, Francis 1996) but these data were not included here.

The Aquatic Science and Fisheries Abstracts (ASFA) database was used extensively to source references. ASFA provided references for feeding studies on New Zealand fish species (including international and New Zealand literature) published since 1970. Earlier New Zealand studies were actively sought by other means, but no attempt was made to search out overseas references prior to 1970.

Feeding data collected during New Zealand trawl surveys and published in individual trawl survey reports have not as a rule been summarised as part of the literature review. Instead, all research trawl data have been summarised separately in Appendices B and C of the report.

#### **2.1.3 Data quality and interpretation**

The early records of fish feeding in New Zealand (Thomson 1892, Phillipps 1926, Graham 1938, 1953) were generally descriptive rather than quantitative assessments of fish diet. Some stomach content identification errors are likely to have been reported in these early records due to limited expertise. For example, the earliest data compiled from 12 000 stomachs of 40 fish species had been obtained from lighthouse keepers (Thomson 1892). Phillipps (1926) examined 114 stomachs from 10 species obtained from commercial fishers. In some cases, Phillipps was unable to supervise the data collection and there appears to have been confusion over the identification of red and scaly gurnard.

Records by Graham (1953) duplicated many of the observations from Graham (1938) and these data were therefore combined.

Uncertainty about species identification (both predator and prey) was often increased by the use of non-specific common names and redundant generic and specific names. For example, gemfish (*Rexea solandri*) was reported as New Zealand hake (Thomson 1891), kingfish (Graham 1938), or southern kingfish (Graham 1953); bluenose (*Hyperoglyphe antarctica*) as bream (Graham 1938, 1953), and rock cod (*Lotella rhacinus*) as hake (Thomson 1891). In cases where the fish species could not be determined from the common name provided, those data were excluded from this review. Some early references may also have been missed because of scientific name changes.

The following references were used extensively for prey species identification: seaweeds (Adams 1994), polychaetes (Glasby & Read 1998), molluscs (Suter 1913, 1915, Powell 1976, Roper et al. 1984), crustaceans (Wear & Fielder 1985, McClay 1988), fishes (Ayling & Cox 1987, Paulin et al. 1989) and general (Morton & Miller 1968, Barnes 1987).

Scientific name changes also caused difficulties where prey had been grouped. For example, natant decapods (shrimps and prawns) were grouped as suborder Natantia and later separated into the Infraorders Penaeidea, Stenopodidea, and Caridea (Barnes 1987). Similarly, cephalopods initially grouped into the suborders Octopoda (octopuses) and Decapoda (squid and cuttlefish) were later grouped into a number of subclasses and orders (Barnes 1987). In keeping with the majority of references cited, we grouped shrimps and prawns as natant decapods. The terms Cephalopod, Octopod and Decapod are used extensively in the literature and for consistency have been used in tables in this report. In instances where prey groups had been inappropriately grouped (e.g., euphausiids, cumaceans and mysids grouped as shrimps) the data were omitted.

#### 2.1.4 Data presentation

Summarising the published data was complicated by the wide range of methods used for feeding studies and different levels of detail presented by different authors. We standardised across reports by grouping and coding data as follows:

*Empty*: Everted stomachs and empty stomachs are not distinguished in the tables in Appendix A. This information was rarely available and, in a number of studies, fish with everted stomachs were discarded from further analysis (e.g., Sedberry & Musick 1978).

*Fish length*: In a number of studies, feeding data were divided into a number of predator length divisions (e.g., 1 cm size classes for *Parapercis colias*, Mutch 1983) to reflect size specific changes in diet. Although the diet of fish often changes as they grow, incorporating these data was beyond the scope of the current exercise. If overall totals were presented these were used in preference to the size class data.

*Method*: The variety of methods used to collect, analyse and present feeding data have been grouped and coded. To standardise the data summaries as much as possible, and to provide comparison for our New Zealand data, we chose percentage of frequency of occurrence (F) in stomachs containing food as the preferred method. Codes are as follows:

- B – percentage of occurrence by biomass. Edgar & Shaw (1995) calculated the biomass of mobile invertebrates in guts using biomass/sieve size relationships (Edgar 1990) and length/weight relationships. These data were then combined with percent occurrence data for plants and sessile animals.
- C – percentage of transect coverage. Choat & Clements (1992) agitated and spread the gut contents of each specimen evenly over a Perspex tray divided into 10 horizontal transects. Four transects were randomly selected and ‘the extent of each transect covered by particles of each food category’ was recorded as a percentage of the total amount per transect.

- D – percentage of diet composition. A semi-quantitative volumetric estimate based on assigning food points for each category according to stomach fullness, expressed as a percentage of total food points awarded to all stomachs. (e.g., Godfriaux 1968, 1969, 1974a, b, Baker 1971).
- F – percentage of frequency of occurrence of one or more individuals of each prey category in stomachs containing food (e.g., Rapson 1956, Godfriaux 1969, 1974a, b, Rosecchi et al. 1988, Hanchet 1991).
- F\* – percentage of frequency of occurrence of one or more individuals of each prey category in all stomachs. (e.g., Johnston 1983, Rojas et al. 1985). Also includes results from Webb (1973) who scaled down the percentage in stomachs with food so that the total percentage = 100.
- I – number of individual prey in a prey category (e.g., Clarke 1980, 1982).
- IRI – Index of relative importance which incorporates proportional occurrence, number and weight of prey (e.g., Yano 1993).
- N – percentage of number. The total number of individuals of each food category expressed as percentage of the total number of individuals in all food categories (e.g., Godfriaux 1974a,b, Rosecchi et al. 1988).
- P – presence of a food item in a stomach (e.g., Thomson 1892, Phillipps 1926, Graham 1938, 1953).
- V – percentage of volume composition. Food points for each category are expressed as a percentage of the total potential stomach volume (total number of stomachs x 20). This method provides the percentage of stomach volume occupied by each food item in the fishes diet (e.g., Godfriaux 1969, 1974a, b).
- W – percentage of weight. The total weight of each food category expressed as a percentage of total weight of all stomach contents (e.g., Rapson 1956, Rosecchi et al. 1988).

*Unidentified*: predominantly unidentified or unidentifiable prey items, and includes detritus (defined by Webb 1973, as organic matter not attributable to any group, but which was classified as food).

*Extraneous matter*: predominantly benthic substrate (i.e., sand, mud, gravel) but includes plastics, vinyls, wood, rubber, straw, leaves, grass, kelp, faecal pellets and even human remains (a finger).

*Algae*: simple photosynthetic, non-vascular plants, diatoms, dinoflagellates, and blue green algae.

*Seaweeds*: complex macroscopic algal assemblages, including the three main divisions: the green seaweeds (Chlorophyta), brown seaweeds (Phaeophyta), and the red seaweeds (Rhodophyta) (Adams 1994).

*References*: Note that all references marked with an asterisk (\*) were sourced from Cortés 1999.

*Invertebrate and teleost groups*: although identification to species level was sometimes available, data have mostly been combined into higher taxonomic groups. For the fish families, Macrouridae (rattails) and Myctophidae (lantern fishes) and the crustacean family Galatheidae, individual genera or species of significance have been included in some tables, alongside the appropriate prey grouping. For overseas references, if a teleost species was a significant prey item but the family was not present in New Zealand waters, the family was included in the table and marked with an asterisk (\*). If the species was of minor importance it was placed in the other fish category.

*Occurrence estimates*: In some papers, feeding data were presented graphically and values were estimated from pie charts and bar graphs. Where it was not possible to obtain complete data for a prey group the largest value of a family or species was used, and therefore the group total may be underestimated. This problem occurred often with the natant decapods group which comprise a large number of families and species.

Quantitative data were grouped to indicate 3 levels of relative importance of prey in the diet: low (under 10%), moderate (10–50%) and high (over 50%). These levels are indicated on the tables by increasing intensity of shading (Appendix A).

Feeding summaries are presented (Appendix A) for 108 New Zealand species and 97 species from elsewhere (divided into Northern or Southern Hemispheres). These summaries are also divided into four assemblages on the basis of mean depth of occurrence (*see* Anderson et al. 1998, Bagley et al. 2000) and the assemblage analysis of Francis et al. (2002). The depth ranges were: inshore (less than 100 m), continental shelf (100 to 400 m), upper-slope (400 to 700 m), and mid-slope (over 700 m). For species that were studied in New Zealand waters, there are 33 inshore species, 34 continental shelf, 16 upper slope, and 10 mid-slope species, and for New Zealand species that were studied elsewhere, there are 12 inshore, 18 continental shelf, 15 upper slope, and 44 mid-slope species. An alphabetical list of the species by the four depth ranges is given in Table 1.

Each diet table comprises two parts. The first part is a summary of the majority dietary groups. The subsequent one or two parts give more detailed taxonomic data for the following prey groups, where they existed: Mollusca, Crustacea, Echinodermata, Chondrichthyes, and Osteichthyes. For example, the first table in Appendix A (Table A1a) relates to feeding records for fish species found in 0–100 metres depths in New Zealand waters. The first three pages of this table summarises the major dietary groups for each species. The next six pages give more detailed prey data for molluscs, crustaceans, and cartilaginous and bony fish.

## 2.2 Research trawl data (Appendices B and C)

### 2.2.1 Species selection

An outline of the catch data contained in the Ministry of Fisheries research trawl database was given by Anderson et al. 1998. The database incorporates surveys from about 1960 to 2000, although the database is incomplete 1960—1980. Many of the feeding records from these early surveys have already been published and are cited in the literature review (Appendix A).

The usual sample size of dietary information about fish during research trawl surveys was 20 individuals per species, per tow but there are a few cases where larger numbers of fish were sampled (e.g., specific orange roughy feeding studies), or the number of species sampled was expanded.

A summary of the number of feeding records on the database up to the year 2000 is given in Table 2. Attention to detail and the identification of stomach contents has received differing priority on trawl surveys because of time and practical constraints and are therefore often patchy within surveys or inconsistent between surveys. The species sampled were usually those sampled for a full range of biological data (e.g. size, weight, spawning condition, otoliths) and are predominantly those of commercial importance. In selecting the species to include in this report, we used a minimum cutoff of 20 stations or 200 fish examined for a given species over all data. Using these criteria, 25 species were selected for inclusion here: black oreo, *Allocyttus niger*; alfonsino, *Beryx splendens*; red gurnard, *Chelidonichthys kumu*; lookdown dory, *Cyttus traversi*; ling, *Genypterus blacodes*; sea perch, *Helicolenus percoides*; orange roughy, *Hoplostethus atlanticus*; pale ghost shark, *Hydrolagus bemisi*; dark ghost shark, *H. novaezealandiae*; bluenose, *Hyperoglyphe antarctica*; banded stargazer, *Kathetostoma binigrasella*; giant stargazer, *K. giganteum*; hoki, *Macruronus novaezealandiae*; hake, *Merluccius australis*; southern blue whiting, *Micromesistius australis*; blue cod, *Parapercis colias*; hapuku, *Polyprion oxygeneious*; smooth oreo, *Pseudocyttus maculatus*; red cod, *Pseudophycis bachus*; gemfish, *Rexea solandri*; blue warehou, *Serirolella brama*; white warehou, *S. caerulea*; silver warehou, *S. punctata*; barracouta, *Thyrssites atun*; and Murphy's mackerel, *Trachurus murphyi*.



### **2.2.2 Research trawl data presentation**

Distribution maps of the feeding records are presented for each species in Appendix B. The start position was used to determine the location of the tow. The number recorded is the number of stations from which feeding data are available for that species. Overall, coverage by research trawls in the EEZ is extensive (see Anderson et al. 1998, Bagley et al. 2000), although the Fiordland coast, the Kermadec Ridge, and parts of the Challenger Plateau have not been well sampled.

Where sufficient feeding data was available the data was separated into geographic regions. These regions are largely self-explanatory however two regions need to be defined. The ‘Challenger’ region is defined as the Challenger Plateau and extends along the upper west coast of the South Island to Jackson Head. The ‘Southern N.Z.’ region is defined as south of Jackson Head on the west coast of the South Island and south of the Otago Peninsula on the East Coast.

The length frequency of the fish sampled for feeding and number of stomachs sampled is shown in Appendix B, to the right of the maps, by area where appropriate. Lengths recorded here were measured to the nearest centimetre below actual length by the following methods: total, fork, standard, and chimera (tip of snout to posterior end of the dorsal fin). Where the data were adequate, fish size has been divided into two or three arbitrary groups and summaries of percentage frequency of prey items, as a percentage of stomachs containing prey, is presented by area and fish size.

Detailed tables of feeding data, by area, for each species are presented in Appendix C. These tables include dietary summaries by the main prey groups, or more detailed identification of prey if available. For many species, prey items were only identified to higher taxonomic levels and some of these were quite broad (e.g., fish). Key taxa are identified in bold and summarise all subsequent prey or prey groups not in bold. Additional sub-groups not indented also contain summaries of subsequent prey or prey groups. Prey groups (e.g. teleost Families) which do not include totals for all the species in that group are indented.

In general, for invertebrates, the key taxa represent the top taxonomic level recorded in a Phylum (e.g., Phylum Crustacea, or Class Polychaeta in Phylum Annelida) and, for vertebrates, the top taxonomic level recorded in Class (e.g., Chondrichthyes, Osteichthyes etc.). For some invertebrate phyla, there are also summaries of “main” groups at a variety of lower levels, down to Superfamily in a few cases. Note that Class Cephalopoda, although a sub-group of the Phylum Mollusca, appears in bold as it contains a summary of the three further sub-groups. No attempt was made to combine teleosts by family, mainly because of the low proportions of teleosts identified, but they were grouped as mesopelagics and “others”. Summaries of data by the various taxonomic levels may be less than the total of the individual prey items added together because some stomachs contain more than one prey species in that group.

### **2.2.3 Sources of error and uncertainty**

Details of possible sources of uncertainty in the positions of bottom and midwater were outlined by Anderson et al. (1998) and Bagley et al. (2000) and are not repeated here. Many of the fish identification problems outlined in the previous two atlases are not relevant to the fish examined for feeding as the 25 key species chosen were common commercial species and the risk of misidentification very low.

Errors in feeding records may have occurred through:

- variable identification of prey. This may have been influenced by the availability of identification guides as well as the skill of seagoing personnel.
- detection errors and biases (i.e. small prey such as copepods may not be detected in visual inspections at sea).
- potential regurgitation (total or partial) may not always be routinely checked.
- mis-coding of prey
- data transcription or computer entry.

Appendix C uses the method of frequency of occurrence for the diet tables, a method which overestimates the importance of small prey. Any such errors are unlikely to affect the general patterns of feeding observed, although we did correct records of the pelagic octopus *Amphitretus* spp. (code AMP) which were generally incorrect due to confusion with the code for amphipods (APH).

Some errors in fish length measurement may have occurred during data collection, transcribing, or computer entry and not have been corrected during normal error checking procedures. To minimize problems, potential outliers were removed.

### **3. RESULTS**

#### **3.1 Diet summaries for the 25 key New Zealand species**

##### **General**

This summary by species is based on information contained in Appendices A–C. Only the 25 key species included in Appendices B and C are included below. The comparison of these findings (Appendices B and C) is made with other feeding studies that are summarised in Appendix A.

The distribution of feeding records and overall summary of diet by predator size and area of capture come from Appendix B. Descriptions of where fish are caught refer to the occurrence of fish in research trawls (i.e., the grey dots on the distribution plots) and do not necessarily reflect where fish are most abundant (Appendix B).

The details of the number of fish examined, stomach state, and percentage frequency of occurrence of prey in stomachs containing food come from Appendix C. Reference to the number of “main” invertebrate groups relates to the prey groups listed in Appendix C which include groupings at a variety of taxonomic levels from Class down to Superfamily.

The main diet for the 25 key species is summarised in Figure 1.

##### ***Allocyttus niger* black oreo**

Black oreo are caught in depths greater than 500 m mainly on the Chatham Rise, but extending northward along the south-east coast of the North Island and southward to the northern edge of the Pukaki Rise (Figure B1a). Feeding records are mainly available from the southern Chatham Rise (Figure B1a). A total of 6282 fish, mainly 20–45 cm total length, were sampled, of which only 14% had stomachs containing food (Figure B1b, Table C1). A high proportion of stomachs (67%) were everted (Table C1).

Overall, teleosts (48%), crustaceans (36%) salps (24%) and cephalopods (mainly squid, 6%) were important in the diet (Table C1). Mesopelagics (5%), mainly myctophids, were the most commonly identified teleost. Non-mesopelagics made up less than 1 % of those identified. Crustaceans were

mainly natant decapods (22%). In total, at least 12 main invertebrate groups in four phyla and eight teleost species were recorded (Table C1).

The relative proportion of teleosts increased and tunicates decreased slightly as fish grew (Figure B1c, Table C1). Relative importance of prey was slightly different between areas: crustaceans and salps were more important on the Chatham Rise (45% and 31% respectively) than off Southern N.Z. (20% and 10%); teleosts and cephalopods were more important off Southern N.Z. (72% and 13%) than on the Chatham Rise (34% and 2%) (Figure B1c, Table C1).

These findings differ from those reported by Clark et al. (1989) who found that salps (68%), amphipods (72%) and natant decapods (42%) were the most important items in the diet of black oreo from the south-west Chatham Rise (Table A4a).

### ***Beryx splendens alfonso***

Alfonso are caught mainly on the northern Chatham Rise, east coast North Island and WCSI (Figure B2a). Feeding records are available from a few samples around the North Island and Chatham Islands (Figure B2a). A total of 365 fish (mainly 20–50 cm fork length) were sampled, of which 44% had stomachs containing food (Figure B2b, Table C2).

Overall, crustaceans were the most important (over 60%) prey group of smaller (up to 30 cm) and larger fish around the North Island (Figure B2c, Table C2). However, in larger Chatham Rise fish, teleosts were the most important (79%) prey, with crustaceans and cephalopods about equal at 18 and 15% (Figure B2c, Table C2).

Invertebrates from North Island samples were mainly natant decapods (81%) with some euphausiids and squid (both 3%) and mysids (2%) (Figure B2c, Table C2). Teleosts (30%) were mainly mesopelagic myctophids (23%). On the Chatham Rise, natant decapods were also the most important (15%) crustacean but squid importance increased to 15% and teleosts to 79% (Figure B2c, Table C2). Identified teleosts were all mesopelagics (33%), mainly *Maurolicus australis* (24%) with some *Photichthys argenteus* (9%). In total, at least five main invertebrate groups in two phyla and two mesopelagic teleost species were identified (Table C2).

These findings are similar to records from the Naska Seamount (SE Pacific) where crustaceans, mainly euphausiids and natant decapods were important, with teleosts and squid also recorded (Dubochkin & Kotlyar 1989) (Table A3b). The main teleost genus recorded was *Diaphus*, a myctophid.

### ***Chelidonichthys kumu* red gurnard**

Red gurnard are caught in inshore shelf areas around New Zealand and the Chatham Islands. Feeding records are available only from the ECSI (Figure B3a). A total of 987 fish (mainly 20–55 cm fork length) were sampled, of which 30% had stomachs containing food (Figure B3b, Table C3).

Crustaceans comprised 57% overall; those identified were mainly *Munida* spp. (20%), *Ovalipes catharus* (18%) and unidentified crabs (12%) (Table C3). Teleosts comprised 43% overall. Red cod was the most commonly identified (6%). Smaller (up to 35cm) fish and larger fish had a similar diet. In total at least five main invertebrate groups in two phyla and six species of fish were identified (Figure B3c, Table C3).

The importance of crustaceans in the diet of red gurnard is consistent with other records from New Zealand. Godfriaux (1968, 1970) found Hauraki Gulf fish had been eating mainly crustaceans, mostly crabs and decapods and some fish (Table A1a). On the WCSI, Ingerson (1996) found that brachyuran crabs and polychaetes were important (Table A1a). These studies include areas not covered by the Ministry of Fisheries research trawl database but many areas remain un-sampled.

### ***Cyttus traversi* lookdown dory**

Lookdown dory are caught mainly in 200–800 m depth around New Zealand, on the Challenger Plateau and in the Sub-Antarctic (Figure B4a). Feeding records are patchy and are mostly from around the North Island (Figure B4a). A total of 549 fish, (mainly 15–55 cm total length) were sampled, of which 38% had stomachs containing food (Figure B4b, Table C4).

Around the North Island, crustaceans were the dominant prey group (82% overall) in the diet and were mainly natant decapods (74%) (Figure B4c, Table C4). Teleosts comprised 20% overall, the most commonly identified were myctophids and silver roughy, *Hoplostethus mediterraneus* (both 2%). Teleosts also increased in importance in larger fish (36 cm or above) (Figure B4c). The small number of samples from other areas is not adequate to assess relative importance of prey groups. In total, at least five main crustacean groups, and seven fish species were identified (Table C4).

At the time of this review, there were no other feeding records known for this species in New Zealand. However a study of *C. traversi* off Tasmania, also found that crustaceans, in particular natant decapods, and fish, in particular myctophids, were important (Blaber & Bulman 1987) (Table A3b).

### ***Genypterus blacodes* ling**

Ling are caught in shelf and slope areas around New Zealand, the Chatham Islands, and the Sub-Antarctic (Figure B5a). Feeding records are available from most of these areas but are patchy from around the North Island and rare from depths under 200 m (Figure B5a). About 18 000 fish (mainly 15–165 cm total length) were sampled, of which 40% had stomachs containing food (Figure B5b, Table C5).

Teleosts were the most important prey, comprising 65% overall (Table C5). Of the teleosts identified, macrourids were an important group (6%, possibly up to 9% including individual species), and hoki (3%) was the most commonly identified species. The proportion of prey identification beyond phylum level was however very low. Mesopelagics were infrequently identified in the diet (0.1%). Crustaceans were also important (37%) with natant decapods (15%) and *Munida* spp. (15%) the most commonly identified. Cephalopods, particularly squid, were the next most common (3%) prey group. In total, at least 19 main invertebrate groups in six phyla, and over 50 fish species were identified (Table C5).

Teleosts were more important (75%) and crustaceans were less important (27%) in Southern N.Z. than in other areas (Figure B5c, Table C5). In all areas except Southern N.Z., there was a trend of changing dominance of prey groups from crustaceans in the smallest fish (up to 70 cm) to teleosts in the largest fish (111 cm or above). In Southern N.Z., teleost prey always dominated but may also be more important in larger fish (Figure B5c, Table C5).

The main prey groups identified are consistent with other records from around New Zealand (Mitchell 1984, Clark 1985) and Australia (Blaber & Bulman 1987). Mitchell (1984) also found crustaceans to decrease in relative importance between northern and southern New Zealand (Tables A3a, A3b).

### ***Helicolenus percoides* sea perch**

Sea perch are caught in shelf and slope waters down to about 800 m depth around New Zealand and on the Challenger Plateau, but not in the Sub-Antarctic (Figure B6a). Feeding records are patchy and are mainly from the ECSI (Figure B6a). About 300 fish (mainly 15–45 cm total length) were sampled, of which 42% had stomachs containing food (Figure B6b, Table C6).

The main prey items recorded from the ECSI were crustaceans (62%), mainly *Munida* spp., and teleosts (18%) and the diet was similar for smaller (up to 30 cm) and larger fish (Figure B6c, Table C6). The small number of samples from other areas is not adequate to assess relative importance of

prey groups in these areas but, overall, salps, crabs, and squid individually comprised more than 4% of the diet. In total, at least seven main invertebrate groups in four phyla and two fish species were identified (Table C6).

Graham (1938, 1953) recorded crustaceans, particularly decapods and crabs, as important in the diet of sea perch caught off Otago (Table A3a).

### ***Hoplostethus atlanticus* orange roughy**

Orange roughy are caught in depths over about 800 m around New Zealand (Figure B7a). Feeding records are available from most of these areas except that the Sub-Antarctic samples are patchy (Figure B7a). A total of almost 106 000 fish (mainly 10–45 cm standard length) were sampled, of which 29% had stomachs containing food (Figure B7b, Table C7).

Dominant prey groups were crustaceans, teleosts and cephalopods (Figure B7c, Table C7). The relative proportion of the prey groups changed with fish size; smaller fish (up to 20 cm) ate more crustaceans whilst larger fish (31 cm and above) ate more teleosts and cephalopods. Relative proportions of the three prey groups were similar between areas (Figure B7c, Table C7).

Crustaceans were the most important prey, comprising 58% overall (range by area 55–62%) (Table C7). Of those identified, natant decapods were the most important (33%), particularly *Pasiphaea* and *Sergestes* spp., followed by euphausiids (5%), amphipods (4%) and mysids (2%). The Southern N.Z. area had the lowest proportion of natant decapods (22%) and the highest proportion of amphipods (19%) recorded. Molluscs (10%), particularly squid (9%), were also important. A total of at least 21 main invertebrate groups in seven phyla were identified (Figure B7c, Table C7).

Teleosts comprised 41% of the diet overall (range by area 36–52%) (Table C7). A large number of species have been identified, including at least 35 mesopelagic and 48 others. Individually they comprise low percentages, but the most commonly identified groups were myctophids (2%) and macrourids (nearly 1%, including individual species). At least four species of elasmobranch were recorded, but totalled less than 0.1% (Table C7).

These findings are consistent with other records from around New Zealand (Rosecchi et al. 1988), i.e., crustaceans, particularly natant decapods, were the most important prey group in areas (Table A4a). Other published work did not fill in the gap in knowledge from Sub-Antarctic areas of New Zealand. Some seasonal studies have been published from the Challenger area (Rosecchi et al. 1988) and this could be expanded with data from the research trawl database, although most recent data are probably from winter months (Table A4a). An Australian feeding study (Bulman & Koslow 1992), found that teleosts were more important than crustaceans by weight, and that this increased in adult-sized fish (Table A4b). This is consistent with the change in importance of crustaceans by size in New Zealand fish.

### ***Hydrolagus bemisi* pale ghost shark**

Pale ghost shark are caught mainly in slope waters of about 300–1200 m depth around New Zealand, on the Challenger Plateau, and in the Sub-Antarctic (Figure B8a). A total of 109 fish (mainly 30–100 cm chimera length) have been sampled, of which 71% had stomachs containing food (Figure B8b, Table C8). The main prey items recorded were salps (39%), crustaceans (29%, mainly crabs at 13%), echinoderms (23%, mainly starfish at 14%), and molluscs (16%, including and squid). Teleosts were unimportant at only 1%. The small sample size from other areas was not adequate to assess relative importance of prey groups by area or fish size (Figure B8c). In total, 12 main invertebrate groups in five phyla and one fish species were identified (Table C8).

At the time of this review, there were no other feeding records known for this species or genus. The range of prey groups was similar to that recorded here for *Hydrolagus novaezealandiae* but

differences in the relative importance of various prey groups are inconclusive given the small sample sizes.

### ***Hydrolagus novaezealandiae* dark ghost shark**

Dark ghost shark are caught mainly on the shelf edge, in 100–500 m depth around New Zealand, on the Challenger Plateau, and in the Sub-Antarctic (Figure B9a). A total of 226 fish (mainly 30–75 cm chimera length) have been sampled, of which 40% had stomachs containing food (Figure B9b, Table C9).

The main prey items recorded were crustaceans (57%), mainly *Munida* spp. (21%) and crabs (19%) (Table C9). Teleosts (11%, including mesopelagics), molluscs (9%, including bivalves and octopods), echinoderms (8%, mainly starfish), polychaetes (8%), and salps (7%) were also important. The small number of samples is not adequate to assess relative importance of prey groups by area or fish size (Figure B9c). In total, at least 10 main invertebrate groups in five phyla and one teleost family were identified (Table C9).

At the time of this review, there were no other feeding records known for this species or genus. The prey groups were similar to that recorded here for *Hydrolagus bemisi* but differences in the relative importance of various prey groups are inconclusive given the small sample sizes.

### ***Hyperoglyphe antarctica* bluenose**

Bluenose are caught in slope waters, mainly 300–700 m depth, around New Zealand, on the Challenger Plateau, on the Chatham Rise (Figure B10a). They are not recorded from Sub-Antarctic. A total of 224 fish (mainly 45–95 cm length) have been sampled, of which 28% had stomachs containing food (Figure B10b, Table C10).

The main prey items recorded were salps (55%), cephalopods (27%, mainly squids), teleosts (18%, both benthic and mesopelagic at 8 and 10% respectively), and natant decapod crustaceans (13%) (Table C10). The small sample size was not adequate to assess relative importance of prey groups by area or fish size (Figure B10c). In total, five main invertebrate groups in four phyla and at least six fish species were identified (Table C10).

Graham (1938, 1953) recorded tunicates, mollusks, crustaceans, and fish present in bluenose from Otago, New Zealand (Table A3a). Winstanley (1978) also recorded salps as the main (over 50%) prey, with molluscs, crustaceans, and fish also important (each over 10%) (Table A3b).

### ***Kathetostoma binigrasella* banded stargazer**

Banded stargazer are recorded mainly from the Southland shelf, although a few have been recorded at the Chatham Islands (Figure B11a). Feeding records are entirely from Southern N.Z. A total of 406 fish (mainly 20–70 cm total length) were sampled, of which 91% had stomachs containing food (Figure B11b, Table C11). Feeding records should be treated with caution as stargazers are voracious feeders and freshly caught prey taken in the trawl net may have been included in the feeding records.

Overall, cephalopods were the most important prey (66%), comprising mainly squid (59%) but including octopods (10%) (Table C11). Teleosts were the next most important (52%) group. The main species identified was opal fish, *Hemerocoetes* spp. (4%). Minor groups included salps (3%) and crustaceans (2%). In total, at least six main invertebrate groups in three phyla and 28 teleost species were identified (Table C11).

For both size categories, teleosts and cephalopods were the dominant groups identified (Figure B11c, Table C11). Although the proportion of cephalopods doubled in larger (46 cm and above) fish, some

of this increase could be attributable to a reduction in the relatively large proportion of unidentified items (about 20% in smaller fish) (Figure B11c).

There are no other feeding records known for this species. The diet described here is similar to that of the giant stargazer, *Kathetostoma giganteum*, from Southern N.Z., described below.

### ***Kathetostoma giganteum* giant stargazer**

Giant stargazer are caught in shelf and upper slope areas around New Zealand, on the Challenger Plateau, on the Chatham Rise, and occasionally around the northernmost Sub-Antarctic Islands (Figure B12a). Feeding records are available mainly from the ECSI and Southland, with a few from the Chatham Rise. A total of 3470 fish (mainly 15–80 cm total length) were sampled, of which 79% had stomachs containing food (Figure B12b, Table C11). Feeding records should be treated with caution as stargazers are voracious feeders and freshly caught prey taken in the trawl net may have been included in the feeding records.

Teleosts were the most important prey, comprising 58% overall (Table C11). The main species identified were opal fish, *Hemerocoetes* spp. (3%), silver conger, *Gnathophis habenatus*, and red cod, *Pseudophycis bachus* (both 2%). Cephalopods were the next most important group (38%), mainly squid (34%) but including octopods (5%). Crustaceans occurred in 12% of stomachs overall, with crabs (7%) and *Munida* spp. (2%) the most commonly identified. In total, at least 19 main invertebrate groups in seven phyla and 40 teleost (including one mesopelagic) species were identified (Table C11).

For all areas, teleosts were important but the relative importance decreased southwards, from 82% on the Chatham Rise to 48% off Southern N.Z. (Figure B12c, Table C11). Cephalopods (mainly squid) were also important in all areas (17–53%), reaching the maximum off Southern N.Z. which was about equal (53%) to teleosts. Crustaceans were represented in all areas but ranged from 4–15%. Salps were recorded mainly from Southern N.Z. (6%). There were no clear changes in diet with fish size (Figure 12c, Table C11).

There is little other published information on New Zealand stargazer diet. Thomson & Anderton (1921) and Graham (1938, 1953) reported the presence of similar prey groups off Otago (Table A2a). Feeding in WCSI, North Island and Sub-Antarctic areas remains unknown. The diet described here is similar to that of the banded stargazer, *Kathetostoma binigrasella* from Southern N.Z., described above.

### ***Macruronus novaezelandiae* hoki**

Hoki are caught in slope areas around New Zealand, mainly on the Chatham Rise, the southern plateau and west coast of the South Island (Figure B13a). Feeding records are available from most of these areas except that records from the Challenger Plateau and North Island are patchy and there are few records from deepwater (over 800 m) on the Chatham Rise. A total of 33 745 fish (mainly 30–120 cm total length) were sampled, of which 42% had stomachs containing food (Figure B13b, Table C12). The proportion of empty stomachs was high (80%) off the west coast of the South Island, probably because many of the fish were sampled during spawning.

Teleosts were the most important prey, comprising 60% overall (Table C12). Of those identified, mesopelagics were important (15%) in the overall diet, particularly myctophids (about 13%) and particularly around the North Island (36%). Crustaceans comprised 43% of the diet overall, mainly natant decapods (27%) and euphausiids (12%). Cephalopods, particularly squid (5%) and salps (3%), were the next most common invertebrate prey groups. In total, at least 15 main invertebrate groups in five phyla and 49 teleost (including 15 mesopelagic) species were identified (Table C12).

Teleosts were important in all areas (range 55–69%), particularly in the north and west (North Island and Challenger) (Figure B13c, Table C12). Crustaceans were also important in all areas (range 28–

49%), but slightly less so to the west. Natant decapods were important in all areas (25–29%) but euphausiids were only commonly reported from the Chatham Rise (20%). There were no major differences in diet between size groups of fish (Figure B13c, Table C12).

The dominance of teleosts and crustaceans in the diet is consistent with results from the other main studies around New Zealand (Kuo & Tanaka 1984, Clark 1985) (Table A3a) and Tasmania, Australia (Bulman & Blaber 1986) (Table A3b), although Clark (1985) also found salps to be of moderate importance on the Campbell Plateau (Table A3a). The New Zealand studies, combined with the data on the research trawl database, could be used to examine seasonal trends in feeding of hoki in more detail.

### ***Merluccius australis* hake**

Hake are caught in slope areas around New Zealand, mainly on the Chatham Rise, the southern plateau and west coast of the South Island (Figure B14a). Feeding records are available from most of these areas except that records from the Challenger Plateau and North Island are patchy. There are also few records from deepwater (over 800 m) on the Chatham Rise and WCSI and all shallow (under 200 m) areas where hake occur (Figure B14a). A total of 7452 fish (mainly 30–130 cm total length) were sampled, of which 31% had stomachs containing food (Figure B14b, Table C13).

Teleosts comprised 85% of the diet overall (Table C13). Of those identified, hoki (22%) and javelinfish, *Lepidorhynchus denticulatus* (13%), were the most common. Mesopelagics were relatively unimportant (under 2%) in the diet. Cephalopods, mainly squid (9%) were also important and included a variety of genera other than *Nototodarus*. Crustaceans comprised mainly natant decapods (5%). In total, at least eight main invertebrate groups in five phyla and 38 teleost (including five mesopelagic) species were identified (Table C13).

Teleosts made up more than about 70% of the diet in all areas and across all size groups of hake (Figure B14c, Table C13). The main species identified, hoki, varied in importance from 15% off Southern N.Z. to 35% on the WCSI. Crustaceans comprised about 10–25% of the diet of small (up to 60 cm) hake but decreased in importance with increasing hake size. Teleosts and cephalopods showed a corresponding increase in importance with increasing hake size, in some areas (Figure B14c, Table C13).

At the time of this review, there were no other published studies of hake feeding in New Zealand that do not include data summarised here. A feeding study on the same species off Chile, using percentage of stomach mass (Paya 1992), found similar results: teleosts comprised at least 70% of the diet and the importance increased with increasing fish size; merluccids were the most commonly recorded teleost prey; squids were also important (although decreasing in importance with increasing fish size); and natant decapods were the main crustacean comprising 5% of the diet (Table A3b).

### ***Micromesistius australis* southern blue whiting**

Southern blue whiting are caught mainly in 200–700 m depths in the Sub-Antarctic, although they also occur infrequently on the Chatham Rise (Figure B15a). Feeding records reflect this distribution. A total of 5857 fish (mainly 15–60 cm total length) were sampled, of which 40% had stomachs containing food and 16% had stomachs which had everted (Figure B15b, Table C14). For both size groups, crustaceans and teleosts were the dominant prey groups (Figure B15c).

In the Sub-Antarctic, crustaceans comprised 70% of the diet, mainly euphausiids (37%), natant decapods (24%) and amphipods (11%) (Figure B15c, Table C14). Teleosts comprised 32%, of which most of those identified were mesopelagics (10%), mainly myctophids. Salps (7%) and cephalopods (2%) were of less importance. On the Chatham Rise, only seven of the 43 stomachs contained food, mainly crustaceans and teleosts (Figure B15c). In total, 10 main invertebrate groups in four phyla, and 10 teleost (including four mesopelagic) species were identified (Table C14).



The overall diet described here is consistent with other records from the Sub-Antarctic waters of around New Zealand (Clark 1985, Shpak 1976) (Table A3a). Shpak (1976) found more teleosts (mainly myctophids) in the diet of Bounties fish and correspondingly less crustaceans (particularly less amphipods) (Table A3a).

### ***Parapercis colias* blue cod**

Blue cod are caught in inshore shelf waters around the mainland and at the Chatham Islands (Figure B16a). They occur primarily over rough ground and are therefore not readily accessible to trawls, but a few feeding records were available from the ECSI and Southern N.Z. (Figure B16a). A total of 131 fish (mainly 25–55 cm total length) were sampled, of which 46% had stomachs containing food (Figure B16b, Table C15).

Overall, teleosts were the most important (47%), with opal fish, *Hemerocoetes* spp., (3%) as the main species identified (Table C15). Crustaceans (23%), mainly *Munida* spp. (12%), and tunicates (17%) were also important. Sample sizes were too small to allow between area comparisons (Figure B16c). In total, at least six main invertebrate groups in three phyla and four teleost species were identified (Table C15).

There are many reports of feeding of blue cod in New Zealand waters (Thomson & Anderton 1921, Graham 1938, 1953, Rapson 1956, Mutch 1983, Russell 1983) (Table A2a). These may not be directly comparable to our findings as the fish may have been caught over different habitats. However, prey groups recorded as being important by these authors are teleosts, crabs, mysids, bivalves, octopods, echinoderms, ctenophores and salps.

### ***Polyprion oxygeneious* hapuku**

Hapuku are caught over the shelf around the mainland and on the Chatham Rise (Figure B17a). They occur primarily over rough ground and are therefore not readily accessible to trawls, especially as adults. Feeding records are available mainly from Southland but include some from the Chatham Rise (Figure B17a). A total of 319 fish (mainly 50–110 cm total length) were sampled, of which 47% had stomachs containing food (Figure B17b, Table C16).

Overall, teleosts were the most important (68%), with red cod (18%) the main species identified (Table C16). Molluscs (25%), mainly squid (20%), and crustaceans (18%), mainly crabs (10%) and *Munida* spp. (7%) were also important. Most fish off Southern N.Z. were small (up to 80 cm) so diet was not summarised by size groups (Figure B17b). Comparison of diet by area was restricted because of the small sample size from the Chatham Rise (Figure B17c). However, feeding from both areas was dominated by fish, with cephalopods of secondary importance. In total, at least six main invertebrate groups in two phyla and 13 teleost species were identified (Table C16).

Other feeding records of hapuku in New Zealand waters are mainly from Cook Strait (Thomson 1892, Thomson & Anderton 1921, Graham 1938, 1953, Johnston 1983) (Table A2a). These studies also found fish (particularly macrourids and myctophids), squid, and crustaceans, particularly *Munida gregaria* and natant decapods to be important (Table A2a). A study of similar sized hapuku in the south-east Pacific (Rojas et al. 1985) recorded similar prey groups but included bryozoans and coelenterates (Table A2b).

### ***Pseudocyttus maculatus* smooth oreo**

Smooth oreo are caught in depths over about 500 m mainly on the Chatham Rise, the south-east coast of the North Island, Southern N.Z. and the WCSI (Figure B18a). Feeding records are mainly available from the Chatham Rise and Southern N.Z. (Figure B18a). A total of 8125 fish (mainly 15–60 cm total length) were sampled, of which 31% had stomachs containing food and 26% were everted (Figure B18b, Table C17).

Salps comprised 80% of the diet overall. Molluscs were the next most important (9%), comprising mainly of squid (8%) but including octopods (Table C17). Teleosts comprised 5% overall, with mesopelagics the most important group identified. Coelenterates comprised 4% and crustaceans 3%, mainly amphipods and natant decapods. In total, at least 11 main invertebrate prey groups in six phyla and seven teleost species were recorded (Table C17).

Diet of fish on the Chatham Rise was dominated by salps, regardless of fish size, but larger fish (31 cm or above) off Southern N.Z. had a more varied diet, dominated by teleosts but including tunicates, cephalopods and echinoderms as important (over 10%) groups (Figure B18c). The few samples from the North Island also indicated a more varied diet with tunicates, cephalopods and coelenterates important (Figure B18c).

These findings are similar to those of Clark et al. (1989) who recorded mainly salps (82%) as being the most important item in the diet of about 240 smooth oreo from the south-west Chatham Rise (Table A4a). However, their study also recorded amphipods as important (38%) whereas crustaceans were relatively unimportant (3%) in our findings.

### ***Pseudophycis bachus* red cod**

Red cod are caught in shelf areas around New Zealand, shallower parts of the Chatham Rise, and occasionally around Sub-Antarctic Islands (Figure B19a). Most feeding records are available from the ECSI with some from Southern N.Z. and a few from the Chatham Rise (Figure B19a). A total of 3703 fish (mainly 10–75 cm total length) were sampled, of which 50% had stomachs containing food (Figure B19b, Table C18).

Crustaceans were the most important prey group, comprising 79% overall, and were mainly *Munida* spp. (61%) (Table C18). Teleosts comprised 25% overall, the most commonly identified species were red cod, stargazer, tarakihi and witch. In total, at least 12 main invertebrates groups in three phyla and 23 species of fish (non-mesopelagic) were identified (Table C18).

There was some indication that crustaceans may become less dominant in the diet of larger (41 cm or above) ECSI fish; samples from other areas were insufficient to determine any trends (Figure 19c). *Munida* spp. was more important in the diet of fish on the ECSI (64%) compared to Southern N.Z. (22%) where other invertebrates such as crabs (24%) and squid (11%) were also important (Figure B19c, Table C18).

The importance of *Munida* spp. in the diet of ECSI red cod is consistent with other records from this area ECSI (Graham 1938, 1953, Habib 1975) (Table A2a). Habib (1975) also sampled off the WCSI and a few east coast North Island areas not represented in this study. Crustaceans and teleosts are also important in these areas, although the predominance of *Munida gregaria* decreases and is replaced by natant decapods and mysids on the east coast of the North Island and merlucciids on the WCSI (Table A2a). Areas not well sampled to date include the North Island, the Chatham Rise and Sub-Antarctic Islands.

### ***Rexea solandri* gemfish**

Gemfish are caught in outer shelf areas around New Zealand, and infrequently on the Chatham Rise (Figure B20a). Feeding records are available only from Southland. A total of 1109 fish (mainly 30–105 cm fork length) were sampled, of which 52% had stomachs containing food (Figure B20b, Table C19).

Teleosts were the most important prey, comprising 63% overall (Table C19). Of those species identified, *Trachurus* spp. (over 6%) and hoki (3%) were the most common. A few mesopelagics (2%) were also recorded. Cephalopods, all squid (43%), were also important and were mainly *Nototodar* species (over 27%). Other invertebrate groups made up less than 1%. In total, at least 20

teleost species and three main invertebrate groups in three phyla were identified (Table C19). There was no consistent trend in change of diet between small (up to 71 cm), medium and large (91 cm and above) gemfish (Figure B20c).

A variety of fish species and squids were recorded as present in other New Zealand studies from Otago (Thomson & Anderton 1921, Graham 1938, 1953) and Cook Strait (Phillips 1926). *Munida gregaria* were also recorded from Otago (Graham 1938, 1953) (Table A2a).

### ***Seriolella brama* blue warehou**

Blue warehou are caught mainly in shelf areas around New Zealand and infrequently on the Chatham Rise and in the Sub-Antarctic (Figure B21a). Feeding records are available only from Southern N.Z. (Figure B21a). A total of 974 fish (mainly 25–65 cm fork length) were sampled, of which 87% had stomachs containing food (Figure B21b, Table C20).

Salps were the main (97%) prey group; the only other group to exceed 1% occurrence was crustaceans (7.5%), mainly euphausiids (7%) (Table C20). There were no major differences in diet between three size classes of blue warehou (Figure 21c). A total of at least six main invertebrate groups in two phyla and one teleost species were recorded (Table C20).

The most comparable New Zealand feeding study is that of South Island fish by Gavrilov & Markina (1979) who found similar results: salps made up more than 50% of the diet by weight, with euphausiids of moderate importance (actually 27% by weight) (Table A2a). Other New Zealand feeding studies include presence records from Portland Island (Thomson 1892) and Otago (Thomson & Anderton 1921, Graham 1938, 1953) which include a variety of invertebrate groups not recorded here (polychaetes, molluscs, natant decapods) (Table A2a).

### ***Seriolella caerulea* white warehou**

White warehou are caught on the Chatham Rise, off Southland and in the Sub-Antarctic (Figure B22a). They occur infrequently on the WCSI and south-east coast of the North Island. Feeding records are available mainly from the Chatham Rise and Southland (Figure B22a). A total of 262 fish (mainly 20–50 cm fork length) were sampled, of which 68% had stomachs containing food (Figure B22b, Table C20).

Salps were the main (96%) prey and this was consistent for the two main sampling areas and for both size groups (Figure B22c, Table C20). The only other prey groups to exceed 1% occurrence were crustaceans (6%), mainly amphipods (3%) and euphausiids (1%), and unidentified teleosts (2%). A total of at least three main invertebrate groups in two phyla and one teleost species were recorded (Table C20).

Gavrilov & Markina (1979) also found salps were the main prey of white warehou (actually 98% by weight) in fish 30–64 cm, with amphipods also present (Table A3a).

### ***Seriolella punctata* silver warehou**

Silver warehou are mainly caught in shelf and upper slope areas around central and southern New Zealand and on the Chatham Rise (Figure B23a). Catches from the north of the North Island and the Sub-Antarctic are sporadic. Feeding records are available mainly from the Chatham Rise and Southland (Figure B23a). A total of 2022 fish (mainly 20–60 cm fork length) were sampled, of which 90% had stomachs containing food (Figure B23b, Table C21).

Salps were the main (97%) prey and this was consistent for the two main sampling areas and for all size groups (Figure B23c, Table C21). The only other prey groups to exceed 1% occurrence were

polychaetes (1.4%) and crustaceans (3%), mainly euphausiids. A total of at least 10 main invertebrate groups in six phyla and one teleost species were recorded (Table C21).

Gavrilov & Markina (1979) also found salps were the main prey of adult silver warehou (Table A2a). However, they also examined small fish, 12–18cm, and found other invertebrate prey groups were important: amphipods (75%) chaetognaths (60%) euphausiids (38%) and copepods (23%). In 24–31 cm fish, salps were found in at least 85% of stomachs with food, but chaetognaths and amphipods were still present (Table A2a).

### ***Thyrsites atun barracouta***

Barracouta are caught in shelf areas around New Zealand, the Chatham Islands, and occasionally at the Auckland Islands (Figure B24a). Feeding records are available from most of the main areas except the west coast of the North Island (Figure B24a). A total of 15 542 fish (mainly 30–100 cm fork length) were sampled, of which 48% had stomachs containing food (Figure B24b, Table C22). The proportion of empty stomachs was high (79%) from the North Island area, possibly related to sampling during the spawning season.

Crustaceans were the most important prey, comprising 77% overall (Table C22). The main crustaceans identified were euphausiids (74%), and *Munida* spp. (4%). Teleosts comprised 18% overall, with hoki the most commonly identified (4%), mainly from the WCSI, and some myctophids (1%) and sprats (0.6%). Cephalopods, particularly squid, were also important (9%). In total, at least eight main invertebrate groups in four phyla and 24 teleost (including two mesopelagic) species were identified (Table C22).

There were no major differences in diet between smaller (up to 60 cm) and larger fish (Figure B24c). However, in northern and western areas (North Island and Challenger), teleosts were more important in the diet (about 50%) than off the east and southern coasts of the South Island and at the Chatham Islands (about 10%) (Table 22). Euphausiids (about 40–60%) and cephalopods (1%) were correspondingly less important in the north and west and more important in the other areas (about 70–90% euphausiids and 6–15% squid). *Munida* spp. was locally important (up to 9%) off the east and southern coasts of the South Island (Figure B24c, Table 22).

These findings are consistent with other records (Thomson 1892, Thomson & Anderton 1921, Phillips 1926, Graham 1938, 1953, Mehl 1969, Hurst 1980, Russell 1983, O’Driscoll 1998) from around New Zealand (i.e., euphausiids and fish were important in all studies; *Munida gregaria*, hoki and sprats are locally important) (Table A2a). Other published work did not fill in the gap in knowledge from the west coast of the North Island. These studies, combined with the data on the research trawl database, could be used to examine seasonal trends in feeding of barracouta.

### ***Trachurus murphyi* Murphy’s mackerel**

Murphy’s mackerel have been recorded in research trawls from shelf areas around New Zealand and on the Chatham Rise, mainly down to 500 m depth (Figure B25a). This species is known to occur off the north and west coasts of the North Island, but has not always been identified beyond genus level in research catches. Feeding records are mainly from Southland, with a few from the Chatham Rise (Figure B25a). A total of 414 fish (mainly 40–60 cm fork length) were sampled, of which 48% had stomachs containing food (Figure 25b, Table C23).

Crustaceans (55%) were the main prey group, mainly euphausiids (38%), amphipods (12%) and *Munida* spp. (6%) (Table C23). Salps comprised 36% and teleosts 11%. Crustaceans and teleosts were important in both areas while cephalopods were more important on the Chatham Rise and salps were only recorded in Southern N.Z. (Figure B25c). A total of at least seven main invertebrate groups in three phyla and one teleost group (myctophid) were recorded (Table C23).

There are no published records of feeding for *T. murphyi* from New Zealand. A summary of feeding from catch sampling of purse seine catches off the north-east of the North Island found euphausiids in 98% of stomachs (Paul Taylor, NIWA, Wellington, pers. comm.). A study by Hurst (1980) of two related species, *T. novaezelandiae* and *T. declivis* from the Bay of Plenty, Northland and the WCSI; found euphausiids to be the dominant prey with few amphipods and fish (Tables A1a, A2a). Godfriaux (1968, 1970) also recorded crustaceans (in at least nine groups) as the dominant prey of *T. novaezelandiae* from the Hauraki Gulf, as well as some fish and polychaetes (Table A1a). Studies of *T. murphyi* off Peru (Konchina 1978, 1979, 1980) found that crustaceans dominated the diet, mainly euphausiids, but including copepods, ostracods and decapods (Table A2b). Other prey groups of less importance included fish and molluscs. Salps were not recorded.

### 3.2 Overall Summary

Appendix A comprises a literature review of feeding studies up to and including 2000. However, a number of important dietary studies have been conducted since 2000, in particular a comprehensive trophic study of the Chatham Rise (Dunn 2009, Dunn et al. 2010 a, b, c, Horn et al. 2010, Stevens and Dunn 2011). Although these studies were beyond the scope of this report, it is acknowledged that our understanding of the diet and trophic relationships of some of these species has improved.

Appendices B and C presents feeding data collected opportunistically at sea by several researchers with varying levels of experience in prey identification over several research trawl surveys up to and including 2000. Although these data were groomed and obvious errors corrected or removed, there are likely to be unresolved issues with the identification of some prey, in particular small and/or delicate species. However such errors are unlikely to affect the overall diet of these species. The level of potential error or bias was not studied here.

A summary of key results for the 25 key species is provided in Figure 1.

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## 5. REFERENCES

- Adams, N.M. (1994). Seaweeds of New Zealand. Canterbury University Press, Christchurch. 360 p.
- Anderson, O.F.; Bagley, N.W.; Hurst, R.J.; Francis, M.P.; Clark, M.R.; McMillan, P.J. (1998). Atlas of New Zealand fish and squid distributions from research bottom trawls. *NIWA Technical Report 42*. 303 p.
- Ayling, T.; Cox, G.J. (1987). Collins Guide to the sea fishes of New Zealand. Collins, Auckland. 343 p.
- Baba, O.; Taniuchi, T.; Nose, Y. (1987). Depth distribution and food habits of three species of sharks off Choshi. *Nippon Suisan Gakkaishi* 53: 417–424.
- Bagley, N.W.; Anderson, O.F.; Hurst, R.J.; Francis, M.P.; Taylor, P.R.; Clark, M.R.; Paul, L.J. (2000). Atlas of New Zealand fish and squid distributions from midwater trawls, tuna longline sets and aerial sightings. *NIWA Technical Report 72*. 171 p.
- Baker, A.B. (1971). Food and feeding of kahawai (Teleostei: Arripididae). *New Zealand Journal of Marine and Freshwater Research* 5: 291–299.

- Barnes, R.D. (1987). *Invertebrate Zoology*. CBS College Publishing, USA. 893 p.
- Bass, A.J.; D'Aubrey, J.D.; Kistnasamy, N. (1975). Sharks of the east coast of southern Africa 5. The families Hexanchidae, Clamydoselachidae, Heterodontidae, Pristophoridae and Squantidae. *Invest. Res. Inst., Durban* 43: 50 p.
- Bass, A.J.; D'Aubrey, J.D.; Kistnasamy, N. (1976). Sharks of the east coast of southern Africa 6. The families Oxynotidae, Squalidae, Dalatiidae, and Echinorhinidae. *Invest. Rep Oceanogr. Res. Inst., Durban* 45: 103 p.
- Beentjes, M.P.; Bull, B.; Hurst, R.J.; Bagley, N.W. (2002). Demersal fish assemblages along the continental shelf and upper slope of the east coast of the South Island, New Zealand. *New Zealand Journal of Marine and Freshwater Research* 36: 197–223.
- Bjelland, O.; Bergstad, O.A. (1998). Trophic ecology of deepwater fishes associated with the continental slope of the eastern Norwegian Sea. *ICES 1998/O:51*. 26 p.
- Blaber, S.J.M.; Bulman, C.M. (1987). Diets of fishes of the upper continental slope of eastern Tasmania: content, calorific values, dietary overlap and trophic relationships. *Marine Biology* 95: 345–356.
- Bull, B.; Livingston, M.E.; Hurst, R.; Bagley, N. (2001). Upper-slope fish communities on the Chatham Rise, New Zealand, 1992-99. *New Zealand Journal of Marine and Freshwater Research* 35: 795–815.
- Bulman, C.M.; Blaber, S.J.M. (1986). Feeding ecology of *Macruronus novaezelandiae* (Hector) (Teleostei: Merlucciidae) in south-east Australia. *Australian Journal of Marine and Freshwater Research* 37: 621–639.
- Bulman, C.M.; Koslow, J.A. (1992). Diet and food consumption of a deep-sea fish, orange roughy *Hoplostethus atlanticus* (Pisces: Trachichthyidae), off southeastern Australia. *Marine Ecology progress series* 82: 115–129.
- Capapé, C. (1975). Observations sur le régime alimentaire de 29 Selaciens pleurotêrmes des côtes tunisiennes. *Archives de l'Institut Pasteur de Tunis* 52: 395–414.
- Carrassón, M.; Mattallanas, J.; Casadevall, M. (1997). Feeding strategies of deep-water morids on the western Mediterranean slope. *Deep-Sea Research I* 44: 1685–1699.
- Carrassón, M.; Stefanescu, C.; Cartes, J.E. (1992). Diets and bathymetric distributions of the Catalan deep sea (western Mediterranean). *Marine Ecology Progress Series* 82: 21–30.
- Castillo, J.L.; Uribe, J.A.; Bonfil, R.; Vélez, R.; Mendizabal, D.; de Anda, D.; Mena, R.; Márquez, J.F. (1992). Estado actual de la pesquería de tiburón en México. Instituto Nacional de la Pesca, México. 44 p.
- Choat, J.H.; Clements, K.D. (1992). Diet in Odacid and Aplodactylid fishes from Australia and New Zealand. *Australian Journal of Marine & Freshwater Research* 43: 1451–1459.
- Clark, F.E. (1899). Notes on parore (mangrove fish). *Transactions and Proceedings of the New Zealand Institute* 31: 96–97.
- Clark, M.R. (1985). The food and feeding of seven fish species from the Campbell Plateau, New Zealand. *N.Z. Journal of Marine and Freshwater Research* 19: 339–363.
- Clark, M.R.; King, K.J. (1989). Deepwater fish resources off the North Island, New Zealand: results of a trawl survey, May 1985 to June 1986. *New Zealand Fisheries Technical Report No. 11*.
- Clark, M.R.; King, K.J.; McMillan, P.J. (1989). The food and feeding relationships of black oreo, *Allocyttus niger*, smooth oreo, *Pseudocyttus maculatus*, and eight other fish species from the continental slope of the South-west Chatham Rise, New Zealand. *Journal of Fish Biology* 35: 465–484.
- Clarke, M.R.; Merrett, N. (1972). The significance of squid, whale and other remains from the stomachs of bottom-living deep-sea fish. *Journal of the Marine Biological Association of the United Kingdom* 52: 599–603.
- Clarke, T.A. (1980). Diets of fourteen species of vertically migrating mesopelagic fishes in Hawaiian waters. *Fishery Bulletin* 78: 619–640.
- Clarke, T.A. (1982). Feeding habits of stomiatoid fishes from Hawaiian waters. *Fishery Bulletin* 80: 287–304.

- Clarke, T.A. (1984). Diet and morphological variation in snipefishes, presently recognised as *Macrorhamphosus scolopax*, from southeast Australia: Evidence for two sexually dimorphic species. *Copeia* 1984: 595–608.
- Clements, K.D.; Choat, J.H. (1997). Comparison of herbivory in the closely-related marine fish genera *Girella* and *Kyphosus*. *Marine Biology* 127: 579–586.
- Colman, J.A. (1972). Food of snapper, *Chrysophrys auratus* (Forster), in the Hauraki Gulf, New Zealand. *N.Z. Journal of Marine and Freshwater Research* 6: 221–239.
- Cortés, E. (1999). Standardised diet compositions and trophic levels of sharks. *ICES Journal of Marine Science* 56: 707–717.
- Davenport, J. (1993). Ventilation of the gills by the pectoral fins in the fangtooth *Anoplogaster cornutum*: how to breathe with a full mouth. *Journal of Fish Biology* 42: 967–970.
- Doak, W. (1984). Fishes of the New Zealand region. Hodder & Stroughton, Auckland. 135 p.
- Doak, W. (1991). Wade Doak's World of New Zealand Fishes. Hodder & Stroughton, Auckland. 222 p.
- Dubochkin, A.S.; Kotlyar, A.N. (1989). On the feeding of alfoncino (*Beryx splendens*). *Journal of Ichthyology* 29: 1–8.
- Dunn, M.R. (2009). Feeding habits of the ommastrephid squid *Nototodarus sloanii* on the Chatham Rise, New Zealand. *New Zealand Journal of Marine and Freshwater Research* 43: 1103–1113.
- Dunn, M.R.; Connell, A.M.; Forman, J.; Stevens, D.W.; Horn, P.L. (2010a). Diet of Two Large Sympatric Teleosts, the Ling (*Genypterus blacodes*) and Hake (*Merluccius australis*). *PLoS ONE* 5(10): e13647.
- Dunn, M.R.; Griggs, L.; Forman, J.; Horn, P. (2010b). Feeding habits and niche separation among the deep-sea chimaeroid fishes *Harriotta raleighana*, *Hydrolagus bemisi* and *Hydrolagus novaezealandiae*. *Marine Ecology Progress Series* 407: 209–225.
- Dunn, M.R.; Szabo, A.; McVeagh, M.S.; Smith, P.J. (2010c). The diet of deepwater sharks and the benefits of using DNA identification of prey. *Deep Sea Research Part I: Oceanographic Research Papers* 57: 923–930.
- Du Buit, M. (1978). The diet of some sea teleost fishes in the Wyville Thomson ridge area. *Oceanologica acta* 1: 129–134.
- Ebert, D.A. (1989). Life history of the sevengill shark, *Notorynchus cepedianus* Peron, in two northern California bays. *California Fish and Game* 75: 102–112.
- Ebert, D.A. (1991). Diet of the sevengill shark *Notorynchus cepedianus* in the temperate coastal waters of southern Africa. *South African Journal of Marine Science* 11: 565–572.
- Ebert, D.A.; Compagno, L.J.V.; Cowley, P.D. (1992). Benguela Trophic Functioning: A preliminary investigation of the feeding ecology of squaloid sharks off the west coast of southern Africa. *South African Journal of Marine Science* 12: 601–609.
- Edgar, G.J. (1990). The use of the size-structure of benthic macrofaunal communities to estimate faunal biomass and secondary production. *Journal of Experimental Marine Biology and Ecology* 137: 195–214.
- Edgar, G.J.; Shaw, C. (1995). The production and trophic ecology of shallow-water fish assemblages in southern Australia. II. Diets of fishes and trophic relationships between fishes and benthos at Western Port, Victoria. *Journal of Experimental Marine Biology and Ecology* 194: 83–106.
- Ellis, J.R.; Pawson, M.G.; Shackley, S.E. (1996). The comparative feeding ecology of six species of shark and four species of ray (Elasmobranchii) in the north-east Atlantic. *Journal of the Marine Biological Association U.K.* 76: 89–106.
- Fenwick, G.D. (1978). Plankton swarms and their predators at the Snares Islands. *New Zealand Journal of Marine and Freshwater Research* 12: 223–224.
- Francis, M.P. (1996). Coastal fishes of New Zealand. An Identification guide. Reed, Auckland. 72 p.
- Francis, M.P.; Hurst, R.J.; McArdle, B.; Bagley, N.W.; Anderson, O.F. (2002). New Zealand demersal fish assemblages. *Environmental Biology of Fishes* 65: 215–234.
- Galván-Magana, F.; Nienhuis, H.J.; Klimley, A.P. (1989). Seasonal abundance and feeding habits of sharks of the lower Gulf of California, Mexico. *Californian Fish and Game* 75: 74–84.
- Gartner, J.V.; Musick, J.A. (1989). Feeding habits of the deep-sea fish, *Scopelogadus beanii* (Pisces: Melamphaidae), in the western North Atlantic. *Deep-Sea Research* 36: 1457–1469.

- Gavrilov, G.M.; Markina, N.P. (1979). The feeding ecology of fishes of the genus *Seriolella* (fam. Nomeidae) on the New Zealand Plateau. *Journal of Ichthyology* 19: 128–135.
- Glasby, J.C.; Read, G.B. (1998). A chronological review of polychaete taxonomy in New Zealand. *Journal of the Royal Society of New Zealand* 28: 347–374.
- Godfriaux, B.L. (1968). The food and feeding relationships of snapper (*Chrysophrys auratus*) and some other fish-species trawled in Hauraki Gulf. Unpublished MSc (Hons) thesis, Victoria University of Wellington.
- Godfriaux, B.L. (1969). Food of predatory demersal fish in Hauraki Gulf. 1. Food and feeding habits of snapper. *N.Z. Journal of Marine and Freshwater Research* 3: 518–544.
- Godfriaux, B.L. (1970). Food of predatory demersal fish in Hauraki Gulf. II. Five fish species associated with snapper. *N.Z. Journal of Marine and Freshwater Research* 4: 248–266.
- Godfriaux, B.L. (1974a). Food of tarakihi in western Bay of Plenty and Tasman Bay, New Zealand. *N.Z. Journal of Marine and Freshwater Research* 8: 111–153.
- Godfriaux, B.L. (1974b). Food of snapper in western Bay of Plenty, New Zealand. *N.Z. Journal of Marine and Freshwater Research* 8: 473–504.
- Gomes, T.M.; Sola, E.; Grós, M.P.; Menezes, G.; Pinho, M.R. (1998). Trophic relationships and feeding habits of demersal fishes from the Azores: Importance to multispecies assessment. *ICES 1998/O:7*, 35 p.
- Gorelova, T.A. (1981). Notes on feeding and gonad condition in three species of the genus *Gonostoma* (Gonostomatidae). *Journal of Ichthyology* 21: 82–92.
- Gorelova, T.A.; Agafonova, T.B.; Lipskaya, N.J. (1994). Feeding of cigarfishes (Genus *Cubiceps*, Stromateoidei). *Journal of Ichthyology* 34: 70–82.
- Gorelova, T.A.; Krasil'nikova, N.A. (1990). On the diets of *Maurolicus muelleri* in the vicinity of seamounts Discovery, Nasca, and Mt. Africana. *Journal of Ichthyology* 30: 42–52.
- Graham, D.H. (1938). Food of the fishes of Otago Harbour and adjacent sea. *Transactions of the Royal Society of New Zealand* 68: 421–436.
- Graham, D.H. (1953). A treasury of New Zealand fishes. Reed, Wellington. 404 p.
- Habib, G. (1975). Aspects of the biology of the red cod *Pseudophycis bacchus*. Unpublished Ph.D. thesis, Department of Zoology, University of Canterbury, New Zealand. 203 p.
- Hanchet, S. (1991). Diet of spiny dogfish, *Squalus acanthias* Linnaeus, on the east coast, South Island, New Zealand. *Journal of Fish Biology* 39: 313–323.
- Heath, E.; Moreland, J.M. (1967). Marine Fishes of New Zealand. Reed, Wellington. 56 p.
- Hopkins, T.L.; Baird, R.C. (1985). Aspects of the trophic ecology of the mesopelagic fish *Lampanyctus alatus* (Family Myctophidae) in the eastern Gulf of Mexico. *Biological Oceanography* 3: 285–313.
- Horn, P.L.; Forman, J.; Dunn, M.R. (2010). Feeding habits of alfonsino *Beryx splendens*. *Journal of Fish Biology* 76: 2382–2400.
- Hurst, R.J. (1980). Studies on the life cycle of some New Zealand Anisakidae (Nematoda). Unpublished PhD thesis, Victoria University of Wellington, New Zealand. 212 p.
- Hurst, R.J.; Bagley, N.W.; Anderson, O.F.; Francis, M.P.; Griggs, L.H.; Clark, M.R.; Taylor, P.R. (2000). Atlas of juvenile fish and squid distributions from bottom and midwater trawls and tuna longline sets. *NIWA Technical Report 84*. 162 p.
- Ingerson, J.K.V. (1996). Food and feeding of red gurnard (Triglidae : *Chelidonichthys kumu*) from the West Coast, Golden and Tasman Bays, New Zealand. Unpublished MSc (Hons) thesis, Victoria University of Wellington.
- James, G.D. (1972). Food of a surface school of trevally (Teleostei: Carangidae) in the North-West Bay of Plenty. Indo-Pacific Fisheries Council Proceedings, 15<sup>th</sup> Session, Wellington, New Zealand. 388–389.
- Johnston, A.D. (1983). The southern Cook Strait groper fishery. *Fisheries Technical Report No. 159*. The Ministry of Agriculture and Fisheries, Wellington, New Zealand. 33 p.
- Kerstan, S.L. (1989). The food of silver roughy (*Hoplostethus mediterraneus*, Beryciformes, Trachichthyidae) from New Zealand waters. *Meeresforsch* 32: 241–247.



- King, K.J.; Clark, M.R. (1984). The food of rig (*Mustelus lenticulatus*) and the relationship of feeding to reproduction and condition in Golden Bay. *New Zealand Journal of Marine & Freshwater Research* 18: 29–42.
- Konchina, Y.V. (1978). Some data on the feeding of the Peruvian Jack mackerel, *Trachurus symmetricus murphyi*. *Journal of Ichthyology* 18: 687–690.
- Konchina, Y.V. (1979). The feeding of the Peruvian Jack mackerel, *Trachurus symmetricus murphyi*. *Journal of Ichthyology* 19: 52–61.
- Konchina, Y.V. (1980). The Peruvian Jack mackerel, *Trachurus symmetricus murphyi*, a facultative predator in the coastal upwelling ecosystem. *Journal of Ichthyology* 20: 46–59.
- Kubota, T.; Shiobara, Y.; Kubodera, T. (1991). Food habits of frilled shark *Chlamydoselachus anguineus* collected from Suruga Bay, Central Japan. *Bulletin of the Japanese Society of Scientific Fisheries* 57: 15–20.
- Kubota, T. (1977). Food of lancetfish, *Alepisaurus ferox* Lowe, fished by gill-net in Suruga Bay, Japan. *Journal of the faculty of Marine Science and Technology, Tokai University* 10: 137–146.
- Kuo, C.; Tanaka, S. (1984). Feeding habit of hoki *Macruronus novaezelandiae* (Hector) in waters around New Zealand. *Bulletin of the Japanese Society of Scientific Fisheries* 50: 783–786.
- Lancraft, T.M.; Hopkins, T.L.; Torres, J.J. (1988). Aspects of the ecology of the mesopelagic fish *Gonostoma elongatum* (Gonostomatidae, Stomiiformes) in the eastern Gulf of Mexico. *Marine Ecology Progress Series* 49: 27–40.
- Lenanton, R.C.J.; Robertson, A.I.; Hansen, J.A. (1982). Nearshore accumulations of detached macrophytes as nursery areas for fish. *Marine Ecology Progress Series* 9: 51–57.
- Lipinsky, M.R.; Payne, A.I.L.; Rose, B. (1992). The importance of cephalopods as prey for hake and other groundfish in South African waters. *South African Journal of marine Science* 12: 651–662.
- Litvinov, F.F. (1990). Ecological characteristics of the dogfish, *Squalus mitsukurii*, from the Sala-y-Gomez Seamounts. *Journal of Ichthyology* 30: 104–115.
- Livingston, M.E. (1987). Food resource use among five flatfish species (Pleuronectiformes) in Wellington Harbour. *New Zealand Journal of Marine and Freshwater Research* 21: 281–293.
- Lyle, J.M.; Smith, D.C. (1997). Abundance and biology of warty oreo (*Allocyttus verrucosus*) and spiky oreo (*Neocyttus rhomboidalis*) (Oreosomatidae) off south-eastern Australia. *Marine and Freshwater Research* 48: 91–102.
- Macpherson, E. (1980). Régime alimentaire de *Galeus melastomus* Rafinesque, 1810, *Etmopterus spinax* (L., 1758) et *Scymnorhinus licha* (Bonnaterre, 1788) en Méditerranée occidentale. *Vie Milieu* 30: 139–148.
- Macpherson, E. (1981). Resource partitioning in a Mediterranean demersal fish community. *Marine Ecology Progress Series* 4: 183–193.
- Macpherson, E. (1985). Daily ration and feeding periodicity of some fishes off the coast of Namibia. *Marine Ecology Progress Series* 26: 253–260.
- Marshall, N.B.; Merrett, N.R. (1977). The existence of a benthopelagic fauna in the deep-sea. In: A voyage of discovery. Edited by M. Angel. (Suppl.), *Deep-Sea Research*. George Deacon 70th anniversary volume.
- Marwick, G. (1942). A study of the tarakihi (*Nemadactylus macropterus*): economic importance, taxonomy, growth rate, size and age distribution in the trawl catches landed at Wellington. Unpublished thesis held in the NIWA Greta Point library.
- Matallanas, J. (1982). Feeding habits of *Scymnorhinus licha* in Catalan waters. *Journal of Fish Biology* 20: 155–163.
- Mauchline, J.; Gordon, J.D.M. (1983a). Diets of the sharks and chimaeroids of the Rockall Trough, northeastern Atlantic Ocean. *Marine Biology* 75: 269–278.
- Mauchline, J.; Gordon, J.D.M. (1983b). Diets of clupeoid, stomiatoid and salmonoid fish of the Rockall Trough, northeastern Atlantic Ocean. *Marine Biology* 77: 67–78.
- Mauchline, J.; Gordon, J.D.M. (1984a). Occurrence and feeding of berycomorphid and percomorphid teleost fish in the Rockall Trough. *J. Cons. Int. Explor. Mer.* 41: 239–247.
- Mauchline, J.; Gordon, J.D.M. (1984b). Diets and bathymetric distributions of the macrourid fish of the Rockall Trough, northeastern Atlantic Ocean. *Marine Biology* 81: 107–121.

- Mauchline, J.; Gordon, J.D.M. (1984c). Feeding and bathymetric distribution of the gadoid and morid fish of the Rockall Trough. *Journal of the Marine Biological Association of the U.K.* 64: 657–665.
- McClay, C.L. (1988). Brachyura and crab-like anomura of New Zealand. *Leigh Laboratory Bulletin* 22. 463 p.
- McKee, M. (1981). Blackfish *Centrolophus niger* (Gmelin) taken from the Irish Sea. *Irish Naturalists' Journal* 20: 258.
- McLellan, T. (1977). Feeding strategies of macrourids. *Deep-Sea Research* 24: 1019–1036.
- Mehl, J.A.P. (1969). Food of the barracouta (Teleostei: Gempylidae) from eastern Cook Strait. *N.Z. Journal of Marine and Freshwater Research* 3: 389–394.
- Mel'nikov, Y.S. (1981). Feeding peculiarities of *Allocyttus verrucosus*, Family Oreosomatidae. *Journal of Ichthyology* 20: 99–105.
- Menni, R.C.; Cousseau, M.B.; Gosztonyi, A.R. (1982). Sobre la biología de los tiburones costeros de la provincia de Buenos Aires. *An. Soc. Cient. Argentina, vol. ccxiii*.
- Menni, R.C.; Lopez, H.L. (1979). Biological data and otolith (sagitta) morphology of *Polyprion americanus* and *Schedophilus griseolineatus* (Osteichthyes, Serranidae and Centrolophidae). *Studies on Neotropical Fauna and Environment* 14: 17–32.
- Meyer, M.; Smale, M.J. (1991a). Predation patterns of demersal teleosts from the cape south and west coasts of South Africa. 1. Pelagic predators. *South African Journal of Marine Science* 10: 173–191.
- Meyer, M.; Smale, M.J. (1991b). Predation patterns of demersal teleosts from the cape south and west coasts of South Africa. 2. Benthic and epibenthic predators. *South African Journal of Marine Science* 11: 409–442.
- Mitchell, S.J. (1984). Feeding of ling *Genypterus blacodes* (Bloch and Schneider) from 4 New Zealand offshore fishing grounds. *N.Z. Journal of Marine and Freshwater Research* 18: 265–274.
- Minchin, T. (1988). A record of the deep-sea anglerfish, *Cryptopsaras couesi* Gill, from the north-eastern Atlantic. *Journal of Fish Biology* 32: 313.
- Moreland, J.M. (1963). Native Sea Fishes. Reed, Wellington. 64 p.
- Moreland, J.M.; Heath, E. (1983). Marine Fishes I. Reed, Wellington. 79 p.
- Morton, J.; Miller, M. (1968). The New Zealand Sea Shore. Collins, Auckland. 638 p.
- Moteki, M.; Fujita, K.; Kohno, H. (1992). Stomach contents of longnose lancetfish, *Alepisaurus ferrox*, in Hawaiian and central equatorial Pacific waters. *Journal of Tokyo University of Fisheries* 80: 121–137.
- Mundy, A.R. (1968). A study of the biology of the sand flounder *Rhombosolea plebeia* (Richardson) off the Canterbury Coast. Unpublished PhD thesis, University of Canterbury.
- Mutch, P.G. (1983). Factors affecting the density and distribution of the blue cod (*Parapercis colias*) (Pisces: Mugiloididae). Unpublished MSc thesis, University of Auckland.
- O'Driscoll, R.L. (1998). Feeding and schooling behaviour of barracouta (*Thyrssites atun*) off Otago, New Zealand. *Marine and Freshwater Research* 49: 19–24.
- Parin, N.V.; Pavlov, Y.P.; Andrianov, D.P. (1988). Ecological characteristics of mirror dory *Zenopsis nebulosus* from the Nazca Ridge. *Journal of Ichthyology* 28: 707–716.
- Parrott, A.W. (1957). Sea anglers fishes of New Zealand. Hodder and Stoughton, London. 176 p.
- Parrott, A.W. (1960). The queer and the rare fishes of New Zealand. Hodder and Stoughton, London. 192 p.
- Pascoe, P.I. (1986). Fish otoliths from the stomach of a thresher shark, *Alopias vulpinus*. *Journal of the Marine Biological Association. U.K.* 66: 315–317.
- Paul, L.J.; Heath, E. (1985). Marine Fishes II. Reed, Wellington. 80 p.
- Paul, L.J. (2000). New Zealand Fishes: identification, natural history & fisheries. Reed, Auckland. 253 p.
- Paulin, C.; Stewart, A.; Roberts, C.; McMillan, P. (1989). New Zealand fish. A complete guide. *National Museum of New Zealand Miscellaneous Series* 19. 279 p.
- Paya, I. (1992). The diet of Patagonian hake *Merluccius australis polylepis* and its daily ration of Patagonian grenadier *Macrouronus magellanicus*. *South African Journal of Marine Science* 12: 753–760.

- Pearks, D.S.J. (1985). *Rhombosolea leporina* of the Manukau Harbour. Unpublished MSc thesis, University of Auckland.
- Phillipps, W.J. (1926). Food of New Zealand fishes. *Transactions and Proceedings of the N.Z. Institute* 56: 525–529.
- Platell, M.E.; Potter, I.C.; Clarke, K.R. (1998). Do the habits, mouth morphology and diets of the mullids *Upeneichthys stotti* and *U. lineatus* in coastal waters of south-western Australia differ? *Journal of Fish Biology* 52: 398–418.
- Powell, A.W.B. (1937). Animal communities of the sea-bottom in Auckland and Manukau Harbours. *Transactions and Proceedings of the Royal Society of New Zealand* 66: 354–401.
- Powell, A.W.B. (1976). Shells of New Zealand. Whitcoulls, Christchurch. 154 p.
- Rapson, A.M. (1940). The reproduction, growth, and distribution of the lemon soles (*Pelotretis flavilatus*) of Tasman Bay and Marlborough Sounds. *New Zealand Marine Department Fisheries Bulletin* 7.
- Rapson, A.M. (1956). Biology of the blue cod (*Parapercis colias* Forster) of New Zealand. Unpublished PhD thesis, Victoria University of Wellington.
- Rojas, Z.P.; Flores, G.H.; Sepulveda, J.I. (1985). Feeding of Juan Fernandez codfish *Polyprion oxygeneios* (Bloch & Schneider, 1801) (Pisces: Percichthyidae). *Marine Investigations in the Juan Fernandez Archipelago 1985*: 305–309.
- Roper, C.F.E.; Sweeney, M.J.; Nauen, C. (1984). FAO species catalogue. Vol. 3. Cephalopods of the world. An annotated and illustrated catalogue of species of interest to fisheries. *FAO Fisheries Synopsis (125)* Vol. 3. 277 p.
- Rosecchi, E.; Tracey, D.M.; Webber, W.R. (1988). Diet of orange roughy, *Hoplostethus atlanticus* (Pisces: Trachichthyidae) on the Challenger Plateau, New Zealand. *Marine Biology* 99: 293–306.
- Roshchin, E.A. (1986). Some biological properties of redbait, *Emmelichthys nitus* (Emmelichthyidae), from a seamount in the notal zone of the Indian ocean. *Journal of Ichthyology* 25: 44–50.
- Russell, B.C. (1983). The food and feeding habits of rocky reef fish of north-eastern New Zealand. *New Zealand Journal of Marine and Freshwater Research* 17: 121–145.
- Sauer, W.H.H.; Smale, M.J. (1991). Predation patterns on the inshore spawning grounds of the squid *Loligo vulgaris reynaudii* (Cephalopoda: Lologinidae) off the south-eastern Cape, South Africa. *South African Journal of Marine Science* 11: 513–523.
- Sedberry, G.R.; Musick, J.A. (1978). Feeding strategies of some demersal fishes of the continental slope and rise off the mid-Atlantic coast of the USA. *Marine Biology* 44: 357–375.
- Sherrin, R.A.A. (1886). Handbook of the fishes of New Zealand. Wilsons and Horton, Auckland. 307 p.
- Shpak, V.M. (1976). Feeding behaviour of the southern blue whiting *Micromesistius australis* Norman, 1937 in the south-west Pacific. *Research in Fish Biology and Fisheries Oceanography* 7: 37–45. [In Russian, Translation No. 198 held in NIWA Greta Point library, Wellington.]
- Smale, M.J. (1991). Occurrence and feeding of three shark species, *Carcharhinus brachyurus*, *C. obscurus* and *Sphyrna zygaena*, on the eastern cape coast of South Africa. *South African Journal of Marine Science* 11: 31–42.
- Stergiou, K.I.; Fourtouni, H. (1991). Food habits, ontogenetic diet shift and selectivity in *Zeus faber* Linnaeus, 1758. *Journal of Fish Biology* 39: 589–603.
- Stevens, D.W.; Dunn, M.R. (2011). Different food preferences in four sympatric deep-sea Macrourid fishes. *Marine Biology* 158: 59–72.
- Stevens, J.D.; Hausfeld, H.F.; Davenport, S.R. (1984). Observations on the biology, distribution and abundance of *Trachurus declivis*, *Sardinops neopilchardus* and *Scomber australasicus* in the Great Australian Bight. *CSIRO Marine Laboratories Report No.164*. 27 p.
- Strasburg, D.W. (1958). Distribution, abundance and habits of pelagic sharks in the central Pacific Ocean. *Fisheries Bulletin of the Fisheries Wildlife Service U.S.* 58: 335–361.
- Sulak, K.J.; Wenner, C.A.; Sedberry, G.R.; Van Guelpen, L. (1985). The life history and systematics of deep-sea lizard fishes, genus *Bathysaurus* (Synodontidae). *Canadian Journal of Zoology* 63: 623–642.
- Suter, H. (1913). Manual of the New Zealand Mollusca. John Mackay, Government Printer, Wellington. 1120 p.

- Suter, H. (1915). Manual of the New Zealand Mollusca: Atlas of Plates. John Mackay, Government Printer, Wellington.
- Taniuchi, T. (1988). Aspects of reproduction and food habits of the Japanese swellshark *Cephaloscyllium umbratile*. *Nippon Suisan Gakkaishi* 54: 627–633.
- Thomson, G.M. (1891). Notes on Sea-Fishes. *Transactions and Proceedings of the N.Z. Institute* 24: 202–215.
- Thomson, G.M.; Anderton, T. (1921). History of the Portobello Marine Fish-Hatchery and Biological station. *Board of Science and Art Bulletin No. 2*. 131 p.
- Thomson, J.M. (1954). The organs of feeding and the food of some Australian mullet. *Australian Journal of Marine and Freshwater Research* 5: 469–485.
- Thomson, J.M. (1957). Biological studies of economic significance of the yellow-eye mullet, *Aldrichetta forsteri* (Cuvier & Valenciennes) (Mugilidae). *Australian Journal of Marine and Freshwater Research* 8: 1–13.
- Vinnichenko, V.I. (1997). Russian investigations and deep water fishery on the Corner Rising Seamount in subarea 6. *NAFO Sci. Coun. Studies* 30: 41–49.
- Wear, R.G.; Fielder, D.R. (1985). The Marine Fauna of New Zealand: Larvae of the Brachyura (Crustacea, Decapoda). *New Zealand Oceanographic Institute Memoir* 92. 90 p.
- Webb, B.F. (1973). Fish populations of the Avon-Heathcote estuary 3. Gut contents. *New Zealand Journal of Marine and Freshwater Research* 7: 223–234.
- Wilson, C.D.; Seki, M.P. (1994). Biology and population characteristics of *Squalus mitsukurii* from a seamount in the central North Pacific Ocean. *Fishery Bulletin* 92: 851–864.
- Winstanley, R.H. (1978). Food of the trevalla *Hyperoglyphe porosa* (Richardson) off south-eastern Australia. *New Zealand Journal of Marine and Freshwater Research* 12: 77–79.
- Yano, K. (1991). Catch distribution, stomach contents, and size at maturity of two squaloid sharks, *Deania calceus* and *D. crepidalbus*, from the southeast Atlantic off Namibia. *Bulletin of the Japanese Society of Fisheries and Oceanography*. 55: 189–196.
- Yano, K. (1993). Distribution and food habits of the slender smoothhound, *Gollum attenuatus*, from waters around New Zealand. *Japanese Journal of Ichthyology* 39: 345–356.
- Yano, K.; Tanaka, S. (1984). Some biological aspects of the deep sea squaloid shark *Centroscymnus* from Suruga Bay, Japan. *Bulletin of the Japanese Society of Scientific Fisheries* 50: 249–256.
- Young, J.W.; Blaber, S.J.M. (1986). Feeding ecology of three species of midwater fishes associated with the continental slope of eastern Tasmania, Australia. *Marine Biology* 93: 147–156.

**Table 1: Alphabetical list of species included in the literature review, by depth range.**

<b>Inshore (&lt;100 m)</b>	<b>Shelf (100–400 m)</b>	<b>Upper slope (400-700 m)</b>	<b>Mid slope (&gt; 700 m)</b>
<i>Aldrichetta forsteri</i>	<i>Alopias vulpinus</i>	<i>Argentina elongata</i>	<i>Alepocephalus australis</i>
<i>Arripis trutta</i>	<i>Arnoglossus scapha</i>	<i>Beryx decadactylus</i>	<i>Alepisaurus ferox</i>
<i>Auchenoceros punctatus</i>	<i>Caelorinchus biclinozonalis</i>	<i>B. splendens</i>	<i>Alloctytus niger</i>
<i>Callorhynchus milii</i>	<i>Caesioperca lepidoptera</i>	<i>Brama brama</i>	<i>A. verrucosus</i>
<i>Chelidonichthys kumu</i>	<i>Cephaloscyllium isabellum</i>	<i>Caelorinchus aspercephalus</i>	<i>Anoplogaster cornuta</i>
<i>Colistium guntheri</i>	<i>Congiopodus leucopaecilus</i>	<i>C. fasciatus</i>	<i>Antimora rostrata</i>
<i>C. nudipinnis</i>	<i>Cyttus novaezelandiae</i>	<i>C. oliverianus</i>	<i>Aristomias</i> spp.
<i>Conger verreauxi</i>	<i>Emmelichthys nitidus</i>	<i>Centriscoops humerosus</i>	<i>Astronesthes indicus</i>
<i>C. wilsoni</i>	<i>Galeorhinus galeus</i>	<i>Centrolophus niger</i>	<i>A. splendidus</i>
<i>Crapatalus novaezelandiae</i>	<i>Gonorynchus gonorynchus</i>	<i>Cyttus traversi</i>	<i>Bathysaurus ferox</i>
<i>Dasyatis brevicaudatus</i>	<i>Helicolenus percoides</i>	<i>Epigonus denticulatus</i>	<i>Caelorinchus fasciatus</i>
<i>Engraulis australis</i>	<i>Heptranchias perlo</i>	<i>Etmopterus lucifer</i>	<i>Caelorinchus innotabilis</i>
<i>Genyagnus monopterygius</i>	<i>Hemerocoetes</i> sp.	<i>Genypterus blacodes</i>	<i>Centrophorus squamosus</i>
<i>Girella tricuspidata</i>	<i>Kathetostoma giganteum</i>	<i>Gollum attenuatus</i>	<i>Centroscyminus coelolepis</i>
<i>Latridopsis ciliaris</i>	<i>Latris lineata</i>	<i>Helicolenus percoides</i>	<i>C. crepidater</i>
<i>Leptoscopus macropygus</i>	<i>Lepidopus caudatus</i>	<i>Hoplostethus mediterraneus</i>	<i>C. owstoni</i>
<i>Myliobatis tenuicaudatus</i>	<i>Lepidotrigla brachyoptera</i>	<i>Hyperoglyphe antarctica</i>	<i>Chauliodus sloani</i>
<i>Nemadactylus douglasi</i>	<i>Macrorhamphosus scolopax</i>	<i>Lampanyctodes hectoris</i>	<i>Chaunax pictus</i>
<i>Notolabrus celidotus</i>	<i>Mustelus lenticulatus</i>	<i>Lepidorhynchus denticulatus</i>	<i>Chlamydoselachus anguineus</i>
<i>Pagrus auratus</i>	<i>Nemadactylus macropterus</i>	<i>Macruronus novaezelandiae</i>	<i>Coryphaenoides subserrulatus</i>
<i>Paranotothenia angustata</i>	<i>Notolabrus cintus</i>	<i>Malacocephalus laevis</i>	<i>Cryptopsaras couesi</i>
<i>Parika scaber</i>	<i>Notorynchus cepedianus</i>	<i>Maurollicus muelleri</i>	<i>Cubiceps baxteri</i>
<i>Paristiopterus labiosus</i>	<i>Polyprion americanus</i>	<i>Merluccius australis</i>	<i>Dalatias licha</i>
<i>Pelt. novaezeelandiae</i>	<i>P. oxygeneios</i>	<i>Micromesistius australis</i>	<i>Deania calcea</i>
<i>Pseudocaranx dentex</i>	<i>Parapercis colias</i>	<i>Neophrynichthys latus</i>	<i>Diaphus danae</i>
<i>Rhombosolea leporina</i>	<i>Pelotretis flavilatus</i>	<i>Paranotothenia microlepidota</i>	<i>Diastranchius capensis</i>
<i>R. plebeia</i>	<i>Pseudolabrus miles</i>	<i>Seriotelella caerulea</i>	<i>Epigonus lenimen</i>
<i>Sardinops neopilchardus</i>	<i>Pseudophycis bachus</i>		<i>E. telescopus</i>
<i>Scomber australasicus</i>	<i>P. breviscula</i>		<i>Etmopterus baxteri</i>
<i>Seriola lalandi</i>	<i>Raja nasuta</i>		<i>Etmopterus pusillus</i>
<i>Sphyrna zygaena</i>	<i>Rexea solandri</i>		<i>Gonostoma elongatum</i>
<i>Sprattus antipodum</i>	<i>Rhombosolea tapirina</i>		<i>Halargyreus johnsonii</i>
<i>Trachurus novaezelandiae</i>	<i>Seriotelella brama</i>		<i>Halosauropsis macrochir</i>
<i>Upeneichthys lineatus</i>	<i>Seriotelella punctata</i>		<i>Harriotta raleighana</i>
<i>Zeus faber</i>	<i>Squalus acanthias</i>		<i>Hoplostethus atlanticus</i>
	<i>S. mitsukurii</i>		<i>Idiacanthus fasciola</i>
	<i>Thyrsites atun</i>		<i>Lampanyctus alatus</i>
	<i>Trachurus declivis</i>		<i>L. nobilis</i>
	<i>T. murphyi</i>		<i>Macrourus carinatus</i>
	<i>Typhlonarke aysoni</i>		<i>Malacocephalus laevis</i>
	<i>Zenopsis nebulosus</i>		<i>Malacosteus niger</i>
			<i>Mora moro</i>
			<i>Neocyttus rhomboidalis</i>
			<i>Odontomacrus murrayi</i>
			<i>Photostomias</i> , 2 spp.
			<i>Pseudocyttus maculatus</i>
			<i>Raja hyperborea</i>
			<i>Scopelogadus beanii</i>
			<i>Simenchelys parasiticus</i>
			<i>Stomias boa ferox</i>
			<i>Xenodermichthys copei</i>

**Table 2: Feeding records on the fish communities database (Anderson et al (1998), for species with 10 or more fish sampled (\*includes some data additional to that in the Ministry of Fisheries trawl database). Where species are indicated in bold, feeding data have been summarised in this report.**

Species Scientific name	Common name	Number of tows		Number of stomachs <sup>1</sup>			
		Empty	Everted	Regurgitated	With prey	Examined	
<b><i>Alloctylus niger</i></b>	<b>Black oreo</b>	<b>408</b>	<b>4 214</b>		<b>1 061</b>	<b>6 282</b>	
<i>Alloctylus verrucosus</i> *	Warty oreo	6	6		9	16	
<b><i>Beryx splendens</i></b>	<b>Alfonsino</b>	<b>32</b>	<b>4</b>		<b>162</b>	<b>365</b>	
<i>Brama brama</i>	Ray's bream	4	5		43	28	
<i>Caelorhynchus matamua</i> *	Mahia rattail	6	2	1	8	10	
<i>Centroscymnus coelolepis</i> *	Portuguese dogfish	9	7		9	15	
<b><i>Chelidonichthys kumu</i></b>	<b>Red gurnard</b>	<b>89</b>	<b>557</b>	<b>132</b>	<b>331</b>	<b>987</b>	
<i>Coryphaenoides murrayi</i>	Abyssal rattail	6	13	3	2	18	
<b><i>Cyttus triversi</i></b> *	<b>Lookdown dory</b>	<b>53</b>	<b>330</b>	<b>13</b>	<b>224</b>	<b>549</b>	
<i>Deania calcea</i> *	Shovelnose dogfish	6	8		5	13	
<i>Diastrobranchius capensis</i> *	Basketwork eel	16	18		16	29	
<i>Epigonus telescopus</i>	Black cardinalfish	15	60		54	166	
<i>Etmopterus baxteri</i> *	Baxter's dogfish	6	4	63	7	11	
<b><i>Genypterus blacodes</i></b>	<b>Ling</b>	<b>1 535</b>	<b>10 592</b>	<b>183</b>	<b>201</b>	<b>8 148</b>	
<i>Harriota raleighana</i> *	Longnose spookfish	11	8		9	14	
<b><i>Helicolenus percoides</i></b> *	<b>Sea perch</b>	<b>18</b>	<b>166</b>	<b>6</b>	<b>133</b>	<b>296</b>	
<b><i>Hoplostethus atlanticus</i></b> *	<b>Orange roughy</b>	<b>5 856</b>	<b>75 249</b>	<b>246</b>	<b>1</b>	<b>35 496</b>	
<b><i>Hydrolagus novaezealandiae</i></b> *	<b>Dark ghost shark</b>	<b>8</b>	<b>133</b>	<b>2</b>	<b>120</b>	<b>226</b>	
<b><i>Hydrolagus bemisi</i></b> *	<b>Pale ghost shark</b>	<b>24</b>	<b>31</b>	<b>1</b>	<b>105</b>	<b>109</b>	
<b><i>Hyperoglyphe antarctica</i></b>	<b>Blue nose</b>	<b>36</b>	<b>144</b>	<b>18</b>	<b>79</b>	<b>224</b>	
<b><i>Kathetostoma giganteum</i></b>	<b>Giant stargazer</b>	<b>474</b>	<b>733</b>	<b>3</b>	<b>3 535</b>	<b>3 470</b>	
<b><i>Kathetostoma binigrasella</i></b>	<b>Banded stargazer</b>	<b>77</b>	<b>36</b>		<b>530</b>	<b>406</b>	
<i>Lepidopus caudatus</i>	Frostfish	3	19		4	23	
<i>Lepidorhynchus denticulatus</i> *	Javelinfish	4	9	24	10	42	
<b><i>Macruronus novaezelandiae</i></b> *	<b>Hoki</b>	<b>1 846</b>	<b>18 744</b>	<b>827</b>	<b>9</b>	<b>16 310</b>	
<b><i>Merluccius australis</i></b> *	<b>Hake</b>	<b>1 423</b>	<b>4 904</b>	<b>228</b>	<b>16</b>	<b>7 452</b>	
<i>Mesobius antipodum</i>	Black javelinfish	7	30	27	18	73	
<b><i>Micromesistius australis</i></b>	<b>Southern blue whiting</b>	<b>357</b>	<b>2 560</b>	<b>939</b>	<b>2 853</b>	<b>5 857</b>	
<i>Mora moro</i> *	Ribaldo	50	22	174	2	198	

**Table 2 (continued)**

Species Scientific name	Common name	Number of tows	Number of stomachs <sup>1</sup>				
			Empty	Everted	Regurgitated	With prey Examined	
<i>Nemadactylus macropterus</i>	Tarakihi	15	106	2		28	135
<i>Neocyttus rhomboidalis</i> *	Spiky oreo	29	107	265		11	382
<i>Nototodarus sloanii</i>	Sloan's arrow squid	4	66			2	68
<i>Paraperis colias</i>	<b>Blue cod</b>	<b>25</b>	<b>71</b>			<b>66</b>	<b>131</b>
<i>Plagiogeneion rubiginosum</i>	Rubyfish	2	43	2		31	68
<i>Polyprion oxygeneios</i>	<b>Hapuku</b>	<b>85</b>	<b>150</b>	<b>20</b>		<b>182</b>	<b>319</b>
<i>Pseudocyttus maculatus</i> *	<b>Smooth oreo</b>	<b>592</b>	<b>3 446</b>	<b>2 121</b>	<b>2</b>	<b>2 713</b>	<b>8 125</b>
<i>Pseudophycis bachus</i>	<b>Red cod</b>	<b>298</b>	<b>1 623</b>	<b>225</b>		<b>2 112</b>	<b>3 703</b>
<i>Raja nasuta</i>	Rough skate	2				23	23
<i>Rexea solandri</i>	<b>Gemfish</b>	<b>177</b>	<b>528</b>			<b>631</b>	<b>1 109</b>
<i>Rhinchimera pacifica</i> *	Pacific spookfish	11	15			2	17
<i>Seriotelella brama</i> *	<b>Blue warehou</b>	<b>88</b>	<b>130</b>			<b>894</b>	<b>974</b>
<i>Seriotelella caerulea</i>	<b>White warehou</b>	<b>44</b>	<b>83</b>			<b>192</b>	<b>262</b>
<i>Seriotelella punctata</i> *	<b>Silver warehou</b>	<b>170</b>	<b>198</b>			<b>1 927</b>	<b>2 022</b>
<i>Squalus acanthias</i>	Spiny dogfish	5	11			106	93
<i>Thyrsites atun</i> *	<b>Barracouta</b>	<b>688</b>	<b>8 035</b>	<b>10</b>		<b>7 997</b>	<b>15 542</b>
<i>Trachurus declivis</i>	Jack mackerel	5	8			21	29
<i>Trachurus murphyi</i>	<b>Murphy's mackerel</b>	<b>46</b>	<b>214</b>			<b>225</b>	<b>414</b>
<i>Trachyrincus longirostris</i> *	White rattail	9	9	18		2	29
<i>Trachyscorpia capensis</i> *	Cape scorpionfish	9	9			7	16
<i>Tubbia tasmanica</i> *	<i>Tubbia tasmanica</i>	1	1			0	1
<i>Xenodermichthys</i> sp. *	Black slickhead	8	23			7	30
Total		14 758	130 768	9 804	229	89 074	21 8300

Note 1: Some totals don't add up as prey items were occasionally recorded from regurgitated and everted stomachs.

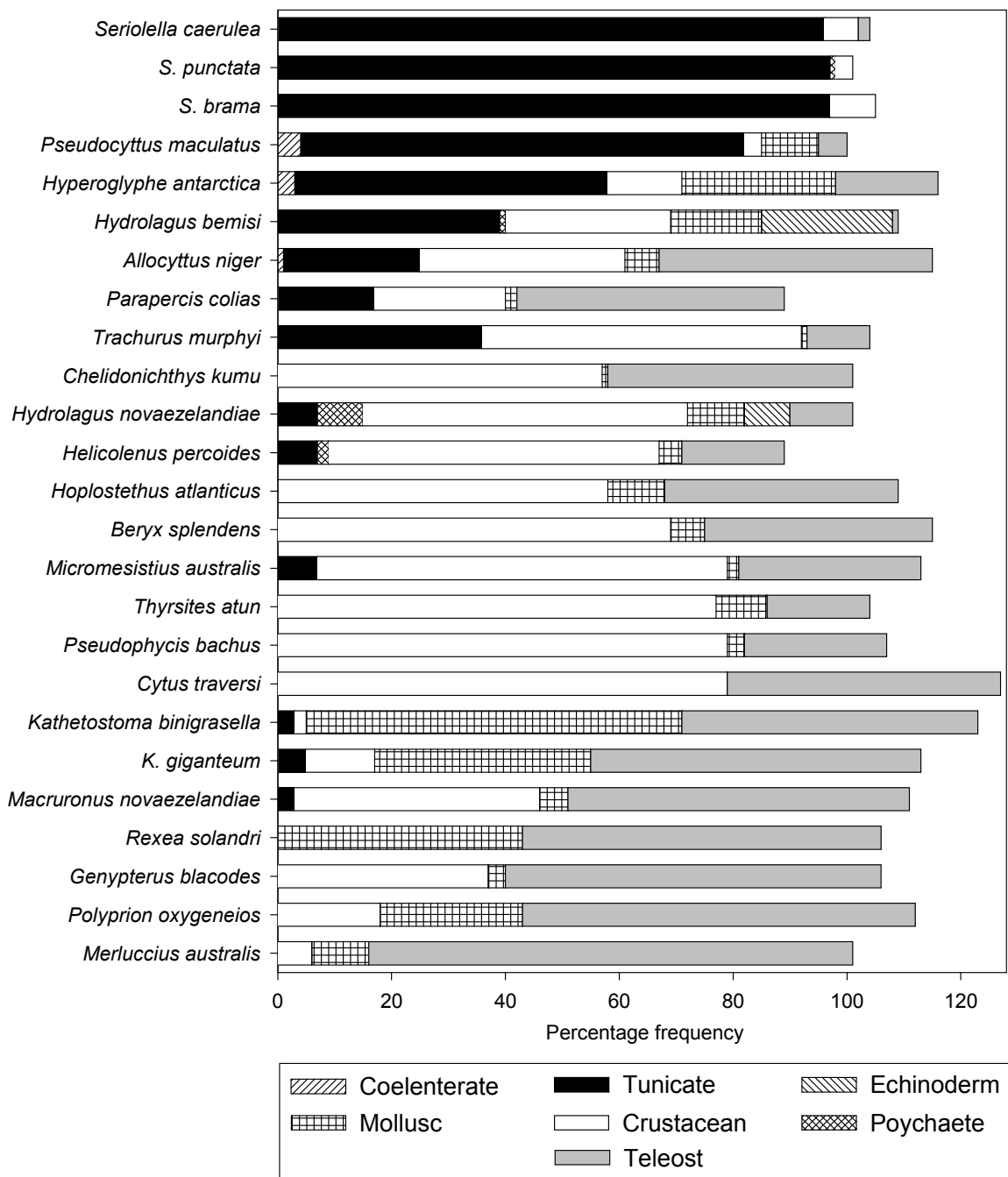





Figure 1: Summary of the percentage frequency of occurrence (%F) of prey groups in the stomach contents of 25 key species collected from New Zealand research trawls.



**APPENDIX A: Tabulated summaries of the dietary characteristics of New Zealand fish species, by depth zone, as reported and standardised in this study from cited literature sources.**

**APPENDIX A: Tabulated summaries of the dietary characteristics of New Zealand fish species, by depth zone, as reported and standardised in this study from cited literature sources.**

**Table A1a: New Zealand inshore (0–100 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤ 10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

















Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Porifera	Coelenterates	Tunicates	Ctenophores	Bryozoans	Echinians	Polychaetes	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	Otago	496	37	< 15	F*																Graham 1938, 1953
2	Avon-Heathcote estuary	1868	30	> 15	F*																Webb 1973
3	Avon-Heathcote estuary	101			P																Webb 1973
1	New Zealand	350+			P																Thomson 1891
2	Otago	326	8	4–52	D																Graham 1938, 1953
3	Wellington Harbour	215	66	>18	P																Baker 1971
4	Avon-Heathcote estuary	9	33	41–64	F																Webb 1973
5	NE North Island				P																Russell 1983
1	Otago				P																Graham 1938, 1953
1	Otago				P																Thomson & Anderton 1921
2	Otago				P																Graham 1938, 1953
1	Moeraki, Fiordland	14	0		P																Thomson 1891
2	Hawke Bay	21	5		P																Phillips 1926
3	Otago	250+			P																Graham 1938, 1953
4	Hauraki Gulf	85	2	21–48	F																Godfriaux 1968, 1970
5	WCSI, Colden & Tasman Bays	2304	18	16–52	F																Ingerson 1996
1	Otago				P																Graham 1938, 1953
1	Otago				P																Graham 1938, 1953
1	New Zealand	23			P																Thomson 1891
2	Otago				P																Graham 1938, 1953
1	NE North Island	2	0	67–97	F																Russell 1983
1	Otago				P																Graham 1938, 1953
1	Ohiwa & Manukau Harbours				P																Graham 1953
2	Hauraki Gulf	8	5	39–67	F																Godfriaux 1968
1	Otago				P																Graham 1938, 1953
1	N North Island	17	0	21–43	F																Clark 1899
2	NE North Island	17		17–33	C																Russell 1983
3	NE North Island				C																Choat & Clements 1992

Table A1a (continued)

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Porifera	Coelenterates	Tunicates	Ctenophores	Bryozoans	Echinurans	Polychaetes	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	New Zealand	711	33		P				P							P	P	P			Thomson 1891
2	Otago				P													P			Thomson & Anderton 1921
3	Palliser Bay	25	0		P			P								P	P	P			Phillipps 1926
4	Otago				P						P					P	P	P			Graham 1938, 1953
5	NE North Island	4	0	57-80	F																Russell 1983
1	Otago				P											P	P	P			Graham 1938, 1953
1	Hauraki Gulf	49	6	15-63	F																Godfriaux 1968, 1970
2	NE North Island	4	25	21-51	F																Russell 1983
1	NE North Island	7	0	45-64	F																Russell 1983
1	New Zealand	36			P																Thomson 1891
2	Otago				P																Thomson & Anderton 1921
3	Otago				P																Graham 1938, 1953
4	Avon-Heathcote estuary	124	2	>12	P						P										Webb 1973
5	NE North Island	18	0	11-24	F																Russell 1983
1	North Island	510			P																Thomson 1891
2	Hawke Bay	2	0		P																Phillipps 1926
3	Hauraki Gulf	957	10		F																Powell 1937
4	Otago & Bay of Plenty				P																Graham 1938, 1953
5	Hauraki Gulf	1194	2	6-76	F																Godfriaux 1968, 1969
6	Hauraki Gulf	6152	36	5-40+	F																Colman 1972
7	W Bay of Plenty	772		8-50+	D																Godfriaux 1974b
8	NE North Island	23	22	27-56	F																Russell 1983
1	Otago	8			P																Thomson 1891
2	Otago				P																Thomson & Anderton 1921
3	Otago				P																Graham 1938, 1953
1	NE North Island	5	0	62-84	F																Russell 1983
1	Mokohinau Islands	7			P																Thomson 1891
2	Otago				P																Graham 1953
3	NE North Island	16	0	25-29	F																Russell 1983

Table A1a (continued)

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Porifera	Coelenterates	Tunicates	Ctenophores	Bryozoans	Echinurans	Polychaetes	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	Southland	77			P																Thomson 1891
2	Otago				P																Thomson & Anderton 1921
3	Otago				P																Graham 1938, 1953
4	Avon-Heathcote estuary	219	22		F*																Webb 1973
5	Wellington Harbour	98	17	15-45	F																Livingston 1987
1	New Zealand	60			P																Thomson 1891
2	Otago - demersal				P																Graham 1938, 1953
3	Hauraki Gulf - demersal	108	0	15-57	F																Godfriaux 1968, 1970
4	Bay of Plenty - surface	24	4	40-47	D																James 1972
5	NE North Island - demersal	7	0	10-49	F																Russell 1983
1	Otago				P																Graham 1938, 1953
2	Avon-Heathcote estuary	360	34	>10	F*																Webb 1973
3	Manukau Harbour	350			F																Pearks 1985
4	Wellington Harbour	116	18	20-39	F																Livingston 1987
1	Otago				P																Thomson & Anderton 1921
2	Otago				P																Graham 1938, 1953
3	Lyttelton Harbour				P																Mundy 1968
4	Akaroa Harbour				P																Mundy 1968
5	Pegasus Bay				P																Mundy 1968
6	off Timaru				P																Mundy 1968
7	Avon-Heathcote estuary	3696	13	> 8	F*																Webb 1973
8	Wellington Harbour	621	9	15-45	F																Livingston 1987
1	Otago				P																Graham 1938, 1953
1	North Island & Cook Strait	7			P																Thomson 1891
2	Otago			< 128	P																Graham 1938, 1953
3	NE North Island	6	33	66-138	F																Russell 1983
1	NE North Island	2	0	85-124	F																Russell 1983
1	Otago				P																Graham 1953

**Table A1a (continued)**

	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Porifera	Celenterates	Tunicates	Ctenophores	Bryozoans	Echinurans	Polychaetes	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
<i>Trachurus novaezelandiae</i>	1	Otago	94	1	9-45	P	▨	▨		▨							▨	▨	▨	▨	P	Graham 1938, 1953
	2	Hauraki Gulf	222	91	Adu	F			▨									▨	▨	▨	▨	Godfriaux 1968, 1970
	3	Bay of Plenty	147	0	Adu	F												▨	▨	▨	▨	Hurst 1980
	4	North Cape	15	20	16-36	F												▨	▨	▨	▨	Hurst 1980
<i>Upeneichthys lineatus</i>	1	NE North Island	270			P				P							▨	▨	▨	▨	P	Russell 1983
	1	Fjordland				P													▨	▨	▨	Thomson 1891
<i>Zeus faber</i>	2	Otago	21	5	15-51	F	▨												▨	▨	▨	Graham 1938
	3	Hauraki Gulf	18	22	25-50	F	▨														▨	Godfriaux 1968, 1970
	4	NE North Island				F															▨	Russell 1983

Table A1a (continued)

	Study no.	Method	Molluscs	Shellfish siphons	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Branchiopods	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Natant decapods	Thalassinids	Hermit crabs	Porcelain crabs	Brachyurans	Myids	Cumaceans	Tanaidaceans	Iso-pods	Amphipods							
<i>Aldrichetta forsteri</i>	1	P	P		P			P				P	P	P																			P					
	2	F*	P																																			
	3	F*																																				
<i>Arripis trutta</i>	1	P	P																																			
	2	P																																				
	3	D																																				
	4	P	P																																			
	5	F																																				
<i>Auchenoceros punctatus</i>	1	P	P																																			
	1	P	P																																			
<i>Callorhynchus mili</i>	2	P	P																																			
<i>Chelidonichthys kumu</i>	1	P	P																																			
	2	P	P																																			
<i>Colistium guntheri</i>	3	P	P																																			
	3	P	P																																			
	4	F																																				
	4	F																																				
	5	F																																				
<i>C. nudipinnis</i>	1	P	P																																			
	1	P	P																																			
	1	P	P																																			
<i>Conger verreauxi</i>	1	P	P																																			
	2	P	P																																			
<i>C. wilsoni</i>	1	F																																				
<i>Crapatus novaehollandiae</i>	1	P																																				
<i>Dasyatis brevicaudatus</i>	1	P																																				
<i>Genyagnus monopterygius</i>	2	F																																				
	1	P	P																																			
<i>Girella tricuspidata</i>	1	P																																				
	2	F																																				
	1	P																																				
	2	F																																				
	3	C																																				

Table A1a (continued)

Species	Study no.	Method	Molluscs	Shellfish siphons	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Branchiopods	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Nantid decapods	Thalassinids	Hermit crabs	Porcelain crabs	Brachyurans	Mysids	Cumaceans	Tanaidaceans	Iso pods	Amphipods							
<i>Latridopsis ciliaris</i>	1	P										P																										
	2	P										P																										
	3	P										P																										
	4	P										P																										
	5	F																																				
<i>Leptoscopus macropygus</i>	1	P										P																										
	1	F																																				
	2	F																																				
	1	F																																				
	1	P											P																									
<i>Notolabrus celidotus</i>	2	P										P																										
	3	P										P																										
	4	P										P																										
	4	P										P																										
	5	F																																				
<i>Pagrus auratus</i>	1	P										P																										
	2	P										P																										
	3	F																																				
	4	P										P																										
	5	F																																				
	2	P																																				
	3	F																																				
	4	P											P																									
<i>Paranotothenia angustata</i>	1	P																																				
	2	P																																				
	3	P										P																										
	1	F																																				
	1	P											P																									
	2	P											P																									
	3	F																																				
	3	F																																				

**Table A1a (continued)**

	Study no.	Method	Molluscs	Shellfish siphons	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Branchiopods	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Nantid decapods	Thalassinids	Hermit crabs	Porcelin crabs	Brachyurans	Mysids	Cumaceans	Tanaidaceans	Iso-pods	Amphipods			
<i>Pelt. novaezeelandiae</i>	1	P	P					P				P																			P			
	2	P										P																		P				
	3	P										P																		P				
	4	F*																																
	5	F																																
<i>Pseudocarax dentex</i>	1	P	P					P				P																						
	2	P										P																						
	3	F																																
	4	D																																
	5	F																																
<i>Rhombosolea leporina</i>	1	P	P									P																						
	2	F*																																
	3	F																																
	4	F																																
<i>R. plebeia</i>	1	P	P									P																						
	2	P										P																						
	3	P										P																						
	4	P										P																						
	5	P										P																						
	6	P										P																						
	7	F*																																
	8	F																																
<i>Sardinops neopilchardus</i>	1	P	P									P																						
	1	P										P																						
<i>Seriola lalandi</i>	2	P										P																						
	3	F																																
<i>Sphyrna zygaena</i>	1	F																																
	1	F																																
<i>Sprattus antipodum</i>	1	P										P																						



**Table A1a (continued)**

	Study no.	Method	Molluscs	Shellfish siphons	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Branchiopods	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Natant decapods	Thalassinids	Hermit crabs	Porcelain crabs	Brachyurans	Mysids	Cumaceans	Tanaidaceans	Iso pods	Amphipods		
<i>Trachurus novaezelandiae</i>	1	P	P									P	P	P																			
	2	F	1,1																														
	3	F																															
	4	F																															
<i>Upeneichthys lineatus</i>	1	F																															
	1	P	P																														
	2	P																															
	3	F																															
<i>Zeus faber</i>	4	F																															

**Table A1a (continued)**

	Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Small fish	Arripidae	Carangidae	Centrolophidae	Clupeidae	Engraulidae	Galaxiidae	Gobiesocidae	Gobiidae	Hemiramphidae	Moridae	Mugilidae	Tripterygiidae	
<i>Aldrichetta forsteri</i>	1	P						P	P														
	2	F*																					
	3	F*						P															
<i>Arripis trutta</i>	1	P						P															
	2	P						P															
	3	D						■															
	4	P						P															
	5	F						■															
<i>Auchenoceros punctatus</i>	1	P																					
<i>Callorhinchus milii</i>	1	P																					
<i>Chelidonichthys kumu</i>	2	P	P					P															
	1	P																					
	2	P																					
	3	P						P															
	4	F																					
	5	F	P					P															
<i>Colistium guntheri</i>	1	P	P					P															
<i>C. nudipinnis</i>	1	P						P															
<i>Conger verreauxi</i>	1	P																					
	2	P						P															
<i>C. wilsoni</i>	1	F																					
<i>Crapatalus novaezelandiae</i>	1	P																					
<i>Dasyatis brevicaudatus</i>	1	P																					
	2	F																					
<i>Genyagnus monopterygius</i>	1	P																					
<i>Girella tricuspidata</i>	1	P						P															
	2	F																					
	3	C																					

**Table A1a (continued)**




	Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Small fish	Arripidae	Carangidae	Centrolophidae	Clupeidae	Engraulidae	Galaxiidae	Gobiesocidae	Gobiidae	Hemiramphidae	Moridae	Mugilidae	Tripterygiidae	
<i>Latridopsis ciliaris</i>	1	P						P															
	2	P																					
	3	P	P																				
	4	P																					
	5	F																					
<i>Leptoscopus macropygus</i>	1	P						P															P
	1	F																					
	2	F																					
<i>Nemadactylus douglasi</i>	1	F																					
	1	P																					
<i>Notolabrus celidotus</i>	2	P																					
	3	P																					
	4	P						P	P														
	5	F																					
	1	P																					
<i>Pagrus auratus</i>	1	P																					
	2	P																					
	3	F																					
	4	P																					
	5	F																					
	6	F																					
	7	D																					
	8	F																					
<i>Paranotothenia angustata</i>	1	P																					
	2	P																					
	3	P																					
<i>Paristiopterus labiosus</i>	1	F																					
	1	P																					
	2	P																					
<i>Parika scaber</i>	2	P																					
	3	F																					

**Table A1a (continued)**

	Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Small fish	Arripidae	Carangidae	Centrolophidae	Clupeidae	Engraulidae	Galaxiidae	Gobiesocidae	Gobiidae	Hemiramphidae	Moridae	Mugilidae	Tripterygiidae	
<i>Pelt. novaezeelandiae</i>	1	P																					
	2	P	P											P									
	3	P	P																				
	4	F*							P														
	5	F								P													
<i>Pseudocaranx dentex</i>	1	P																					
	2	P																					
	3	F																					
	4	D																					
<i>Rhombosolea leporina</i>	5	F																					
	1	P	P																				
	2	F*																					
	3	F																					
	4	F																					
<i>R. plebeia</i>	1	P	P																		P		
	2	P	P																				
	3	P	P																				
	4	P																					
<i>Sardinops neopilchardus</i> <i>Seriola lalandi</i>	5	P																					
	6	P																					
	7	F*																					
	8	F																					
	1	P																					
	1	P																					
	2	P																					
	3	F																					
<i>Sphyrna zygaena</i> <i>Sprattus antipodum</i>	1	F																					
	1	P																					

**Table A1a (continued)**

	Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Small fish	Arripidae	Carangidae	Centrolophidae	Clupeidae	Engraulidae	Galaxiidae	Gobiesocidae	Gobiidae	Hemiramphidae	Moridae	Mugilidae	Tripterygiidae	
<i>Trachurus novaezelandiae</i>	1	P						P															
	2	F							P														
	3	F																					
	4	F																					
<i>Upeneichthys lineatus</i>	1	F																					
	1	P						P															
	2	P																					
	3	F																					
<i>Zeus faber</i>	3	F																					
	4	F																					P

**Table A1b: Southern Hemisphere inshore (0–100 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤ 10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Species	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Eggs	Extraneous matter	Algae	Seaweeds	Coelenterates	Polychaetes	Molluscs	Crustaceans	Insects	Echinoderms	Osteichthyes	Reference
<i>Aldrichetta forsteri</i>	1	W Australia	127		12–23	P	P												Thomson 1954
	2	W Australia	488	37		F													Thomson 1957
	3	W Australia	20		3–5	F													Lenanton, et al. 1982
	4	S Australia	472			B													Edgar & Shaw 1995
<i>Arripis trutta</i>	1	S Australia	40			B													Edgar & Shaw 1995
<i>Callorhynchus milii</i>	1	S Australia	86			B													Edgar & Shaw 1995
<i>Engraulis australis</i>	1	S Australia	23			B													Edgar & Shaw 1995
<i>Girella tricuspidata</i>	1	E Australia	9	33	>15	C													Clements & Choat 1997
<i>Pseudocaranx dentex</i>	1	S Australia	105			B													Edgar & Shaw 1995
<i>Sardinops neopilchardus</i>	1	S Australia	964	6	9–21	F													Stevens et al 1984
	2	S Australia	14			B													Edgar & Shaw 1995
<i>Scomber australasicus</i>	1	S Australia	536	35		F													Stevens et al 1984
<i>Sphyrna zygaena</i>	1	central Pacific	1			F													Strasburg 1958*
	2	E Australia	48	13	151–312	F													Stevens et al 1984
	3	S Africa	144		<200	F													Smale 1991
	4	S Africa	5		>200	F													Smale 1991
	5	S Africa	4			IRI													Sauer and Smale 1991*
<i>Trachurus novaezelandiae</i>	1	S Australia	240	9		F													Stevens et al 1984
<i>Upeneichthys lineatus</i>	2	SW Australia	382	35	6–26	F													Platell et al. 1998

**Table A1b (continued)**

	Study no.	Method	Molluscs	Benthic molluscs	Gastropods	Bivalves	Cephalopods	Decapods	Crustaceans	Crustacean remains	Benthic crustaceans	Planktonic crustaceans	Ostracods	Copepods	Natant decapods	Brachyurans	Mysids	Euphausiids / mysids	Cumaceans	Tanaidaceans	Isopods	Amphipods	Echinoderms	Brittle stars
<i>Aldrichetta forsteri</i>	1	P	P						P															
	2	F																						
	3	F																						
	4	B																						
<i>Arripis trutta</i>	1	B																						
<i>Callorhynchus milii</i>	1	B																						
<i>Engraulis australis</i>	1	B																						
<i>Girella tricuspidata</i>	1	C																						
<i>Pseudocaranx dentex</i>	1	B																						
<i>Sardinops neopilchardus</i>	1	F																						
	2	B																						
<i>Scomber australasicus</i>	1	F																						
<i>Sphyrna zygaena</i>	1	F																						
	2	F																						
	3	F																						
	4	F																						
	5	IRI																						
<i>Trachurus novaezelandiae</i>	1	F																						
<i>Upeneichthys lineatus</i>	2	F																						

**Table A1b (continued)**

	Study no.	Method	Osteichthyes	Fish remains	Other fish	Berycidae	Carangidae	Chelodactylidae	Cupidae	Congiopodidae	Congridae	Engraulidae	Macrouridae	Merlucciidae	Scombridae	Sparidae
<i>Aldrichetta forsteri</i>	1	P														
	2	F														
	3	F														
	4	B														
<i>Arripis trutta</i>	1	B	■													
<i>Callorhynchus milii</i>	1	B														
<i>Engraulis australis</i>	1	B														
<i>Girella tricuspidata</i>	1	C														
<i>Pseudocaranx dentex</i>	1	B														
<i>Sardinops neopilchardus</i>	1	F	▨													
	2	B														
<i>Scomber australasicus</i>	1	F	■													
<i>Sphyrna zygaena</i>	1	F	■													
	2	F	▨													
	3	F	▨													
	4	F	▨													
	5	IRI	■													
<i>Trachurus novaezelandiae</i>	1	F	▨													
<i>Upeneichthys lineatus</i>	2	F	▨													









**Table A2a: New Zealand shelf (100–400 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤ 10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Coelenterates	Tunicates	Bryozoans	Echinurans	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	Otago			< 290	P															Graham 1938, 1953
1	Otago				P											P				Thomson & Anderton 1921
2	Otago				P											P				Graham 1938, 1953
3	Wellington Harbour	103	43	20–35	F															Livingston 1987
1	Otago	2			P															Graham 1953
1	NE North Island	30	13	16–27	F															Russell 1983
1	Otago				P															Graham 1938, 1953
1	Otago	12	100		P															Thomson 1891
2	Otago				P															Graham 1938, 1953
1	Otago				P															Graham 1938, 1953
1	Otago	24			P															Graham 1938, 1953
1	Otago				P															Graham 1938, 1953
1	Otago				P															Graham 1938, 1953
1	Otago	1			P															Graham 1938
2	Otago				P															Thomson & Anderton 1921
1	New Zealand	573	>90		P															Graham 1938, 1953
2	Otago				P															Thomson 1891
3	Otago				P															Thomson & Anderton 1921
1	New Zealand				P															Graham 1938, 1953
2	New Zealand				P															Thomson & Anderton 1921
3	Otago	31	97		P															Graham 1938, 1953
1	Otago				P															Sherrin 1886
1	Otago				P															Thomson 1891
1	Otago				P															Graham 1953
2	Otago	100's			P															Graham 1938
2	Otago	400+			P															Thomson & Anderton 1921
3	Hauraki Gulf	12	0	50–95	F															Graham 1938, 1953
4	Golden Bay	25	♀	juv. ♂	F															Godfriaux 1968
5	Golden Bay	148	♂	mat. ♂	F															King & Clark 1984
6	Golden Bay	49	♀	juv. ♀	F															King & Clark 1984
7	Golden Bay	206	mat. ♀	mat. ♀	F															King & Clark 1984
8	Golden Bay	428			P															King & Clark 1984

**Table A2a (continued)**

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Coelenterates	Tunicates	Bryozoans	Echinurans	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	New Zealand	108			P															Thomson 1891
2	Otago	20	0		P															Thomson & Anderton 1921
3	Palliser Bay & Cook Strait				P															Phillips 1926
4	Otago	50			P															Graham 1938, 1953
5	Cape Campbell & Castlepoint	1420		15-40	D															Marwick 1942
6	W Bay of Plenty	154		15-40	D															Godfriaux 1974a
7	Tasman Bay				D															Godfriaux 1974a
1	Otago	1	0	135	P															Graham 1938, 1953
1	Otago	6286			P															Graham 1938, 1953
1	New Zealand				P															Thomson 1891
2	Otago	400+			P															Thomson & Anderton 1921
3	Otago	250			P															Graham 1938, 1953
4	Chatham Islands	530	47		W															Rapson 1956
5	Malborough Sounds	800	~50		F															Rapson 1956
6	Cook Strait	400		10-30	P															Rapson 1956
7	Auckland	14	14	17-31	F															Mutch 1983
8	NE North Island				F															Russell 1983
1	Otago				P															Thomson & Anderton 1921
2	Otago				P															Graham 1938, 1953
3	Tasman Bay, Marl. Sounds				P															Rapson 1940
4	Wellington Harbour	130	13	20-39	F															Livingston 1987
1	Otago	141			P															Graham 1938, 1953
1	New Zealand	160+			P															Thomson 1891
2	Otago	800+			P															Thomson & Anderton 1921
3	Otago	85			P															Graham 1938, 1953
4	Runnder Point, Cook Strait	2536	~47		F*															Johnston 1983
5	Cook Strait	61			F*															Johnston 1983
6	Cape Campbell				F*															Johnston 1983
1	Otago	10	10	20-35	P															Graham 1938, 1953
2	NE North Island				F															Russell 1983

Table A2a (continued)

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Coelenterates	Tunicates	Bryozoans	Echinurans	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	South Island	372			P															Thomson 1891
2	Otago	100's			P															Thomson & Anderton 1921
3	Cook Strait	2	0		P															Phillipps 1926
4	Otago	1000+			P															Graham 1938, 1953
5	Rangitikei River to Wellington	203		12-69	F															Habib 1975
6	Whakatane R. to Cape Palliser	51		12-47	F															Habib 1975
7	west coast South Island	210		10-57	F															Habib 1975
8	Cloudy Bay to Cape Campbell	219		8-70	F															Habib 1975
9	Canterbury	2294		2-75	V															Habib 1975
10	Otago	258		5-71	F															Habib 1975
11	Southland	122		48-73	F															Habib 1975
1	Otago				P															Graham 1938
2	NE North Island	6	50	6-13	F															Russell 1983
1	Southland	19			P															Thomson 1891
2	Otago				P															Thomson & Anderton 1921
3	Otago				P															Graham 1938, 1953
1	Cook Strait	20			P															Phillipps 1926
2	Otago				P															Thomson & Anderton 1921
3	Otago	350+			P															Graham 1938, 1953
1	Otago				P															Graham 1938, 1953
1	Portland Island	59			P															Thomson 1891
2	Otago				P															Thomson & Anderton 1921
3	Otago				P															Graham 1938, 1953
4	South Island	270		38-68	W															Gavrilov & Markina 1979
1	Otago	4		28-35	P															Graham 1938, 1953
2	South Island	1371		12-18	F															Gavrilov & Markina 1979
3	South Island	1371		24-31	P															Gavrilov & Markina 1979
4	South Island	1371		32-67	?W															Gavrilov & Markina 1979
1	Otago				P															Thomson & Anderton 1921
2	Otago	350+			P															Graham 1938, 1953
3	east coast South Island	7283	29		F															Hanchet 1991

**Table A2a (continued)**

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Algae	Seaweeds	Coelenterates	Tunicates	Bryozoans	Echinurans	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	New Zealand	39	0		P															Thomson 1891
2	Otago				P															Thomson & Anderton 1921
3	Cook Strait	12			P									P						Phillips 1926
4	Otago	400+			P									P						Graham 1938, 1953
5	Castlepoint to Point Gibson	244	58	36-92	P															Mehl 1969
6	west coast South Island	225	47		F															Hurst 1980
7	NE South Island	516	26		F															Hurst 1980
8	Otago - non-schooling	86	3	64-88	W															O'Driscoll 1998
9	Otago - schooling	29	41	64-88	W															O'Driscoll 1998
10	NE North Island	3	0	99-135	F															Russell 1983
1	Bay Of Plenty	49	61		F															Hurst 1980
2	North Cape	16	0		F															Hurst 1980
3	west coast South Island	301	75		F															Hurst 1980
1	Otago				P															Graham 1938, 1953

**Table A2a (continued)**

Study no.	Method	Molluscs	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Natant decapods	Thalassinids	hermit crabs	Porcelain crabs	<i>Munda</i> sp.	Brachyurans	Mysids	Cumaceans	Isopods	Amphipods
1	P																											
1	P																											
2	P																											
3	F																											
1	P																											
1	F																											
1	P																											
1	P																											
2	P																											
1	P																											
1	P																											
1	P																											
1	P																											
2	P																											
1	P																											
2	P																											
3	P																											
1	P																											
2	P																											
3	P																											
1	P																											
2	P																											
3	P																											
1	P																											
2	P																											
3	P																											
1	P																											
2	P																											
3	F																											
4	F																											
5	F																											
6	F																											
7	F																											
8	P																											



**Table A2a (continued)**

Study no.	Method	Molluscs	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Natant decapods	Thalassinids	hermit crabs	Porcelain crabs	<i>Munda</i> sp.	Brachyurans	Mysids	Cumaceans	Isopods	Amphipods
1	P	P								P																		
2	P									P																		
3	P	P								P																		
4	P	P								P																		
5	P	P								P																		
6	D																											
7	D																											
1	P	P								P																		
1	P	P								P																		
1	P	P								P																		
2	P	P								P																		
3	P	P								P																		
4	W																											
5	F																											
6	P	P								P																		
7	P	P								P																		
8	F																											
1	P	P																										
2	P	P								P																		
3	P	P								P																		
4	F																											
1	P	P																										
1	P	P								P																		
2	P	P								P																		
3	P	P								P																		
4	F*																											
5	F*																											
6	F*																											
1	P	P								P																		
2	F																											

**Table A2a (continued)**

	Study no.	Method	Molluscs	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Natant decapods	Thalassinids	ermit crabs	Porcelin crabs	<i>Munda</i> sp.	Brachyurans	Mysids	Cumaceans	Isopods	Amphipods			
<i>Pseudophycis bachus</i>	1	P	P				P				P	P																				
	2	P	P						P		P	P															P					
	3	P									P	P																				
	4	P	P								P	P															P					
	5	F																														
	6	F																														
	7	F																														
	8	F																														
	9	V																														
	10	F																														
	11	F																														
<i>P. breviscula</i>	1	P									P																					
	2	F																														
	1	P	P								P																					
<i>Raja nasuta</i>	2	P																														
	3	P	P								P																					
	1	P	P								P																					
<i>Rexea solandri</i>	2	P	P																													
	3	P	P																													
	1	P	P								P																					
<i>Rhombosolea tapirina</i> <i>Seriolella brama</i>	1	P	P								P																					
	2	P	P																													
	3	P	P								P																					
	1	P									P																					
<i>Seriolella punctata</i>	2	P	P								P																					
	3	P																														
	4	?W																														
	1	P	P																													
<i>Squalus acanthias</i>	1	P	P								P																					
	2	P	P								P																					
	3	F																														

**Table A2a (continued)**

Study no.	Method	Molluscs	Gastropods	Chitons	Bivalves	Shellfish	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Other crustaceans	Seed shrimps	Copepods	Barnacles	Mantis shrimps	Euphausiids	Natant decapods	Thalassinids	hermit crabs	Porcelain crabs	<i>Munda</i> sp.	Brachyurans	Mysids	Cumaceans	Isopods	Amphipods
1	P	P				P				P																		
2	P									P																	P	
3	P	P								P																	P	
4	P	P								P																	P	
5	P									P																	P	
6	F																											
7	F																											
8	W																											
9	W																											
10	F																											
1	F																											
2	F																											
3	F																											
3	F																											
1	P																											

*Thysites atun*

*Trachurus declivis*

*Typhlonarke aysoni*

Table A2a (continued)

Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Fish eggs	Small fish	Other fish	Argentinidae	Bothidae	Carangidae	Centrolophidae	Chelodactylidae	Clupeidae	Engraulidae	Gobiidae	Gobiesocidae	Macrouridae	Merlucciidae	Moridae	Mugilidae	Myctophidae	Notheniidae	Percophidae	Peuronectidae
1	P						P										P											
1	P						P										P											
2	P	P		P			P																					
3	F																											
1	P																											
1	F																											
1	P						P																					
1	P																											
2	P																											
1	P																											
1	P	P					P																					
1	P																											
1	P																											
2	P																											
1	P																											
2	P																											
3	P																											
1	P																											
2	P																											
3	P																											
1	P																											
1	P																											
2	P																											
3	F																											
4	F																											
5	F																											
6	F																											
7	F																											
8	P						P																					

**Table A2a (continued)**

	Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Fish eggs	Small fish	Other fish	Argentinidae	Bothidae	Carangidae	Centrolophidae	Cheilodactylidae	Clupeidae	Engraulidae	Gobiidae	Gobiessocidae	Macrouridae	Mercuroidae	Moridae	Mugilidae	Mycophidae	Nototheniidae	Percophidae	Pleuronectidae
<i>Nemadactylus macropterus</i>	1	P						P																					
	2	P																											
	3	P																											
	4	P																											
	5	P						P	P																				
	6	D																											
	7	D																											
	1	P																											
<i>Notolabrus cinctus</i> <i>Notorynchus cepedianus</i> <i>Paraperis colias</i>	1	P						P																					
	2	P						P																					
	3	P						P																					
	4	W																											
	5	F																											
	6	P						P																					
	7	P						P																					
	8	F																											
<i>Peloretis flavilatus</i>	1	P																											
	2	P						P																					
	3	P																											
	4	F																											
<i>Polyprion americanus</i> <i>P. oxygeneios</i>	1	P																											
	2	P																											
	3	P																											
	4	F*																											
	5	F*																											
	6	F*																											
<i>Pseudolabrus miles</i>	1	P																											
	2	F																											

Table A2a (continued)

	Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Fish eggs	Small fish	Other fish	Argentinidae	Bothidae	Carangidae	Centrolophidae	Cheilodactylidae	Clupeidae	Engraulidae	Gobiidae	Gobiesocidae	Macrouridae	Merlucciidae	Moridae	Mugilidae	Mycophidae	Notothenidae	Percophidae	Pleuronectidae	
<i>Pseudophycis bachus</i>	1	P						P			P		P					P												
	2	P						P					P					P												
	3	P						P					P					P												
	4	P					P	P					P					P												
	5	F																												
	6	F																												
	7	F																												
	8	F																												
	9	V																												
	10	F																												
	11	F																												
<i>P. breviscula</i>	1	P																												
	2	F																												
<i>Raja nasuta</i>	1	P						P																						
	2	P																												
<i>Rexea solandri</i>	3	P																												
	1	P																												
	2	P																												
<i>Rhombosolea tapirina</i>	3	P																												
	1	P																												
	1	P																												
<i>Seriolaella brama</i>	2	P																												
	3	P																												
<i>Seriolaella punctata</i>	4	W																												
	1	P																												
	2	F																												
	3	P																												
<i>Squalus acanthias</i>	4	?W																												
	1	P																												
	2	P																												
	2	P																												
	3	F																												

**Table A2a (continued)**

Study no.	Method	Echinoderms	Starfish	Brittle stars	Urchins	Sea cucumbers	Osteichthyes	Fish remains	Fish eggs	Small fish	Other fish	Argentinidae	Bothidae	Caramidae	Centrolophidae	Chelodactylidae	Clupeidae	Engraulidae	Gobiidae	Gobiesocidae	Macrouridae	Mercenridae	Moridae	Mugilidae	Myctophidae	Nothoenidae	Percophidae	Pleuronectidae
1	P						P							P			P											
2	P						P							P			P						P					
3	P						P							P			P						P					
4	P						P							P			P						P					
5	P						P							P			P						P					
6	F						Diagonal lines (top-left to bottom-right)																					
7	F						Diagonal lines (top-left to bottom-right)																					
8	W						Diagonal lines (top-left to bottom-right)																					
9	W						Diagonal lines (top-left to bottom-right)																					
10	F						Solid black							Diagonal lines (top-left to bottom-right)														
1	F																											
2	F																											
3	F																											
3	F						Diagonal lines (top-left to bottom-right)																					
1	P						P																					

*Thyrsites atun*

*Trachurus declivis*

*Typhlonarke aysoni*

**Table A2a (continued)**

	Study no.	Method	Pomacentridae	Retropinnidae	Scorpaenidae	Triglidae	Tripterygiidae
<i>Alopias vulpinus</i>	1	P					
<i>Arnoglossus scapha</i>	1	P					
	2	P					
	3	F					
<i>Caelorhynchus biclinozonalis</i>	1	P					
<i>Caesioperca lepidoptera</i>	1	F					
<i>Cephaloscyllium isabellum</i>	1	P					P
<i>Congiopodus leucopaecilus</i>	1	P					
	2	P					
<i>Cyttus novaezelandiae</i>	1	P					
<i>Galeorhinus galeus</i>	1	P					
<i>Gonorynchus gonorynchus</i>	1	P			P		
<i>Hemerocoetes</i> sp.	1	P					
<i>Kathetostoma giganteum</i>	1	P					
	2	P					P
<i>Latris lineata</i>	1	P					
	2	P					
	3	P					
<i>Lepidopus caudatus</i>	1	P					
	2	P					
	3	P					
<i>Lepidotrigla brachyoptera</i>	1	P					
<i>Mustelus lenticulatus</i>	1	P					
	2	P					
	3	F					
	4	F					
	5	F					
	6	F					
	7	F					
	8	P					



**Table A2a (continued)**




	Study no.	Method	Pomacentridae	Retropinnidae	Scorpaenidae	Triglidae	Tripterygiidae
<i>Nemadactylus macropterus</i>	1	P					
	2	P					
	3	P					
	4	P					
	5	P					
	6	D					
	7	D					
<i>Notolabrus cinctus</i>	1	P					
	1	P					
	1	P					
	2	P					P
<i>Notorynchus cepedianus</i>	3	P					P
	4	W					
	5	F					
	6	P					P
<i>Peloretis flavilatus</i>	7	P					
	8	F					
	1	P					
	2	P					
	3	P					
	4	F					
	1	P					
	1	P					
<i>Polyprion americanus</i> <i>P. oxygeneios</i>	2	P					
	3	P			P		
	4	F*					
	5	F*					
	6	F*					
	1	P					
<i>Pseudolabrus miles</i>	1	P					
	2	F					

**Table A2a (continued)**

	Study no.	Method	Pomacentridae	Retropinnidae	Scorpaenidae	Triglidae	Tripterygiidae
<i>Pseudophycis bachus</i>	1	P					
	2	P			P		
	3	P			P		
	4	P		P	P		P
	5	F					
	6	F					
	7	F					
	8	F					
	9	V					
	10	F					
	11	F					
<i>P. breviscula</i>	1	P					
2	F						
<i>Raja nasuta</i>	1	P					
	2	P					
	3	P					
<i>Rexea solandri</i>	1	P					
	2	P					
	3	P					
<i>Rhombosolea tapirina</i> <i>Seriolella brama</i>	1	P			P		
	1	P					
	2	P					
	3	P					
<i>Seriolella punctata</i>	4	W					
	1	P					
	2	F					
	3	P					
4	?W						
<i>Squalus acanthias</i>	1	P					
	2	P					
	3	F					

**Table A2a (continued)**

	Study no.	Method	Pomacentridae	Retropinnidae	Scorpaenidae	Triglidae	Tripterygiidae
<i>Thyrsites atun</i>	1	P					
	2	P					
	3	P					
	4	P			P	P	
	5	P					
	6	F					
	7	F					
	8	W					
	9	W					
	10	F					
<i>Trachurus declivis</i>	1	F					
	2	F					
	3	F					
<i>Typhlonarke aysoni</i>	1	P					

**Table A2b: Southern Hemisphere shelf (100–400 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤ 10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Extraneous matter	Foraminiferans	Coelenterates	Tunicates	Bryozoans	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Chondrichthyes	Osteichthyes	Cetacean remains	Piniped remains	Reference
1	WC S Africa	1615		12–36	F															Roshchin 1986
2	WC S Africa	48	8	14–49	F															Meyer & Smale 1991a
3	WC S Africa	82	8	14–49	F															Meyer & Smale 1991a
1	Argentina	12	17		F															Menni et al. 1982*
2	Argentina	18	33		F															Menni et al. 1982*
3	S Africa	5	40		F															Sauer & Smale 1991*
1	S Africa	62	21	31–42	N															Meyer & Smale 1991b
1	E Tasmania	948	28	8–38	F															Blaber & Bulman 1987
1	S Africa	10	40		F															Bass et al. 1975*
1	E Tasmania	78	24	97–149	F															Blaber & Bulman 1987
2	SC, S Africa	75	18	24–60	F															Meyer & Smale 1991a
3	SC, S Africa	40	18	85–134	F															Meyer & Smale 1991a
4	WC, S Africa	66	18	34–60	F															Meyer & Smale 1991a
5	WC, S Africa	19	18	60–85	F															Meyer & Smale 1991a
6	WC, S Africa	37	18	85–124	F															Meyer & Smale 1991a
1	SE Australia - plank. prey	111			F															Clarke 1985
2	SE Australia - benthic prey	71			F															Clarke 1985
1	E Cape, S Africa	91	30		F															Ebert 1991
2	W Cape, S Africa	103	29		F															Ebert 1991
3	Namibia	94	32		F															Ebert 1991
4	Namibia	46	33		F															Ebert 1991
5	Argentina	5			F															Menni et al. 1982*
1	Argentina	20	60	59–79	F															Menni & Lopez 1979
1	SE Pacific	138	13	65–115	F*															Rojas et al. 1985
1	S Australia	69			B															Edgar & Shaw 1995
1	S Australia	43			B															Edgar & Shaw 1995
1	S Africa	176	31	32–91	F															Ebert et al. 1992
1	SE Pacific	110	66		P															Litvinov 1990
2	S Africa	271	22	23–102	F															Ebert et al. 1992
3	S Africa	1			F															Lipinsky et al. 1992*

**Table A2b (continued)**

	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Coelenterates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
<i>Trachurus declivis</i>	1	S Australia - summer	550	24	11-35	F	■	■	■	■	■	■	■	■	■	Stevens et al 1984
	2	S Australia - spring	143	48	11-35	F	■	■	■	■	■	■	■	■	■	Stevens et al 1984
	3	S Australia - winter	485	31	11-35	F	■	■	■	■	■	■	■	■	■	Stevens et al 1984
	4	E Tasmania	169	7	21-37	F	■	■	■	■	■	■	■	■	■	Blaber & Bulman 1987
<i>T. murphyi</i>	1	Peru	92	11	32-38	F	■	■	■	■	■	■	■	■	■	Konchina 1978
	2	Peru	280	10	22-62	F	■	■	■	■	■	■	■	■	■	Konchina 1979
	3	Peru	2381	45	23-64	F	■	■	■	■	■	■	■	■	■	Konchina 1980
<i>Zenopsis nebulosus</i>	1	SE Pacific	20		25-46	F	■	■	■	■	■	■	■	■	■	Parin et al. 1988
	2	SE Pacific	36		27-33	F	■	■	■	■	■	■	■	■	■	Parin et al. 1988
	3	SE Pacific	11		26-40	F	■	■	■	■	■	■	■	■	■	Parin et al. 1988
	4	SE Pacific	5		26-41	F	■	■	■	■	■	■	■	■	■	Parin et al. 1988

**Table A2b (continued)**

Study no.	Method	Molluscs	Benthic molluscs	Gastropods	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Larval crustaceans	Benthic crustaceans	Ostracods	Copepods	Stomatopods	Euphausiids	Decapods	Natant decapods	<i>Munda</i> sp.	Brachyurans	Mysids	Mysid remains	Isopods	Amphipods	Echinoderms	Brittle stars
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal														Diagonal		
3	F	Diagonal						Diagonal	Diagonal														Diagonal		
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
3	F	Diagonal						Diagonal	Diagonal																
1	N	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
3	F	Diagonal						Diagonal	Diagonal																
4	F	Diagonal						Diagonal	Diagonal																
5	F	Diagonal						Diagonal	Diagonal																
6	F	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
3	F	Diagonal						Diagonal	Diagonal																
4	F	Diagonal						Diagonal	Diagonal																
5	F	Diagonal						Diagonal	Diagonal																
1	F*	Diagonal						Diagonal	Diagonal																
1	B	Diagonal						Diagonal	Diagonal																
1	B	Diagonal						Diagonal	Diagonal																
1	F	Diagonal						Diagonal	Diagonal																
1	P	Diagonal						Diagonal	Diagonal																
2	F	Diagonal						Diagonal	Diagonal																
3	F	Diagonal						Diagonal	Diagonal																

**Table A2b (continued)**

	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Crustaceans	Crustacean remains	Larval crustaceans	Ostracods	Copepods	Stomatopods	Euphausiids	Decapods	Natant decapods	Brachyurans	Mysids	Euphausiids / mysids	Isopods	Amphipods	Echinoderms	Brittle stars	
<i>Trachurus declivis</i>	1	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	2	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	3	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	4	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
<i>T. murphyi</i>	1	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	2	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	3	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
<i>Zenopsis nebulosus</i>	1	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	2	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	3	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal
	4	F	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal	Diagonal




Table A2b (continued)

Species	Study no.	Method	Chondrichthyes	Other Chondrichthyes	Elasmobranchii	Hexanchidae	Scyliorhinidae	Squalidae	Triakidae	Dasyatidae	Myliobatidae	Osteichthyes	Fish remains	Fish scales	Other fish	Callionymidae	Carangidae	Cupidae	Emmelichthyidae	Engraulidae	Gempyidae	Gonostomatidae	Mercuroidae	Mycophoridae	Lamppanyctodes hectoris	Photichthyidae	Scorpaenidae	Sparidae	Sternopychidae	Trichuridae
<i>Emmelichthys nitidus</i>	1	F																												
	2	F																												
	3	F																												
<i>Galeorhinus galeus</i>	1	F																												
	2	F																												
	3	F																												
<i>Gonorynchus gonorynchu.</i>	1	N																												
<i>Helicolenus percoides</i>	1	F																												
<i>Hepranchias perlo</i>	1	F																												
<i>Lepidopus caudatus</i>	1	F																												
	2	F																												
	3	F																												
	4	F																												
	5	F																												
	6	F																												
<i>Macrorhamphosus scolop.</i>	1	F																												
	2	F																												
<i>Notorynchus cepedianus</i>	1	F																												
	2	F																												
	3	F																												
	4	F																												
	5	F																												
<i>Polyprion americanus</i>	1	F																												
<i>P. oxygeneios</i>	1	F*																												
<i>Pseudophycis bachus</i>	1	B																												
<i>Rhombosolea tapirina</i>	1	B																												
<i>Squalus acanthias</i>	1	F																												
	1	P																												
<i>S. mitsukurii</i>	2	F																												
	3	F																												



**Table A2b (continued)**

	Study no.	Method
<i>Trachurus declivis</i>	1	F
	2	F
	3	F
	4	F
<i>T. murphyi</i>	1	F
	2	F
	3	F
<i>Zenopsis nebulosus</i>	1	F
	2	F
	3	F
	4	F
Osteichthyes		F
Fish remains		F
Other fish		F
Apogonidae		F
Bathylagidae		F
Bregmacerotidae		F
Carangidae		F
Emmelichthyidae		F
Engraulidae		F
Gempylidae		F
Gonostomatidae		F
Myctophidae		F
<i>Lamppanyctodes hectoris</i>		F
Nomidae		F
Photichthyidae		F
Scorpaenidae		F
Sternopychidae		F
Trichiuridae		F

**Table A2c: Northern Hemisphere shelf (100–400 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤ 10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Species	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Molluscs	Crustaceans	Chondrichthyes	Osteichthyes	Marine mammals	Reference
<i>Alopias vulpinus</i>	1	Mediterranean	4			F							Capape 1975*
	2	NE Atlantic	1			F							Pascoe 1986*
<i>Cephaloscyllium isabellum</i>	3	Mexico	926	59		F							Castillo et al. 1992*
	1	NW Pacific	356	28		F							Taniuchi 1988
<i>Galeorhinus galeus</i>	1	Mediterranean	1			F							Capape 1975*
	2	NE Atlantic	169		< 105	F							Gomes et al. 1998
<i>Hepranchias perlo</i>	3	N Atlantic	46	9		F							Ellis et al. 1996*
	1	Mediterranean	9	22		F							Capape 1975*
<i>Lepidopus caudatus</i>	1	Mediterranean	145		20–69	W							Macpherson 1981
	2	NE Atlantic	70		74–173	F							Gomes et al. 1998
<i>Notorynchus cepedianus</i>	1	NE Pacific	38			F							Ebert 1989*
	1	N Atlantic				P							Vinnichenko 1997
<i>Squalus mitsukurii</i>	1	C Pacific	293	66		F							Wilson & Seki 1994



**Table A3a: New Zealand upper slope (400–700 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Species	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
<i>Argentina elongata</i>	1	Otago				P										Graham 1938, 1953
<i>Caelorinchus aspercephalus</i>	2	Campbell Plateau	132			F										Clark 1985
<i>C. oliverianus</i>	1	Campbell Plateau	46	21–60		F										Clark 1985
<i>Centriscoptes humerosus</i>	1	SW Chatham Rise	10			F										Clark et al 1989
<i>Genypterus blacodes</i>	1	Otago	6	83		P										Graham 1938, 1953
	1	Otago, Milford Sound	6	83		P										Thomson 1891
	2	Otago	100's			P										Thomson & Anderton 1921
	3	CKST	6	17		P										Phillips 1926
	4	Otago	500+			P										Graham 1938, 1953
	5	North Island	57	30–40–160		F										Mitchell 1984
	6	Chatham Rise	300	34–30–140		F										Mitchell 1984
	7	WCSI	56	36–60–150		F										Mitchell 1984
	8	Campbell Plat./Pukaki Rise	126	34–40–120		F										Mitchell 1984
	9	Campbell Plateau	50	70–130		F										Clark 1985
<i>Gollum attenuatus</i>	1	New Zealand	747	33		IRI										Yano 1993
<i>Helicolenus percoides</i>	1	Otago				P										Thomson & Anderton 1921
	2	Otago	300+			P										Graham 1938, 1953
<i>Hoplostethus mediterraneus</i>	1	northern New Zealand	173		5–18	F										Kerstan 1989
<i>Hyperoglyphe antarctica</i>	1	Otago	200+			P										Graham 1938, 1953
<i>Lepidorhynchus denticulatus</i>	1	Campbell Plateau	82	26–50		F										Clark 1985
	2	SW Chatham Rise	40			F										Clark et al 1989
<i>Macruronus novaezelandiae</i>	1	Otago	5			P										Thomson & Anderton 1921
	2	CKST	4	25–125		P										Phillips 1926
	3	Otago	4	507 50*		F										Graham 1938, 1953
	4	East region	207 60*			F										Kuo & Tanaka 1984
	5	Northwest region	451			F										Kuo & Tanaka 1984
	6	South region	544	40–100		F										Kuo & Tanaka 1984
	7	Campbell Plateau	544	40–100		F										Clark 1985
<i>Merluccius australis</i>	1	Otago				P										Graham 1938, 1953

**Table A3a (continued)**

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
1	Bounty Rise	101		♀ + ♂	F										Shpak 1976
2	Bounty Rise			♀ + ♂	F										Shpak 1976
3	Pukaki Rise	110		♀ + ♂	F										Shpak 1976
4	Pukaki Rise			♀ + ♂	F										Shpak 1976
5	Campbell Plateau	231		♀ + ♂	F										Shpak 1976
6	Campbell Plateau			♀ + ♂	F										Shpak 1976
7	Campbell Plateau	65		15-20	F										Shpak 1976
8	Campbell Plateau	475		16-54	F										Clark 1985
1	Otago				P				P						Graham 1938, 1953
1	Shares Islands	3	0	juv.	F										Fenwick 1978
2	Campbell Plateau	56		20-70	F										Clark 1985
1	South Island	224		30-64	W										Gavrillov & Markina 1979

*Neophrynichthys latus*  
*Paranotothenia microlepidota*

*Seriolella caerulea*

**Table A3a (continued)**

	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Other crustaceans	Ostracods	Copepods	Euphausiids	Decapods	Natant decapods	Thalassinids	Hermit crabs	<i>Munda</i> sp.	Brachyurans	Mysids	Iso-pods	Amphipods
<i>Argentina elongata</i>	1	P																				
<i>Caelorrinchus aspercephalus</i>	2	F																				
<i>C. oliverianus</i>	1	F																				
<i>Centriscopus humerosus</i>	1	F																				
<i>Genypterus blacodes</i>	1	P																				
	2	P																				
	3	P																				
	4	P																				
	5	F																				
	6	F																				
	7	F																				
	8	F																				
	9	F																				
<i>Gollum attenuatus</i>	1	IRI																				
<i>Helicolenus percoides</i>	1	P																				
	2	P																				
<i>Hoplostethus mediterraneus</i>	1	F																				
<i>Hyperoglyphe antarctica</i>	1	P																				
<i>Lepidorhynchus denticulatus</i>	1	F																				
	2	F																				
<i>Macruronus novaezelandiae</i>	1	P																				
	2	P																				
	3	P																				
	4	F																				
	5	F																				
	6	F																				
	7	F																				
<i>Merluccius australis</i>	1	P																				

**Table A3a (continued)**

	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Octopods	Crustaceans	Crustacean remains	Other crustaceans	Ostracods	Copepods	Euphausiids	Decapods	Natant decapods	Thalassinids	Hermit crabs	<i>Munda</i> sp.	Brachyurans	Mysids	Iso-pods	Amphipods
<i>Micromesistius australis</i>	1	F						Diagonal lines					Diagonal lines		Diagonal lines							Diagonal lines
	2	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
	3	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
	4	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
	5	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
	6	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
	7	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
	8	F						Diagonal lines				Diagonal lines	Diagonal lines		Diagonal lines							Diagonal lines
<i>Neophrynychthys latus</i>	1	P						Diagonal lines														Diagonal lines
	1	F						Diagonal lines														Diagonal lines
	2	F						Diagonal lines														Diagonal lines
<i>Paranotothenia microlepidota</i>	1	F						Diagonal lines														Diagonal lines
	2	F						Diagonal lines														Diagonal lines
<i>Seriola caerulea</i>	1	W						Diagonal lines														Diagonal lines




Table A3a (continued)



































Species	Study no.	Method	Echinoderms	Brittle stars	Osteichthyes	Fish remains	Other fish	Argentinidae	Bothidae	Clupeidae	Congridae	Gadidae	Macrouridae	Merlucciidae	Moridae	Mycophidae	<i>Lampanyctus hectoris</i>	Ophidiidae	Percophidae	Photichthyidae	Pleuronectidae	Scorpaenidae	Tripterygiidae	
<i>Argentina elongata</i>	1	P																						
<i>Caelorinchus aspercephalus</i>	2	F																						
<i>C. oliverianus</i>	1	F																						
<i>Centriscoops humerosus</i>	1	F																						
<i>Genypterus blacodes</i>	1	P																						
	2	P																						
	3	P																						
	4	P																						
	5	F																						
	6	F																						
	7	F																						
	8	F																						
	9	F																						
<i>Gollum attenuatus</i>	1	IRI																						
<i>Helicolenus percoides</i>	1	P																						
	2	P																						
<i>Hoplostethus mediterraneus</i>	1	F																						
<i>Hyperoglyphe antarctica</i>	1	P																						
<i>Lepidorhynchus denticulatus</i>	1	F																						
	2	F																						
<i>Macruronus novaezelandiae</i>	1	P																						
	2	P																						
	3	P																						
	4	F																						
	5	F																						
	6	F																						
	7	F																						
<i>Merluccius australis</i>	1	P																						



**Table A3a (continued)**

	Study no.	Method	Echinoderms	Brittle stars	Osteichthyes	Fish remains	Other fish	Argentiniidae	Bothidae	Clupeidae	Congridae	Gadidae	Macrouridae	Mercuroidae	Moridae	Mycetophidae	<i>Lampan. hectoris</i>	Ophidiidae	Percophidae	Photichthyidae	Pleuronectidae	Scorpaenidae	Tripterygiidae	
<i>Micromesistius australis</i>	1	F			█																			
	2	F			█																			
	3	F			█																			
	4	F			█																			
	5	F			█																			
	6	F			█																			
	7	F			█																			
	8	F			█																			
<i>Neophrynichthys latus</i>	1	P																						
	1	F																						
	2	F																						
<i>Paranotothenia microlepidota</i>	1	F																						
	1	W																						
<i>Seriola caerulea</i>	1	F																						
	1	W																						

**Table A3b: Southern Hemisphere upper slope (400–700 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; ,  $\leq 10\%$  importance in diet; , 11 to 50 % importance in diet; ,  $>51\%$  importance in diet.**

	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Osteichthyes	Reference
<i>B. splendens</i>	1	SE Pacific	59	0	15–31	W									Dubochkin & Kotlyar 1989
<i>Brama brama</i>	1	E Tasmania	129	7	24–48	F									Blaber & Bulman 1987
<i>Cytus triversi</i>	1	E Tasmania	433	43	9–56	F									Blaber & Bulman 1987
<i>Epigonus denticulatus</i>	1	S Africa	59	12	10–18	F									Meyer & Smale 1991a
	2	S Africa	49	12	18–22	F									Meyer & Smale 1991a
<i>Genypterus blacodes</i>	1	E Tasmania	449	46	45–130	F									Blaber & Bulman 1987
<i>Hyperoglyphe antarctica</i>	1	SE Australia	78	45		F									Winstanley 1978
<i>Lampanyctodes hectoris</i>	1	E Tasmania	975	19	3–7	F									Young & Blaber 1986
<i>Lepidorhynchus denticulatus</i>	1	E Tasmania	655	15	8–51	F									Blaber & Bulman 1987
<i>Macruronus novaezelandiae</i>	1	E Tasmania	243	8	15–29	F									Bulman & Blaber 1986
	2	E Tasmania	841	32	30–120	F									Bulman & Blaber 1986
	3	Bass Strait	452	38	30–120	F									Bulman & Blaber 1986
	4	WC Tasmania	303	42	30–120	F									Bulman & Blaber 1986
<i>Maurolicus muelleri</i>	1	E Tasmania	719	22	2–6	F									Young & Blaber 1986
	2	SE Pacific	59	0		F									Gorelova & Krasil'nikova 1990
	3	S Africa	57	0	3–4	F									Gorelova & Krasil'nikova 1990
<i>Merluccius australis</i>	1	Chile, S Pacific			51–60	W									Paya 1992
	2	Chile, S Pacific			61–70	W									Paya 1992
	3	Chile, S Pacific			71–80	W									Paya 1992
	4	Chile, S Pacific			81–90	W									Paya 1992
	5	Chile, S Pacific			91–100	W									Paya 1992




**Table A3b (continued)**


















	Study no.	Method	Molluscs	Gastropods	Decapods	Crustaceans	Crustacean remains	Ostracods	Copepods	Euphausiids	Decapods	Natant decapods	Thalassinids	<i>Munda</i> sp.	Brachyurans	Mysids	Iso-pods	Amphipods	
<i>B. splendens</i>	1	W	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Brama brama</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Cytus traversi</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Epigonus denticulatus</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	2	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Genypterus blacodes</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Hyperoglyphe antarctica</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Lampanyctodes hectoris</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Lepidorhynchus denticulatus</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Macrurus novaezelandiae</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	2	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	3	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	4	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Maurollicus muelleri</i>	1	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	2	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	3	F	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
<i>Merluccius australis</i>	1	W	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	2	W	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	3	W	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	4	W	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■
	5	W	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■

Table A3b (continued)

	Study no.	Method	Osteichthyes	Fish remains	Fish eggs	Callionymidae	Gadidae	Macrouridae	Coelornchus spp.	Lepidorhynchus denticulatus	Merlucciidae	Moridae	Daphus spp.	Lampanyctodes hectoris	Scorpaenidae	Sternopychidae	
<i>B. splendens</i>	1	W	■														
<i>Brama brama</i>	1	F	■	■										■			
<i>Cytus traversi</i>	1	F	■	■					▨					■			
<i>Epigonus denticulatus</i>	1	F	■	■					▨					■			
	2	F	■	■	■												
<i>Genypterus blacodes</i>	1	F	■	■					▨					■			
<i>Hyperoglyphe antarctica</i>	1	F	■	■					▨					■			
<i>Lampanyctodes hectoris</i>	1	F	■	■					▨					■			
<i>Lepidorhynchus denticulatus</i>	1	F	■	■					▨					■			
<i>Macruronus novaezelandiae</i>	1	F	■	■					▨					■			
	2	F	■	■					▨					■			
	3	F	■	■					▨					■			
	4	F	■	■					▨					■			
<i>Mauroliticus muelleri</i>	1	F	■	■					▨					■			
	2	F	■	■					▨					■			
	3	F	■	■					▨					■			
<i>Merluccius australis</i>	1	W	■	■					▨		■			■			
	2	W	■	■					▨		■			■			
	3	W	■	■					▨		■			■			
	4	W	■	■					▨		■			■			
	5	W	■	■					▨		■			■			

















**Table A3c: Northern Hemisphere upper slope (400–700 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; ,  $\leq 10\%$  importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Osteichthyes	Reference
<i>Beryx decadactylus</i>	1	NE Atlantic	1		25	P								P		Mauchline & Gordon 1984a
	2	NE Atlantic	46	23–48	F											Gomes et al. 1998
<i>B. splendens</i>	1	Atlantic	67	16–33	W											Dubochkin & Kotlyar 1989
	2	NE Atlantic	294	18–39	F								P	P	P	Gomes et al. 1998
<i>Caelorinchus fasciatus</i>	3	NW Atlantic				P					P					Vinnichenko 1997
	1		6	<31	P											McLellan 1977
	1	Irish Sea	1	47	P			P						P		McKee 1981
<i>Centrolophus niger</i>	1	NW Pacific	230			F										Bata et al. 1987*
<i>Eimopterus lucifer</i>	1	NE Atlantic	23	5–24	P									P		Marshall & Merrett 1977
	2	NE Atlantic	51	25–43	F											Du Buit 1978
<i>Lepidorhynchus denticulatus</i>	3	NW Atlantic				P								P		Vinnichenko 1997
	1	Atlantic	4			P								P		McLellan 1977
	1	NW Pacific	9	<52	P								P			McLellan 1977
<i>Malacocephalus laevis</i>	2	N Atlantic				P										McLellan 1977
	3	NE Atlantic	24	33–54	F											Mauchline & Gordon 1984b
<i>Maurolicus muelleri</i>	1	NE Atlantic	180	1–5	F											Mauchline & Gordon 1983b

**Table A3c (continued)**

	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Crustaceans	Crustacean remains	Other crustaceans	Ostracods	Copepods	Euphausiids	Decapods	Decapod remains	Natant decapods	<i>Munida</i> sp.	Mysids	Euphausiids / mysids	Amphipods	Osteichthyes	Fish remains	Mycetophidae	
<i>Beryx decadactylus</i>	1	P																					
	2	F																					
<i>B. splendens</i>	1	W																					
	2	F																					
<i>Caelorinchus fasciatus</i>	3	P																					
<i>Centrolophus niger</i>	1	P																					
<i>Emmopterus lucifer</i>	1	P																					
<i>Hoplostethus mediterraneus</i>	1	F																					
	1	P																					
	2	F																					
<i>Lepidorhynchus denticulatus</i>	3	P																					
<i>Malacocephalus laevis</i>	1	P																					
	1	P																					
	2	P																					
	3	F																					
<i>Maurolicus muelleri</i>	1	F																					



**Table A4a: New Zealand mid slope (> 700 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤ 10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Coelenterates	Tunicates	Polychaetes	Molluscs	Crustaceans	Echinoderms	Osteichthyes	Reference
<i>Alepocephalus australis</i>	1	SW Chatham Rise	26			F								Clark et al 1989
<i>Allocyttus niger</i>	1	SW Chatham Rise	329			F								Clark et al 1989
<i>Caelorinchus fasciatus</i>	1	SW Chatham Rise	21			F								Clark et al 1989
<i>Coryphaenoides subsermilatus</i>	1	SW Chatham Rise	23			F								Clark & King 1989
<i>Deania calcea</i>	1	North Island	63			F								Clark et al 1989
<i>Diatobranchius capensis</i>	1	SW Chatham Rise	10			F								Clark et al 1989
<i>Emtopterus baxteri</i>	1	SW Chatham Rise	117			F								Clark et al 1989
<i>Hoplostethus atlanticus</i>	1	Chatham Rise	33			N								Rosecchi et al 1988
	2	Challenger - March	210	~42		F								Rosecchi et al 1988
	3	Challenger - July	392	~60		F								Rosecchi et al 1988
	4	Challenger - Nov.	85	~38		F								Rosecchi et al 1988
	5	Challenger - Total	687	9-40		N								Rosecchi et al 1988
<i>Macrourus carinatus</i>	1	SW Chatham Rise	34			F								Clark et al 1989
<i>Pseudocyttus maculatus</i>	1	SW Chatham Rise	242			F								Clark et al 1989



**Table A4a (continued)**

	Study no.	Method	Molluscs	Cephalopods	Decapods	Octopods	Crustaceans	Other crustaceans	Ostracods	Copepods	Euphausiids	Natant decapods	Mysids	Isopods	Amphipods	Echinoderms	Brittle stars	Osteichthyes	Fish remains	Larval fishes	Merlucciidae	Mycetophidae	Oreosomatidae
<i>Alepocephalus australis</i>	1	F																					
<i>Allocyttus niger</i>	1	F																					
<i>Caelorinchus fasciatus</i>	1	F																					
<i>Coryphaenoides subserrulatus</i>	1	F																					
<i>Deania calcea</i>	1	F																					
<i>Diastrorhynchus capensis</i>	1	F																					
<i>Eimopterus baxteri</i>	1	F																					
<i>Hoplostethus atlanticus</i>	1	N																					
	2	F																					
	3	F																					
	4	F																					
	5	N																					
<i>Macrourus carinatus</i>	1	F																					
<i>Pseudocyttus maculatus</i>	1	F																					

**Table A4b: Southern Hemisphere mid slope (> 700 m), n, numbers of stomachs examined; P, presence of a food item in the stomach; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Species	Study no.	Study area	No. Stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Chondrichthyes	Osteichthyes	Cetacean remains	Reference
<i>Allocyttus verrucosus</i>	1	S Africa	1802	10-42	F													Mel'nikov 1981
<i>Anoplogaster cornuta</i>	2	SE Australia	3173	10-38	F													Lyle & Smith 1997
<i>Bathysaurus ferrox</i>	1	W Africa	1	0	P													Daveport 1993
<i>Caelorinchus fasciatus</i>	1	NW Africa	714	15-29	P													Marshall & Merrett 1977
	1	Namibia	61	small	F													Macpherson 1985
	2	S Africa	110	26	F													Meyer & Smaale 1991b
	3	S Africa	110	26	F													Meyer & Smaale 1991b
<i>Centrophorus squamosus</i>	1	NW Africa	1	121	P													Marshall & Merrett 1977
	2	S Africa	21	14	F													Ebert et al. 1992
<i>Centroscymnus coelolepis</i>	1	NW Africa	4	43-131	F													Marshall & Merrett 1977
	2	S Africa	93	66-102	P													Ebert et al. 1992
<i>C. crepidater</i>	1	S Africa	6	32-110	F													Ebert et al. 1992
<i>Chaunax pictus</i>	1	NW Africa	5	5-21	P													Marshall & Merrett 1977
<i>Cubiceps baxteri</i>	1	WC S America, EC Aust.	40	3	F													Gorelova et al 1994
	2	WC S America, EC Aust.	50	8	F													Gorelova et al 1994
	3	WC S America, S Africa	56	50	F													Gorelova et al 1994
	4	WC S America, S Africa	8	16-23	F													Gorelova et al 1994
<i>Deania calcea</i>	1	NW Africa	64	66-94	P													Marshall & Merrett 1977
	2	E T asmania	61	42-105	F													Blaber & Bulman 1987
	3	Namibia	81	56-108	IRI													Yano 1991 *
	4	S Africa	2	7	F													Ebert et al. 1992
<i>Diaphus danae</i>	1	Southern Africa	538	3-12	F													Lipinski et al. 1992*
<i>Epigonus lenimen</i>	1	E T asmania	597	40	F													Young & Blaber 1986
<i>Emmopterus pusillus</i>	1	E T asmania	5	10-19	F													Blaber & Bulman 1987
	2	S Africa	7	39-46	F													Ebert et al. 1992
<i>Halosauruspis macrochir</i>	1	NW Africa	5	15-27	P													Bass et al. 1976*
	1	NW Africa	5	15-27	P													Marshall & Merrett 1977

**Table A4b (continued)**

Study no.	Study area	No. Stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Chondrichthyes	Osteichthyes	Cetacean remains	Reference
1	SE Aust - 88 juv	3157	57	<30	W												Bulman & Koslow 1992
2	SE Aust - 88 adults	1416	51	>30	W												Bulman & Koslow 1992
3	SE Aust - 89 juv	1929	65	<30	W												Bulman & Koslow 1992
4	SE Aust - 89 adults	984	68	>30	W												Bulman & Koslow 1992
1	S Africa	54	39	small	F												Meyer & Smale 1991b
2	S Africa	14	39	large	F												Meyer & Smale 1991b
1	E Tasmania	165	61	24-37	F												Blaber & Bulman 1987
2	SE Australia	1427	96	9-41	F												Lyle & Smith 1997

Table A4b (continued)

Species	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Crustaceans	Crustacean remains	Ostracods	Copepods	Stomatopods	Euphausiids	Decapods	Natant decapods	Anomurans	Thalassinids	Hermit crabs	Mysids	Iso-pods	Amphipods	Echinoderms	Brittle stars
<i>Allocytus verrucosus</i>	1	F	█			█	█														█	
<i>Anoplogaster cornuta</i>	2	F				█																
<i>Bathysaurus ferox</i>	1	P					P															
<i>Caelorinchus fasciatus</i>	1	P					P															
	2	F					█	█													█	
	3	F					█	█													█	
<i>Centrophorus squamosus</i>	1	P																				
	2	F					█															
<i>Centroscyms coelelepis</i>	1	P																				
	2	F					█															
<i>C. crepidater</i>	1	F	█																			
<i>Chaunax pictus</i>	1	P					P															
<i>Cubiceps baxteri</i>	1	F	█	█			█															
	2	F	█	█			█															
	3	F	█	█			█															
	4	F	█	█			█															
<i>Deania calcea</i>	1	P																				
	2	F					█															
	3	IRI																				
	4	F																				
	5	F																				
<i>Diaphus danae</i>	1	F	█				█															
<i>Epigonus lenimen</i>	1	F	█				█															
<i>Eimopterus pusillus</i>	1	F	█				█															
	2	F	█				█															
<i>Halosauruspis macrochir</i>	1	P																				
















Table A4b (continued)

Species	Study no.	Method	Chondrichthyes	Elasmobranchii	Squalidae	Osteichthyes	Fish remains	Other fish	Callionymidae	Carangidae	Chauliodontidae	Dirtemidae	Macrouridae	Melanostomidae	Mertuicidae	Mycophidae	<i>Diaphus</i> spp.	<i>Lampanyctodes hectoris</i>	Nemichthyidae	Ophichthidae	
<i>Allocyttus verrucosus</i>	1	F				Diagonal lines															
<i>Anoplogaster cornuta</i>	2	F				P															
<i>Bathysaurus ferox</i>	1	P				P															
<i>Caelorhynchus fasciatus</i>	1	P				P															
	2	F				Diagonal lines												Diagonal lines			
	3	F				Diagonal lines												Diagonal lines			
<i>Centrophorus squamosus</i>	1	P				Diagonal lines															
	2	F				Diagonal lines															
<i>Centroscymnus coelepis</i>	1	P				P															
	2	F				Diagonal lines															
<i>C. crepidater</i>	1	F				Diagonal lines															
<i>Chaunax pictus</i>	1	P				Diagonal lines															
<i>Cubiceps baxteri</i>	1	F				Diagonal lines															
	2	F				Diagonal lines															
	3	F				Diagonal lines															
	4	F				Diagonal lines															
<i>Deania calcea</i>	1	P				P															
	2	F				Diagonal lines															
	3	IRI				Diagonal lines															
	4	F				Diagonal lines															
	5	F				Diagonal lines															
<i>Diaphus danae</i>	1	F				Diagonal lines															
<i>Epigonus lenini</i>	1	F				Diagonal lines															
<i>Etmopterus pusillus</i>	1	F				Diagonal lines															
	2	F				Diagonal lines															
<i>Halosaurus macrochir</i>	1	P				Diagonal lines															

**Table A4b (continued)**

	Study no.	Method	Chondrichthyes	Elasmobranchii	Squalidae	Osteichthyes	Fish remains	Other fish	Callionymidae	Carangidae	Chauliodontidae	Diretmidae	Macrouridae	Melanostomidae	Mertuicidae	Mycophidae	<i>Diphus</i> sp.	<i>Lampanyctodes hectoris</i>	Nemichthyidae	Ophichthidae	
<i>Hoplostethus atlanticus</i>	1	W				█	█														
	2	W				█	█				█										
	3	W				█	█				█										
	4	W				█	█				█										
<i>Malacocephalus laevis</i>	1	F				█	█														
	2	F				█	█														
<i>Neocyttus rhomboidalis</i>	1	F				█	█														
	2	F				█	█														

**Table A4c: Northern Hemisphere mid slope (> 700 m). n, numbers of stomachs examined; P, presence of a food item in the stomach; , ≤10% importance in diet; , 11 to 50 % importance in diet; , >51 % importance in diet.**

Species	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Undertified	Extraneous matter	Foraminiferans	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Chondrichthyes	Osteichthyes	Cetacean remains	Reference
<i>Alepisaurus ferox</i>	1	NW Pacific	73	1		N														Kubota 1977
	2	C Pacific	42			F														Moteki et al. 1992
	3	C Pacific	24			F														Moteki et al. 1992
<i>Antimora rostrata</i>	1	W Atlantic	10	30	29-50	P								P						Sedberry & Musick 1978
	2	NE Atlantic	153	89	8-46	F														Mauchline & Gordon 1984c
<i>Aristomias spp.</i>	1	C Pacific	17	71	33-140	I														Clarke 1982
<i>Astronesthes indicus</i>	1	C Pacific	154	75	2-6	I														Clarke 1982
	2	C Pacific	18	83	6-15	I														Clarke 1982
<i>A. splendidus</i>	1	C Pacific	61	0	2-10	I														Clarke 1982
<i>Bathysaurus ferox</i>	1	SE Atlantic	1	0	50	P														Marshall & Merrett 1977
	2	W Atlantic	14	50	17-61	F														Sedberry & Musick 1978
	3	W Atlantic	30	53		F														Sulak et al. 1985
<i>Caelorinchus innotabilis</i>	1		3		<28	P								P						McLellan 1977
<i>Centrophorus squamosus</i>	1	SE Atlantic	1		121	P														Marshall & Merrett 1977
	2	NE Atlantic	18	50	111-148	F														Mauchline & Gordon 1983a
	3	NE Atlantic	1			F														Lipinski et al. 1992*
<i>Centroscyllium coelelepis</i>	1	NE Atlantic	12		67-106	F														Clarke & Merrett 1972
	2	SE Atlantic	4		66-102	P														Marshall & Merrett 1977
	3	W Atlantic	5	40	37-50	F														Sedberry & Musick 1978
	4	NE Atlantic	18	72	88-149	F														Mauchline & Gordon 1983a
	5	NW Pacific	64	~80	64-109	P								P						Yano & Tanaka 1984
	6	Mediterranean	69	36	19-39	F														Carrasson et al. 1992
	7	Mediterranean	17	41	40-75	F														Carrasson et al. 1992
	8	NE Atlantic	2			F														Lipinski et al. 1992*
<i>C. crepidater</i>	1	NE Atlantic	23	48	26-79	F														Mauchline & Gordon 1983a
<i>C. owstoni</i>	1	NW Pacific	336	~80	38-117	P														Yano & Tanaka 1984



**Table A4c (continued)**

Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Foraminiferans	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Chondrichthyes	Osteichthyes	Cetacean remains	Reference
1	NE Atlantic	151	56	2-26	I														Clarke 1982
2	NE Atlantic	2	100	5-22	F														Mauchline & Gordon 1983b
1	NW Pacific	139	73	118-177	F														Kubota et al 1991
1	NE Atlantic	1	0	35	P														Minchin 1988
1	Mediterranean	4			F														Capape 1975*
2	Mediterranean	31			F														Maepherson 1980*
3	Mediterranean	31			W														Maepherson 1981
4	Mediterranean	97	18	23-101	F														Matallanas 1982
1	NE Atlantic	2		99-147	F														Clarke & Merrett 1972
2	SE Atlantic	8		66-94	P														Marshall & Merrett 1977
3	NE Atlantic	65	69	79-98	F														Mauchline & Gordon 1983a
4	NE Atlantic	46	59	99-111	F														Mauchline & Gordon 1983a
1	NE Atlantic	40	88	27-60	F														Du Buit 1978
2	Mediterranean	311		5-19	W														Maepherson 1981
3	NE Atlantic	525	31	8-34	F														Mauchline & Gordon 1984a
4	NW Atlantic				P														Vinnichenko 1997
1	W Pacific	347	48	2-22	N														Gorelova 1981
2	C Pacific	77		3-21	I														Clarke 1982
3	NE Atlantic	1	0	16	F														Mauchline & Gordon 1983b
4	Gulf of Mexico	114		3-5	N														Lancraft et al. 1988
5	Gulf of Mexico	121		8-10	N														Lancraft et al. 1988
6	Gulf of Mexico	31		11-13	N														Lancraft et al. 1988
1	NE Atlantic	573	94	8-27	F														Mauchline & Gordon 1984c
1	SE Atlantic	5		15-27	P														Marshall & Merrett 1977
2	W Atlantic	83	19	13-38	F														Sedberry & Musick 1978

**Table A4c (continued)**

	Study no.	Study area	No. stomachs	No. empty (%)	Fish length (cm)	Method	Unidentified	Extraneous matter	Foraminiferans	Coelenterates	Tunicates	Polychaetes	Chaetognaths	Molluscs	Crustaceans	Echinoderms	Chondrichthyes	Osteichthyes	Cetacean remains
<i>Harriotta raleighana</i>	1	W Atlantic	10	0	25-91	P	P												
	2	NE Atlantic	10	20		F													
<i>Hoplostethus atlanticus</i>	1	NE Atlantic	53	28	14-30	F													
<i>Idiacanthus fasciola</i>	1	C Pacific	291	82	5-38	I													
<i>Lampanyctus alatus</i>	1	Gulf of Mexico	84			N													
<i>L. nobilis</i>	1	C Pacific	183		4-9	I													
<i>Malacosteus niger</i>	1	C Pacific	100	80	2-19	I												P	
<i>Mora moro</i>	1	Mediterranean	93	97		P												P	
<i>Odontomacrus murrayi</i>	1	N Atlantic	1		50	P												P	
<i>Photostomias, 2 spp.</i>	1	C Pacific	189	74	3-14	I													
<i>Raja hyperborea</i>	1	NE Atlantic	31	16	23-84	F													
<i>Scopelogadus beanii</i>	1	W Atlantic	106	29	4-11	F													
<i>Simencheilus parasiticus</i>	1	W Atlantic	23	0	14-46	P													
<i>Stomias boa ferox</i>	1	NE Atlantic	8	100	3-28	F													
<i>Xenodermichthys copei</i>	1	NE Atlantic - pelagic	22	23	1-14	F													
	2	NE Atlantic - demersal	36	22	5-11	F													
	3	NE Atlantic - demersal	62	47	11-14	F													
	4	NE Atlantic - demersal	64	42	14-17	F													

**Table A4c (continued)**

	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Crustaceans	Crustacean remains	Larval crustaceans	Ostracods	Copepods	Euphausiids	Decapod remains	Natant decapods	Isopods	Amphipods	Chondrichthyes	Elasmobranchii	Holocephali	Osteichthyes	Fish remains	Fish lens	Larval fishes	Other fish
<i>Alepisaurus ferox</i>	1	N																						
	2	F																						
<i>Antimora rostrata</i>	3	F																						
	1	P																						
	2	F																						
<i>Aristomias spp.</i>	1	I																						
<i>Astronesthes indicus</i>	1	I																						
	2	I																						
<i>A. splendidus</i>	1	I																						
<i>Bathysaurus ferox</i>	1	P																						
	2	F																						
	3	F																						
<i>C. innotabilis</i>	1	P																						
<i>Centrophorus squamosus</i>	1	P																						
	2	F																						
	3	F																						
<i>Centroscyllium coelelepis</i>	1	F																						
	2	P																						
	3	F																						
	4	F																						
	5	P																						
	6	F																						
	7	F																						
	8	F																						
<i>C. crepidater</i>	1	F																						
<i>C. owstoni</i>	1	P																						

**Table A4c (continued)**

Species	Study no.	Method	Molluscs	Gastropods	Cephalopods	Decapods	Crustaceans	Crustacean remains	Larval crustaceans	Ostracods	Copepods	Euphausiids	Decapods	Decapod remains	Natant decapods	Mysids	Euphausiids / mysids	Mysid remains	Isopods	Amphipods	Echinoderms	Chondrichthyes	Elasmobranchii	Emopteridae	Scyliorhinidae
<i>Chaulioidus sloani</i>	1	I																							
<i>Chlamydoselachus anguineus</i>	2	F																							
<i>Cryptosaras couesi</i>	1	F																							
<i>Dalatias licha</i>	1	P																							
	1	F																							
	2	F																							
	3	W																							
	4	F																							
<i>Deania calcea</i>	1	F																							
	2	P																							
	3	F																							
	4	F																							
<i>Epigonus telescopus</i>	1	F																							
	2	W																							
	3	F																							
	4	F																							
<i>Gonostoma elongatum</i>	1	N																							
	2	I																							
	3	F																							
	4	N																							
	5	N																							
	6	N																							
<i>Halargyreus johnsonii</i>	1	F																							
<i>Halosaurus macrochir</i>	1	P																							
	2	F																							

**Table A4c (continued)**

Species	Study no.	Method	Molluscs	Gastropods	Cephalopods	Crustaceans	Crustacean remains	Larval crustaceans	Ostracods	Copepods	Euphausiids	Decapods	Decapod remains	Natant decapods	<i>Munda</i> sp.	Mysids	Euphausiids / mysids	Mysid remains	Iso-pods	Amphipods	Osteichthyes	Fish remains	Fish scales	Other fish	Mycetophidae	<i>Lamppanyctus</i> spp.
<i>Harriotta raleighana</i>	1	P	P			P																				
<i>Hoplostethus atlanticus</i>	2	F																								
<i>Idiacanthus fasciola</i>	1	F																								
<i>Lamppanyctus alatus</i>	1	I																								
<i>L. nobilis</i>	1	N																								
<i>Malacosteus niger</i>	1	I																								
<i>Mora moro</i>	1	I																								
<i>Odontomacrus murrayi</i>	1	P																								
<i>Photostomias</i> , 2 spp.	1	P																								
<i>Raja hyperborea</i>	1	I																								
<i>Scopelogadus beanii</i>	1	F																								
<i>Simenechelys parasiticus</i>	1	F																								
<i>Stomias boa ferrox</i>	1	F																								
<i>Xenoderm ichthys copei</i>	1	F																								
	2	F																								
	3	F																								
	4	F																								

**Table A4c (continued)**

Species	Study no.	Method	Alepisauridae	Anoplogastridae	Argentinidae	Chauliodontidae	Chiasmodontidae	Congridae	Engraulidae	Gadidae	Gempylidae	Gonostomatidae	Halosauridae	Macrouridae	Melanocetidae	Moridae	Mycophphidae	Nomidae	Omosudae	Paralepididae	Scombridae	Scorpaenidae	Sternopychidae	Synphobranchidae	Tachipteridae
<i>Alepisaurus ferox</i>	1	N																							
	2	F																							
<i>Antimora rostrata</i>	3	F																							
	1	P																							
	2	F																							
<i>Aristomias</i> spp.	1	I																							
<i>Astronesthes indicus</i>	1	I																							
	2	I																							
<i>A. splendidus</i>	1	I																							
<i>Bathysaurus ferox</i>	1	P																							
	2	F																							
	3	F																							
<i>C. innotabilis</i>	1	P																							
<i>Centrophorus squamosus</i>	1	P																							
	2	F																							
	3	F																							
<i>Centroscymnus coelelepis</i>	1	F																							
	2	P																							
	3	F																							
	4	F																							
	5	P																							
	6	F																							
	7	F																							
	8	F																							
<i>C. crepidater</i>	1	F																							
<i>C. owstoni</i>	1	P																							

**Table A4c (continued)**

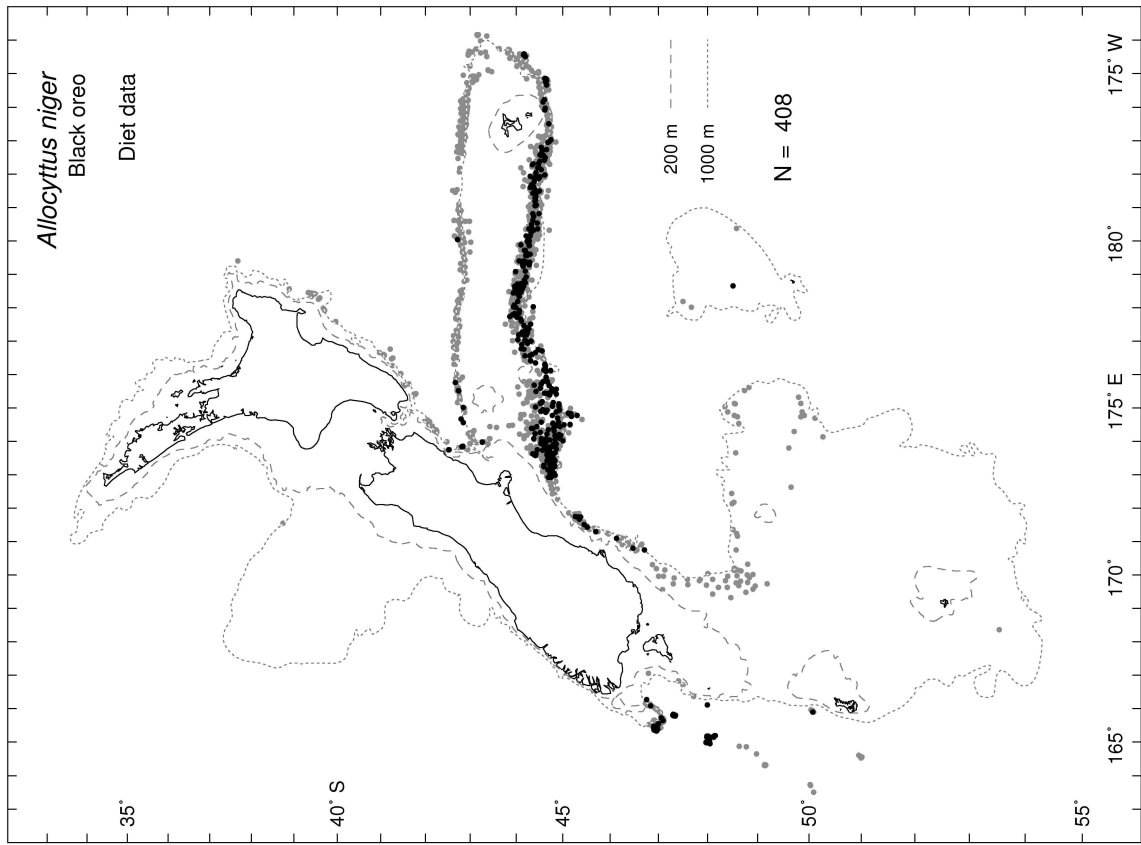
Species	Study no.	Method	Osteichthyes	Fish remains	Fish scales	Fish lens	Fish otolith	Larval fish	Other fish	Argentiniidae	Chauliodontidae	Gadidae	Gonostomatidae	Macrouridae	Moridae	Mycophidae	Lampancyctus spp.	Paralepididae	Scombridae	Scorpaenidae	Sternopychidae	
<i>Chauliodus sloani</i>	1	I	■	▨					▨													
<i>Chlamydoselachus anguineus</i>	2	F																				
<i>Cryptosaras couesi</i>	1	F																		P		
<i>Dalatias licha</i>	1	P	■																			
	1	F	■																			
	2	F	■																			
	3	W	▨																			
	4	F	■	▨																		
<i>Deania calcea</i>	1	F	■	■																		
	2	P	■																			
	3	F	▨	▨																		
	4	F	▨	▨																		
<i>Epigonus telescopus</i>	1	F	▨	▨																		
	2	W	▨																			
	3	F	▨	▨																		
	4	P	▨																			
<i>Gonostoma elongatum</i>	1	N	▨	▨																		
	2	I	▨																			
	3	F	▨																			
	4	N	▨	▨																		
	5	N	▨	▨																		
	6	N	▨	▨																		
<i>Halargyreus johnsonii</i>	1	F	▨	▨																		
<i>Halosaurus macrochir</i>	1	P	▨																			
	2	F	▨	▨																		



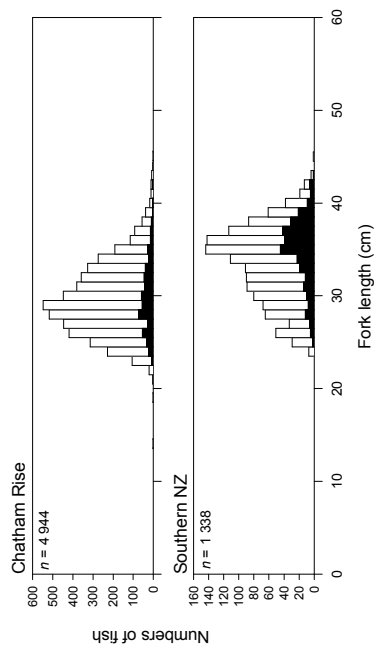


**APPENDIX B: Fish species catch and stomach content sampling distribution maps,  
and ontological diet change (MFish Research Trawl Database 1960-2000).**

**APPENDIX B: Fish species catch and stomach content sampling distribution maps, and ontological diet change (MFish Research Trawl Database 1960-2000).**



**Figure B1a.** The distribution of all black oreo (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.



**Figure B1b.** The length frequency of black oreo where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

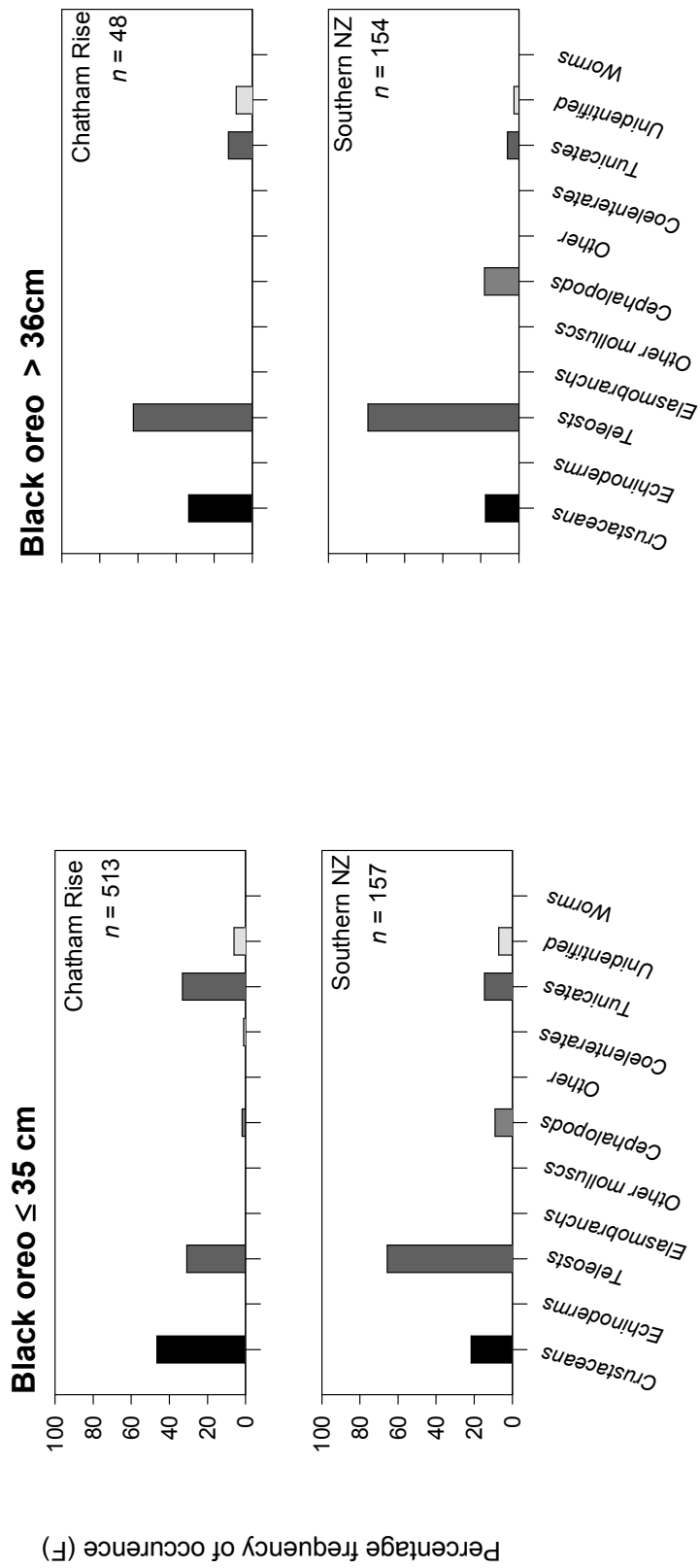


Figure B1c. The importance of major prey groups in the diet of black oreo examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

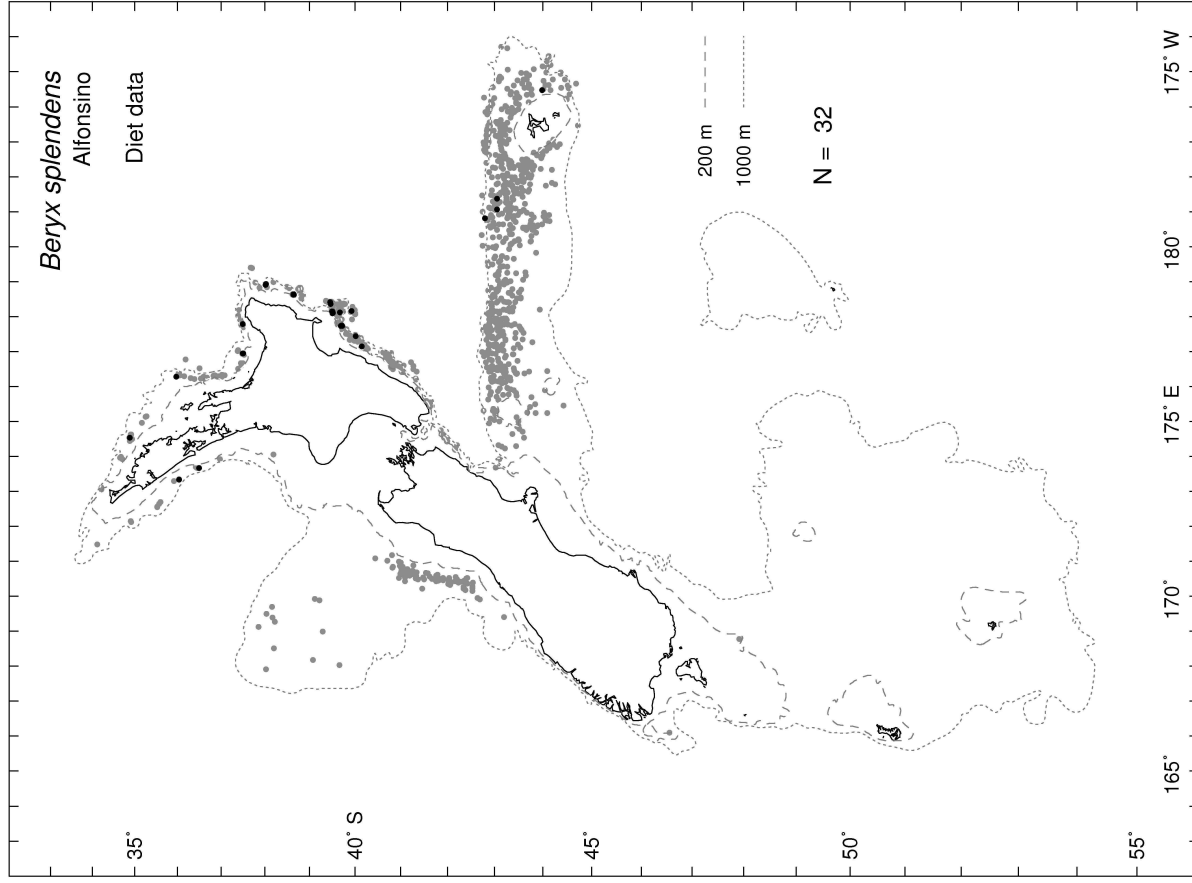


Figure B2a. The distribution of all alfonsino (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

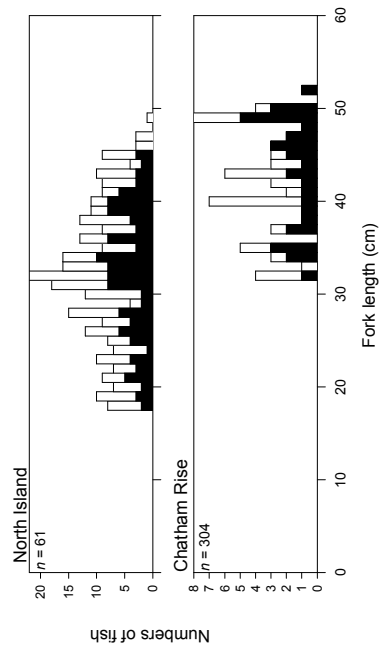


Figure B2b. The length frequency of alfonsino where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

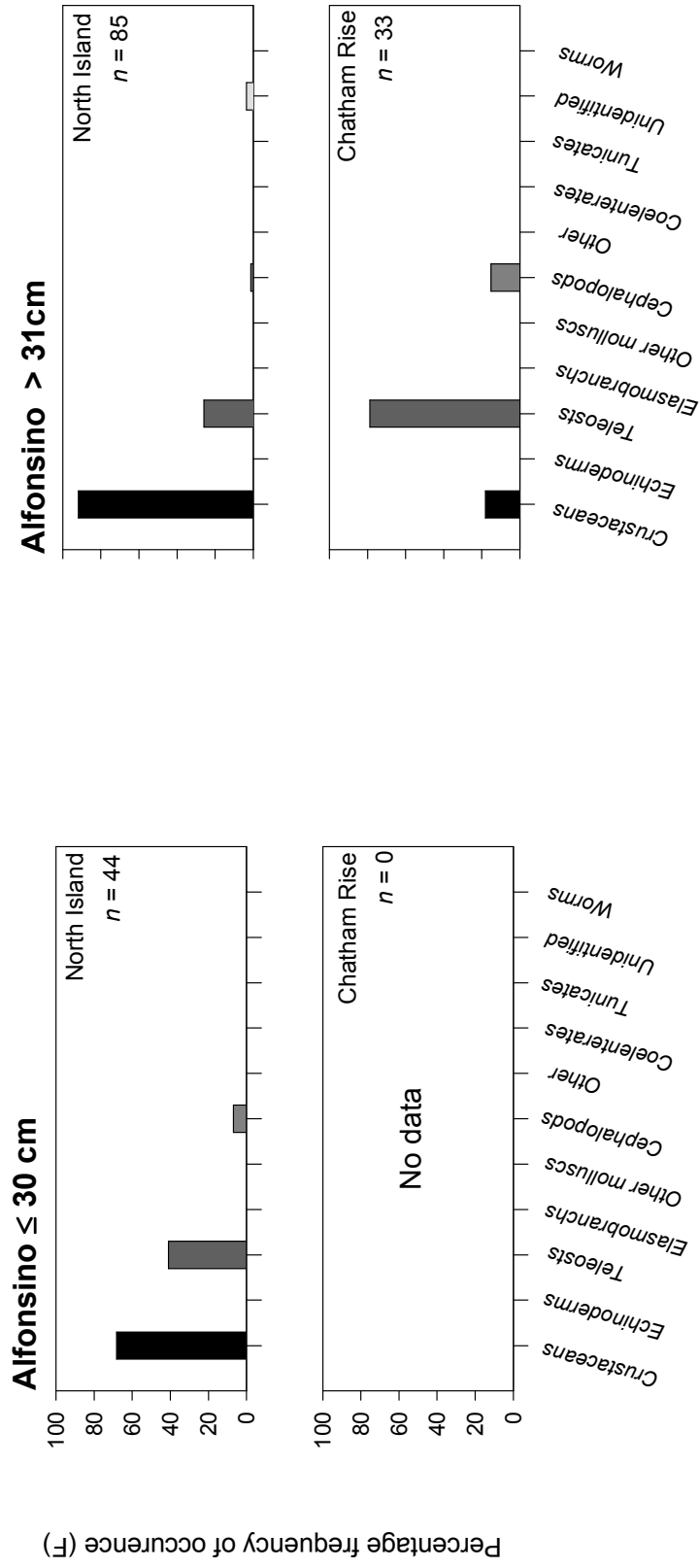


Figure B2c. The importance of major prey groups in the diet of alfonsino examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9.  $n$ , number of fish examined for diet.

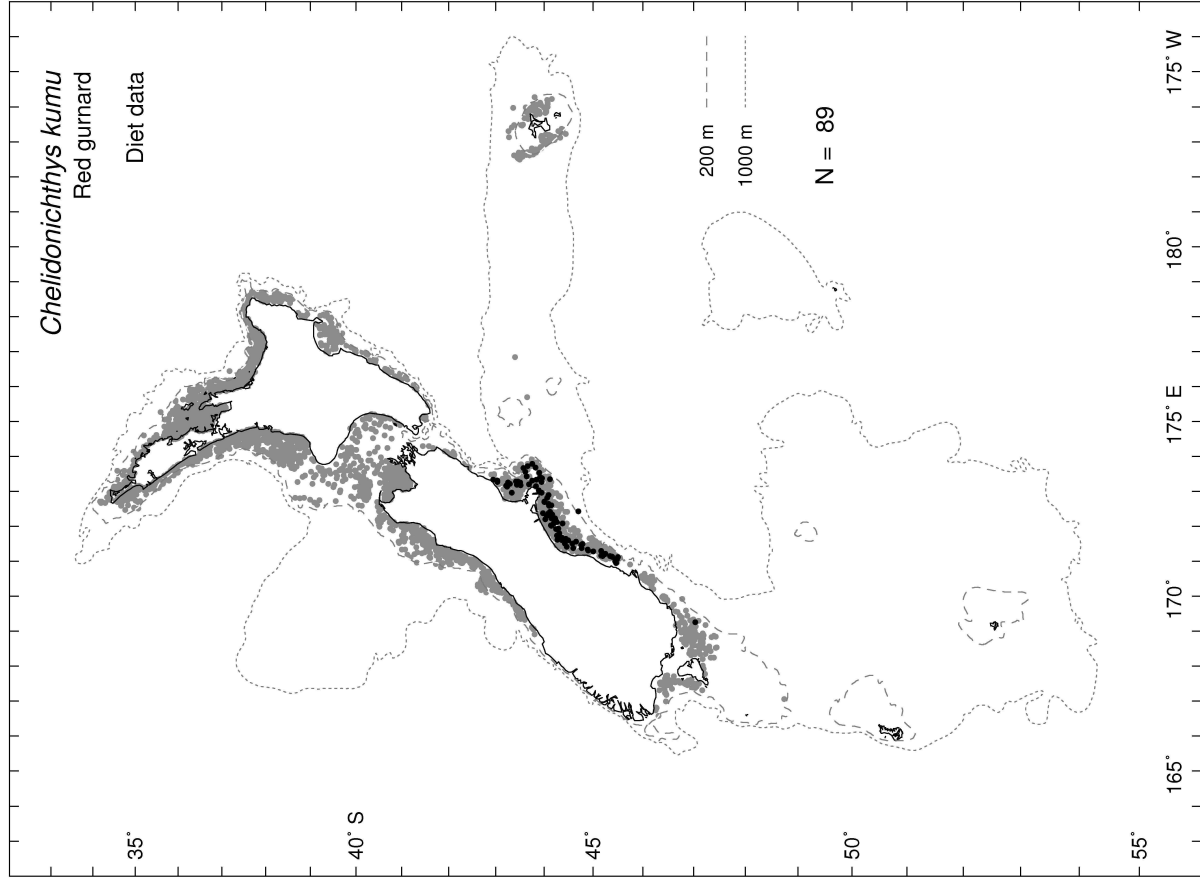


Figure B3a. The distribution of all red gurnard (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

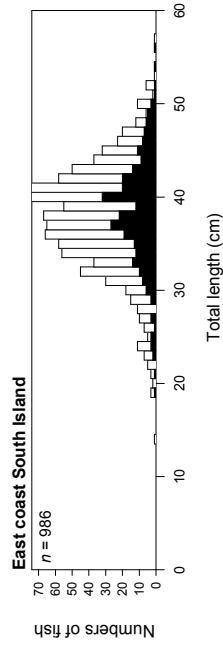


Figure B3b. The length frequency of red gurnard where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

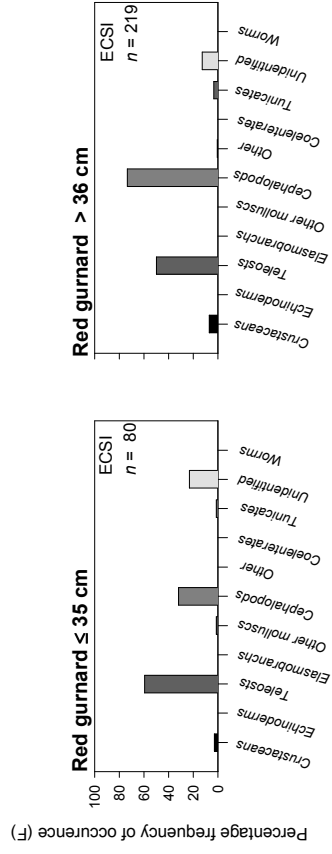


Figure B3c. The importance of major prey groups in the diet of red gurnard examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.



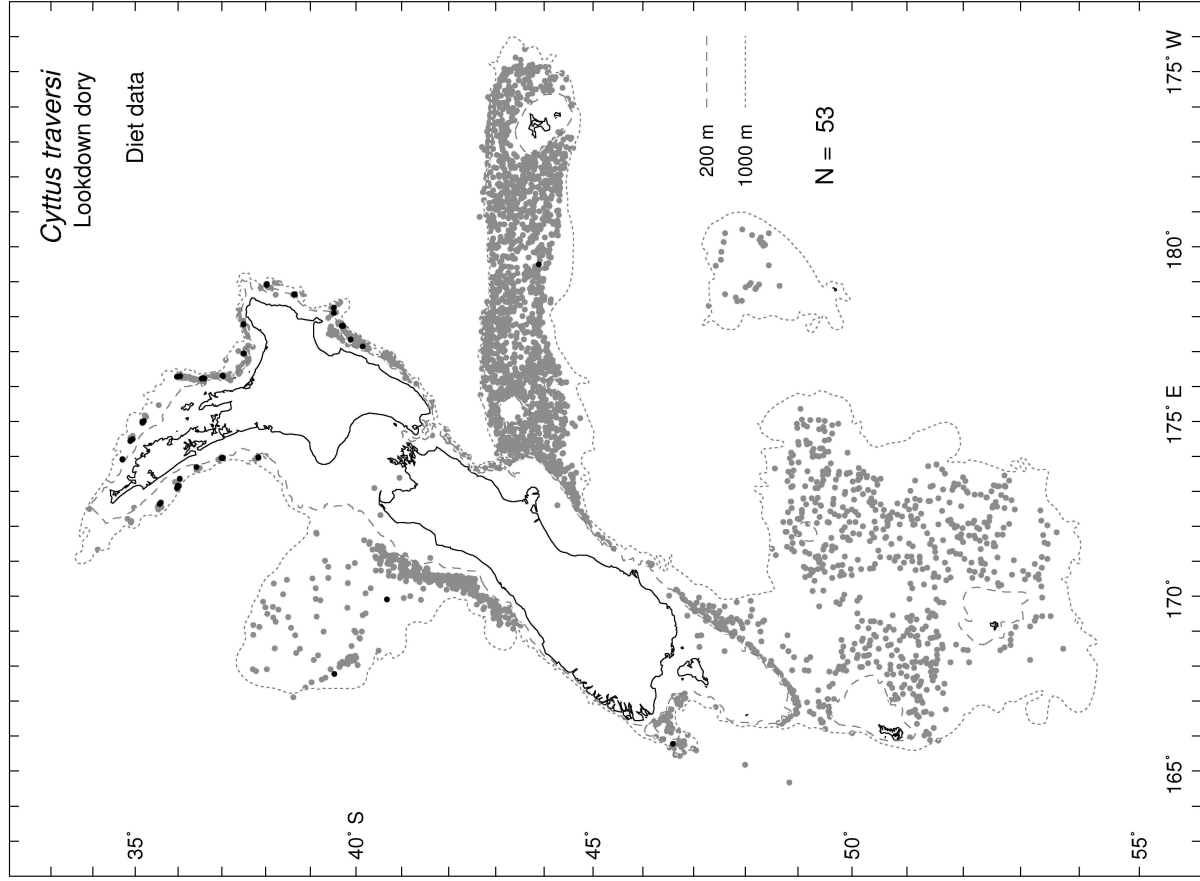


Figure B4a. The distribution of all lookdown dory (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

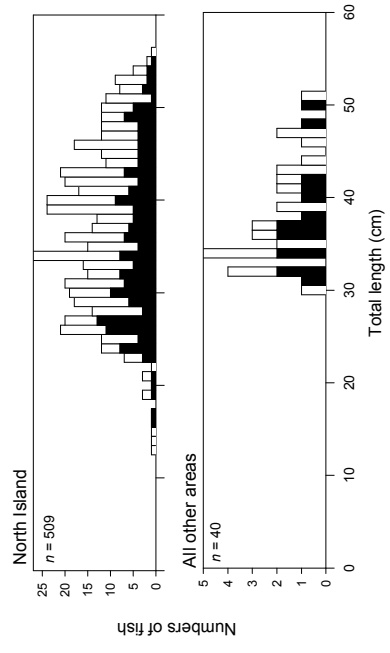


Figure B4b. The length frequency of lookdown dory where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.



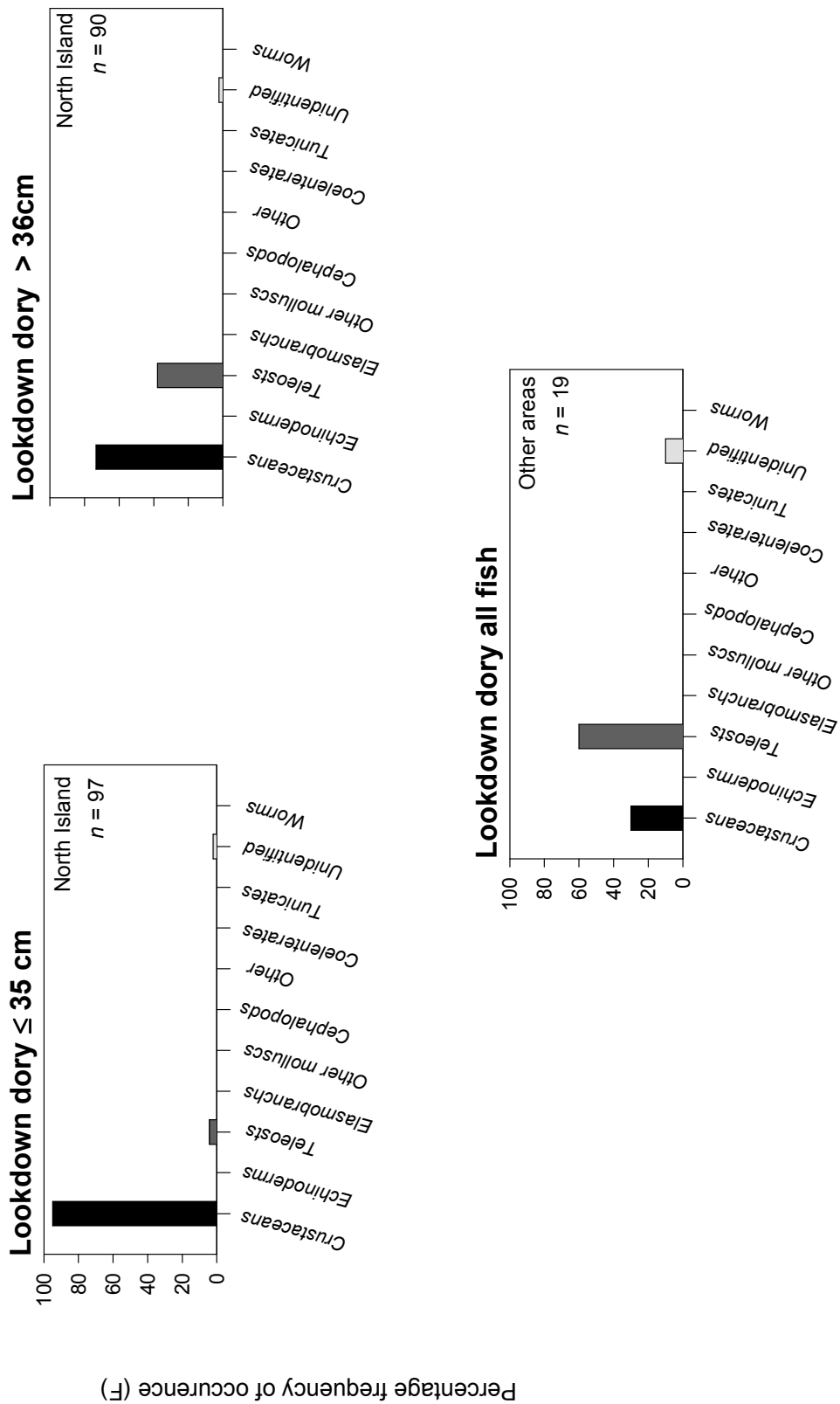


Figure B4c. The importance of major prey groups in the diet of lookdown dory examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

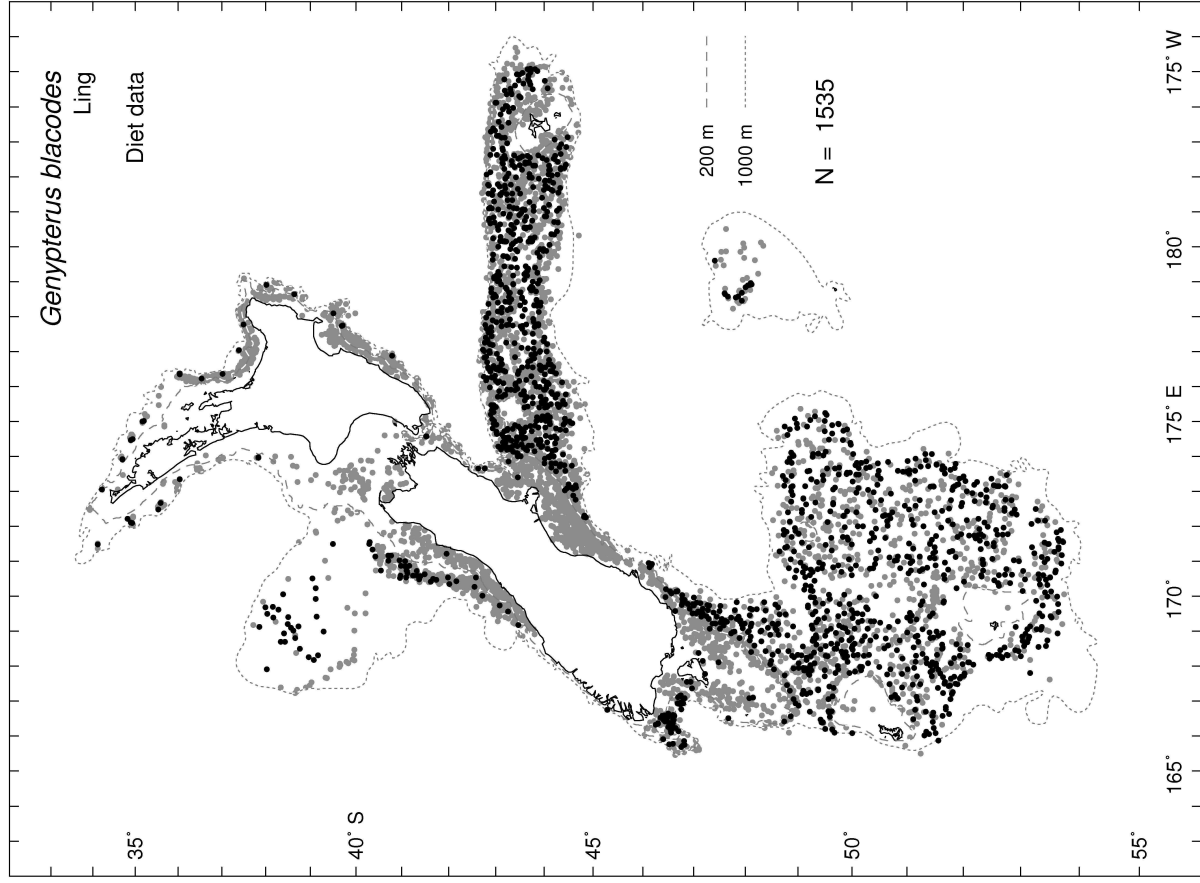


Figure B5a. The distribution of all ling (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

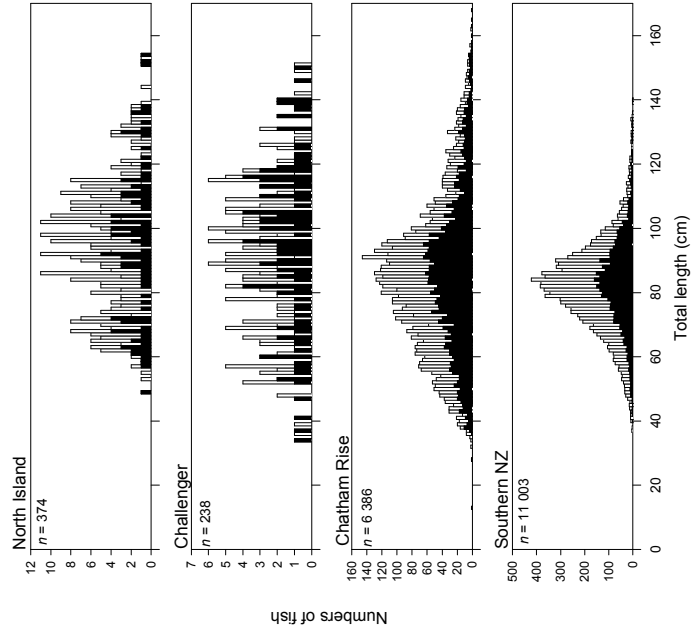


Figure B5b. The length frequency of ling where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

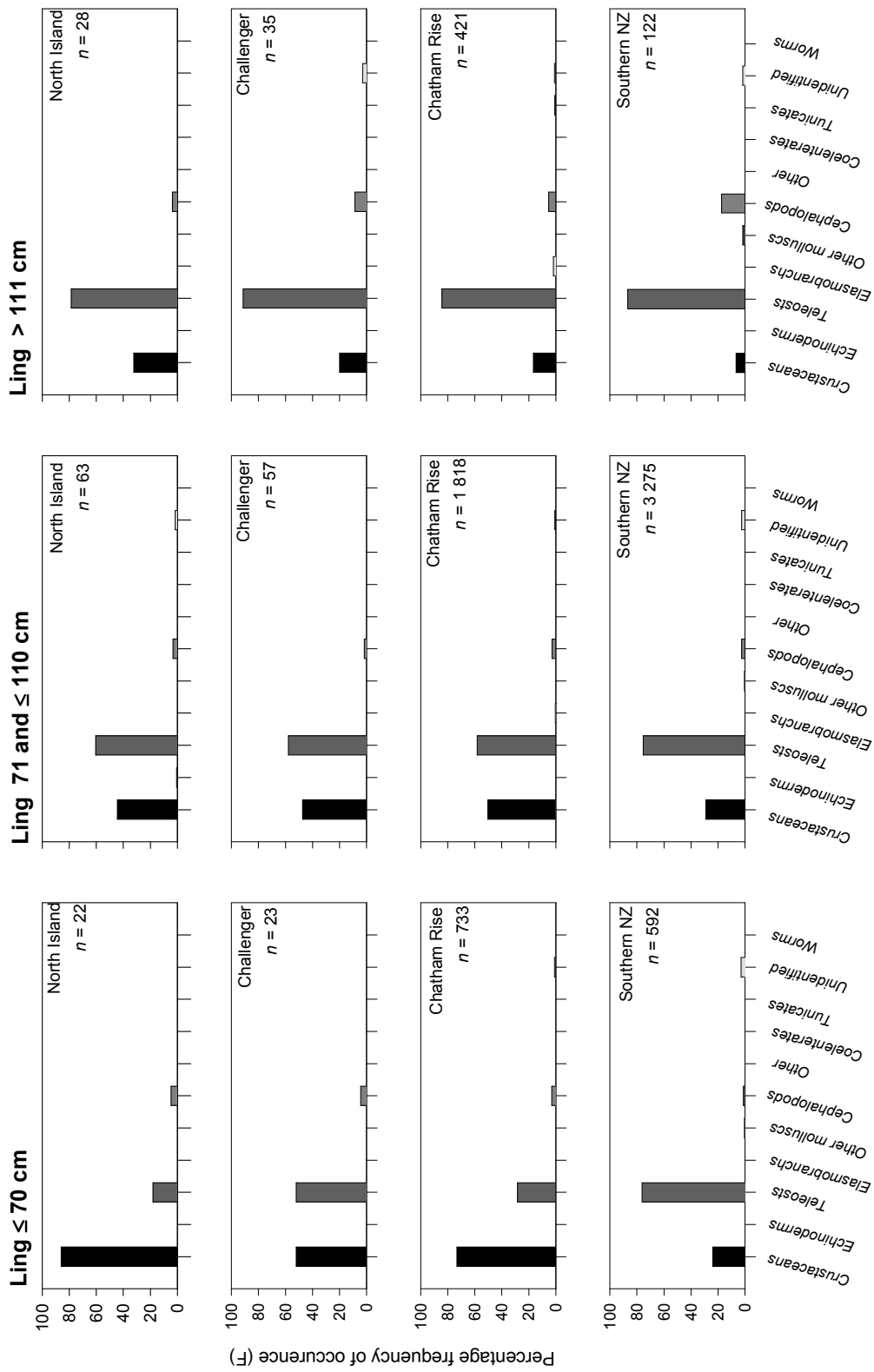


Figure B5c. The importance of major prey groups in the diet of ling examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

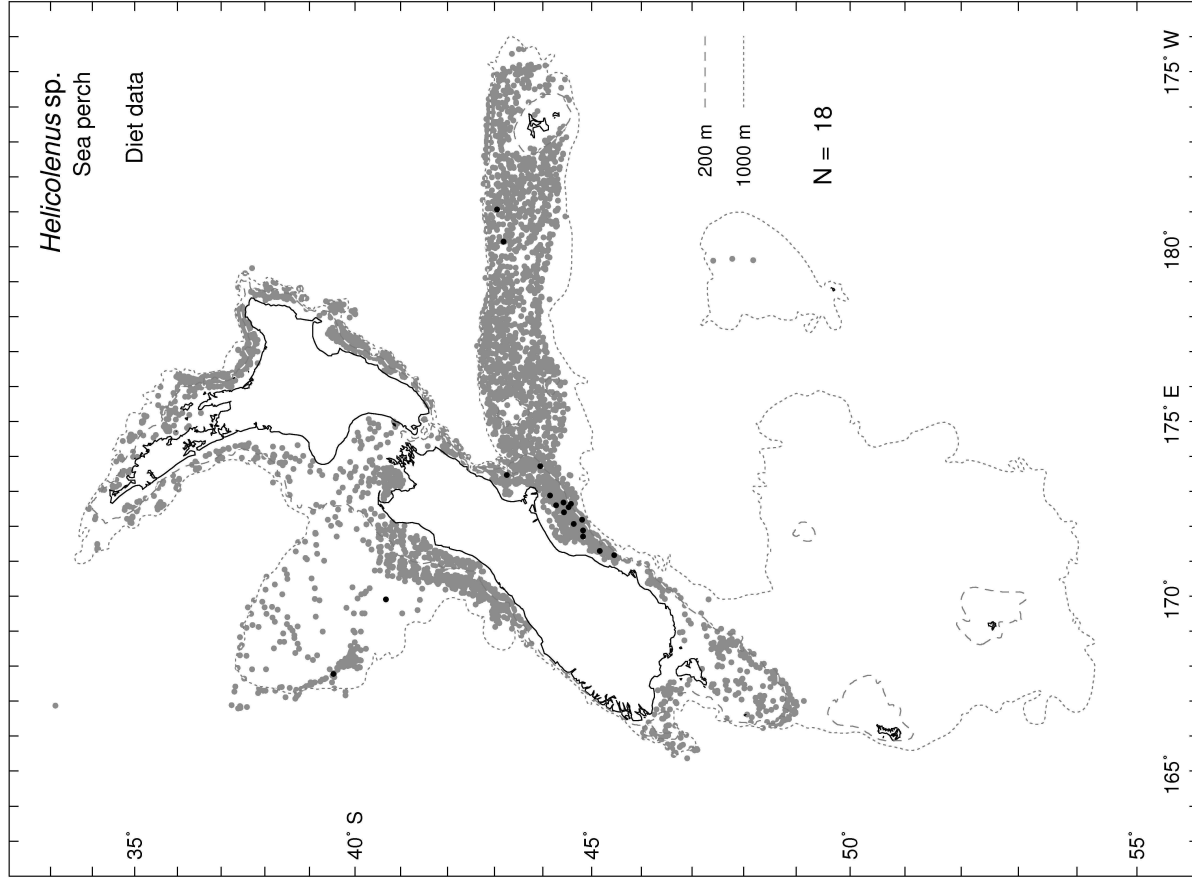


Figure B6a. The distribution of all sea perch (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

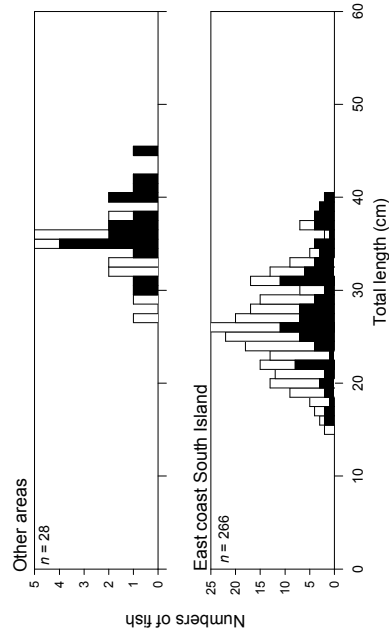


Figure B6b. The length frequency of sea perch where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. *n*, number of fish examined for diet.

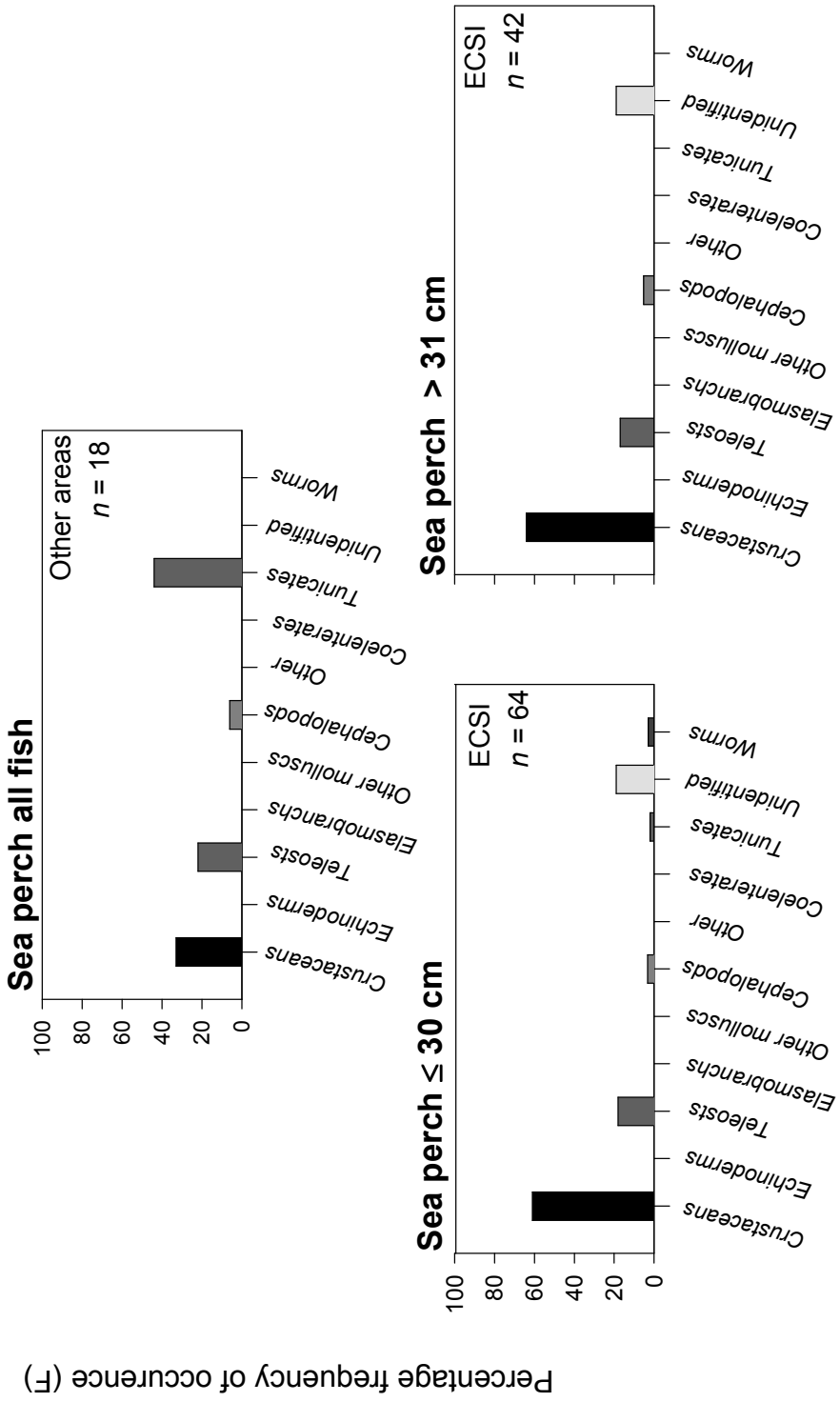


Figure B6c. The importance of major prey groups in the diet of sea perch examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

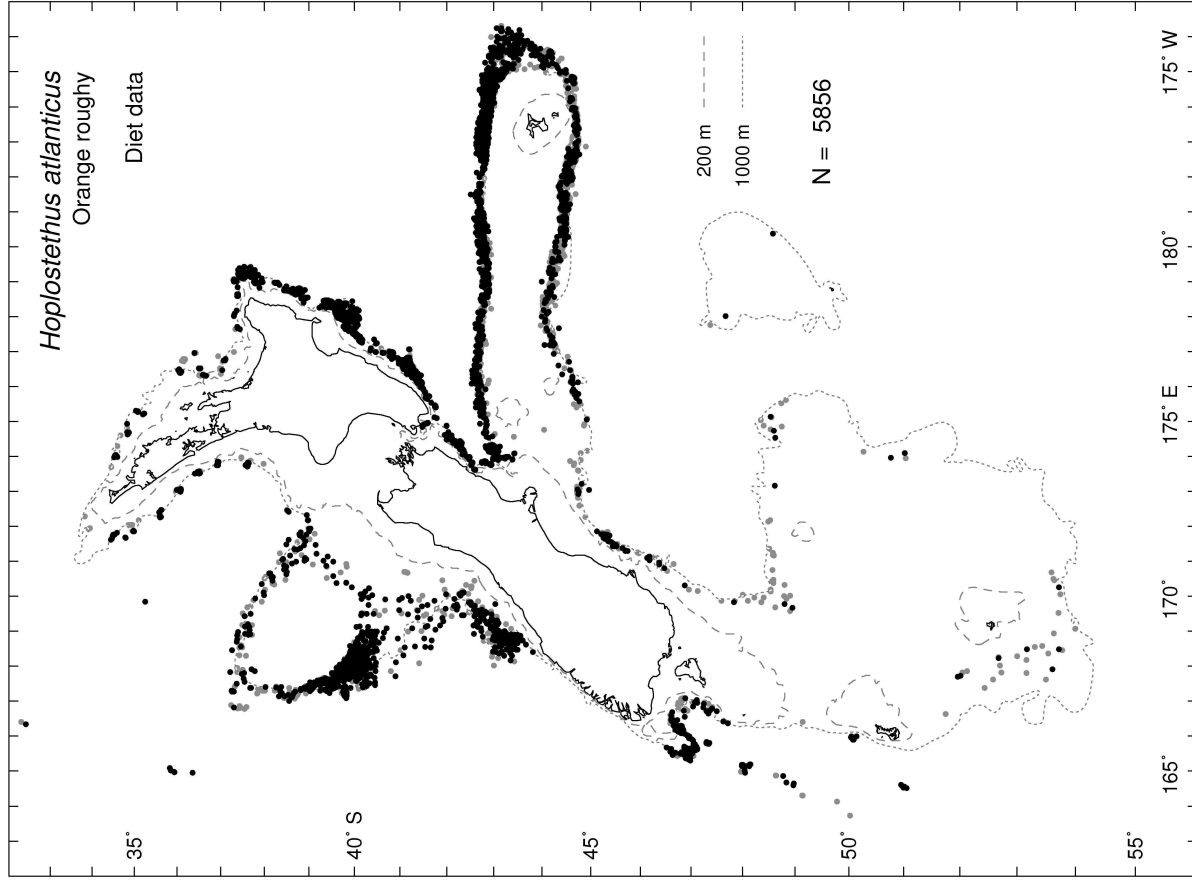


Figure B7a. The distribution of all orange roughy (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

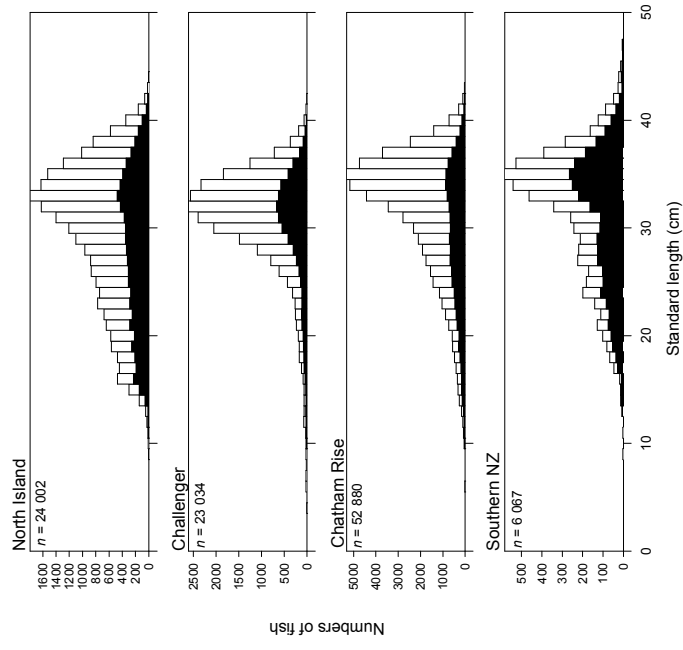


Figure B7b. The length frequency of orange roughy where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

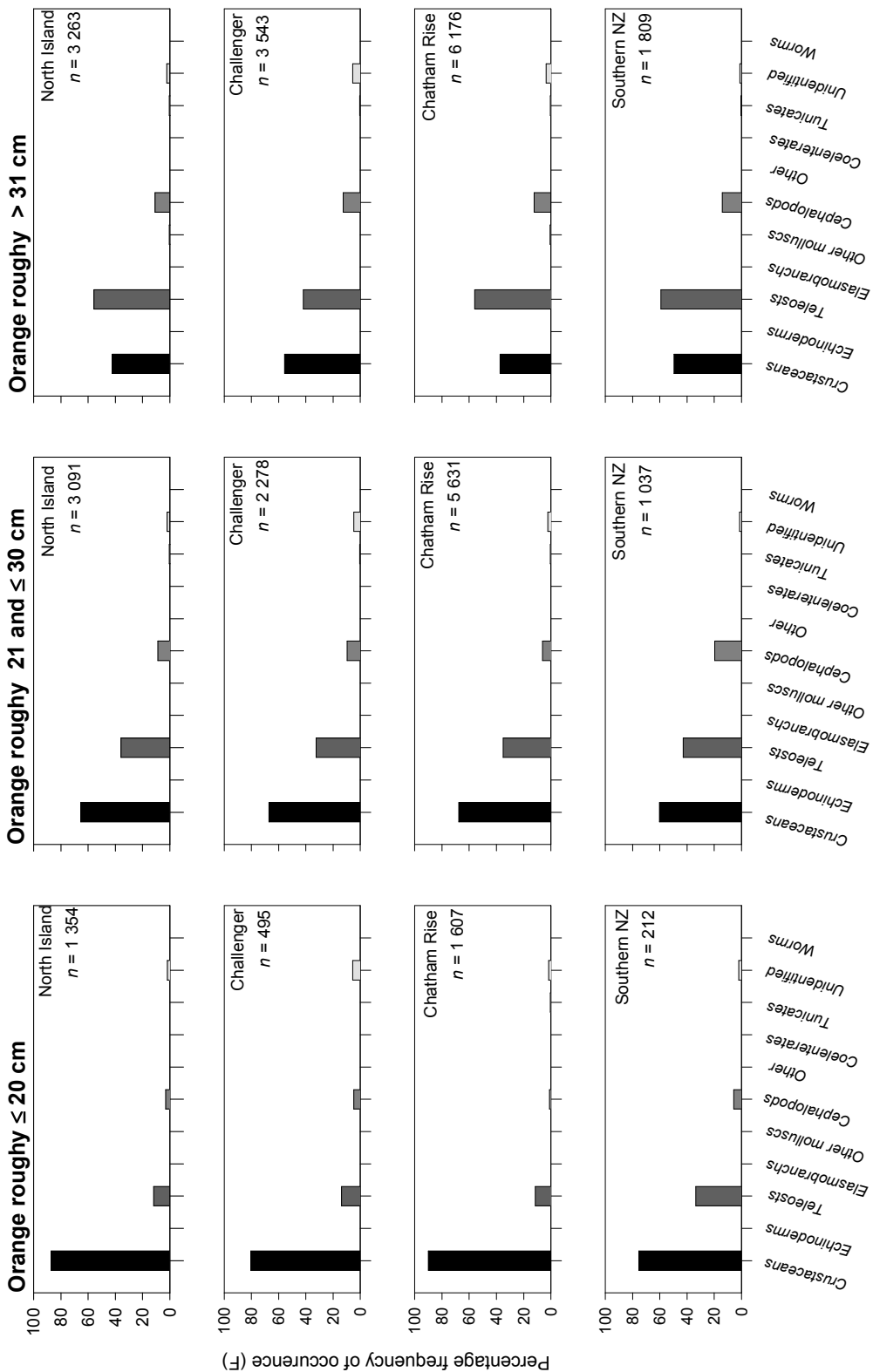


Figure B7c. The importance of major prey groups in the diet of orange roughy examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

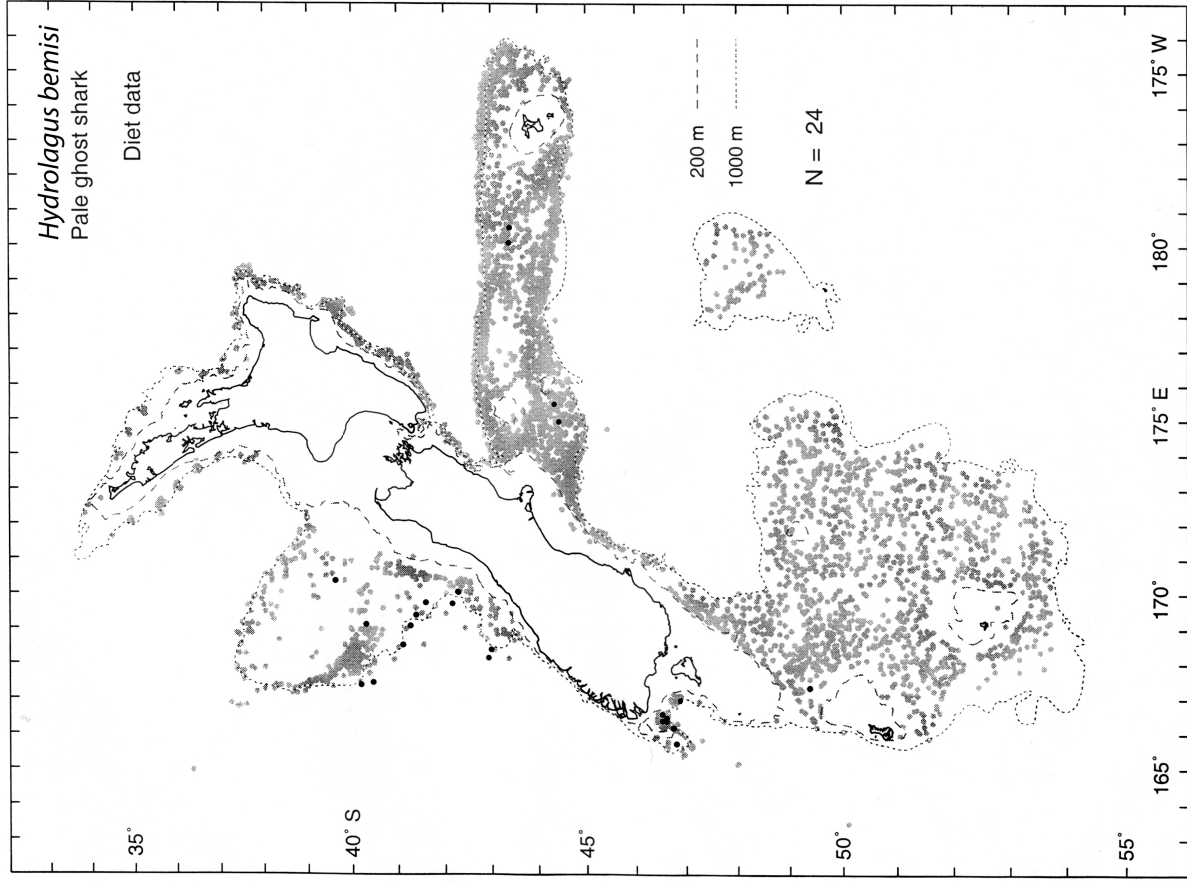
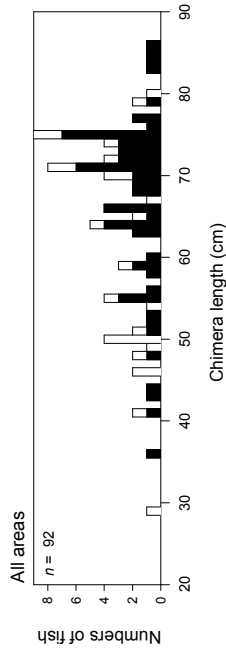


Figure B8a. The distribution of all pale ghost shark (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.



Note: Challenger sample not included, appears to be total length not chimeria length

Figure B8b. The length frequency of pale ghost shark where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

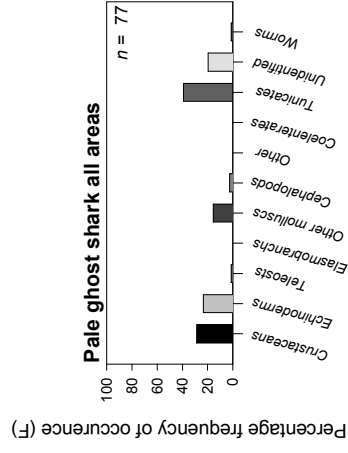


Figure B8c. The importance of major prey groups in the diet of pale ghost shark examined on research trawl surveys. Fish size groups are arbitrary designations. n, number of fish examined for diet.





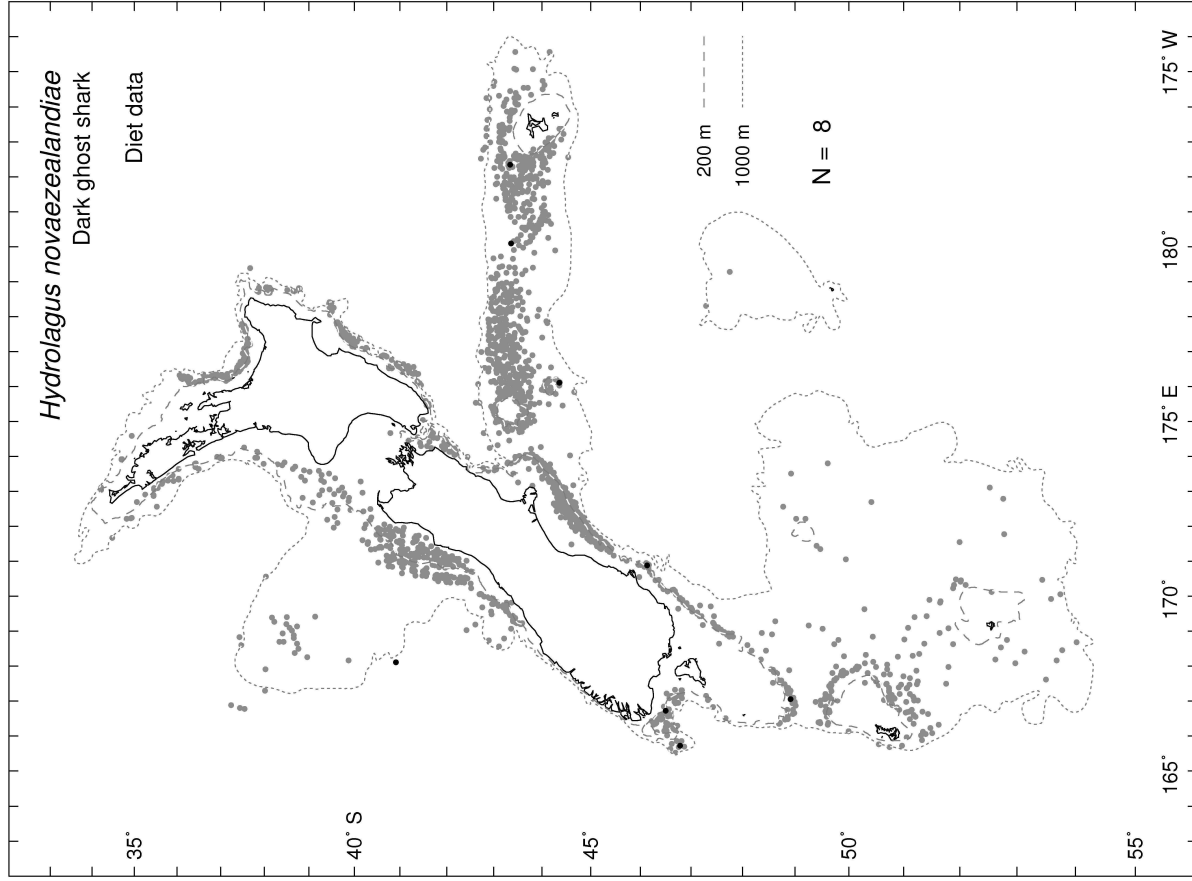


Figure B9a. The distribution of all dark ghost shark (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

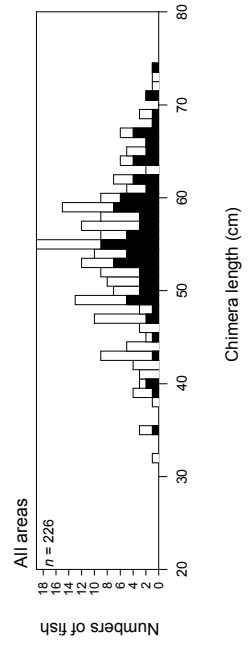


Figure B9b. The length frequency of dark ghost shark where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

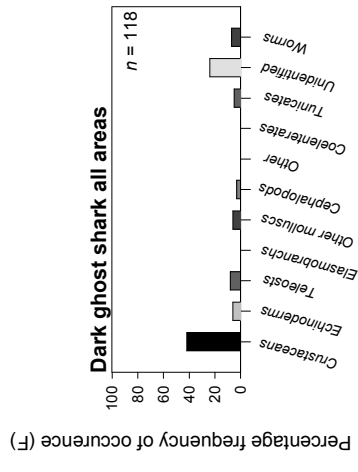


Figure B9c. The importance of major prey groups in the diet of dark ghost shark examined on research trawl surveys. Fish size groups are arbitrary designations. n, number of fish examined for diet.



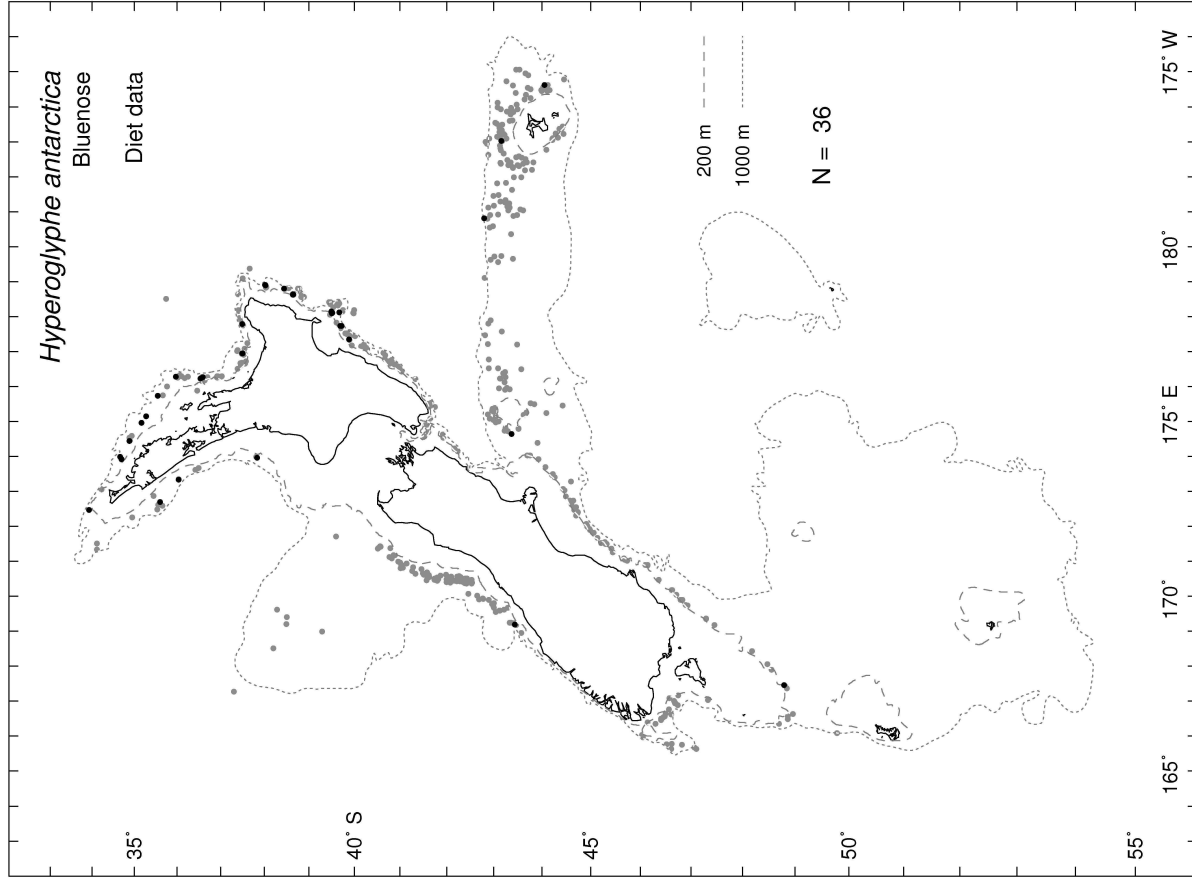


Figure B10a. The distribution of all bluenose (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

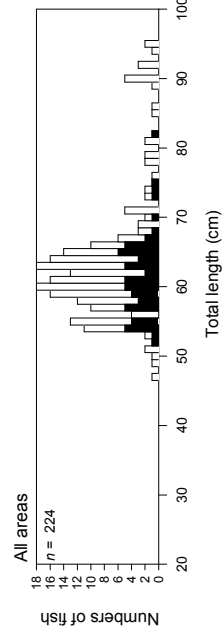


Figure B10b. The length frequency of bluenose where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

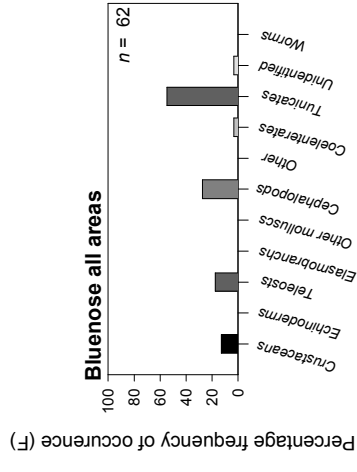


Figure B10c. The importance of major prey groups in the diet of bluenose examined on research trawl surveys. Fish size groups are arbitrary designations. n, number of fish examined for diet.



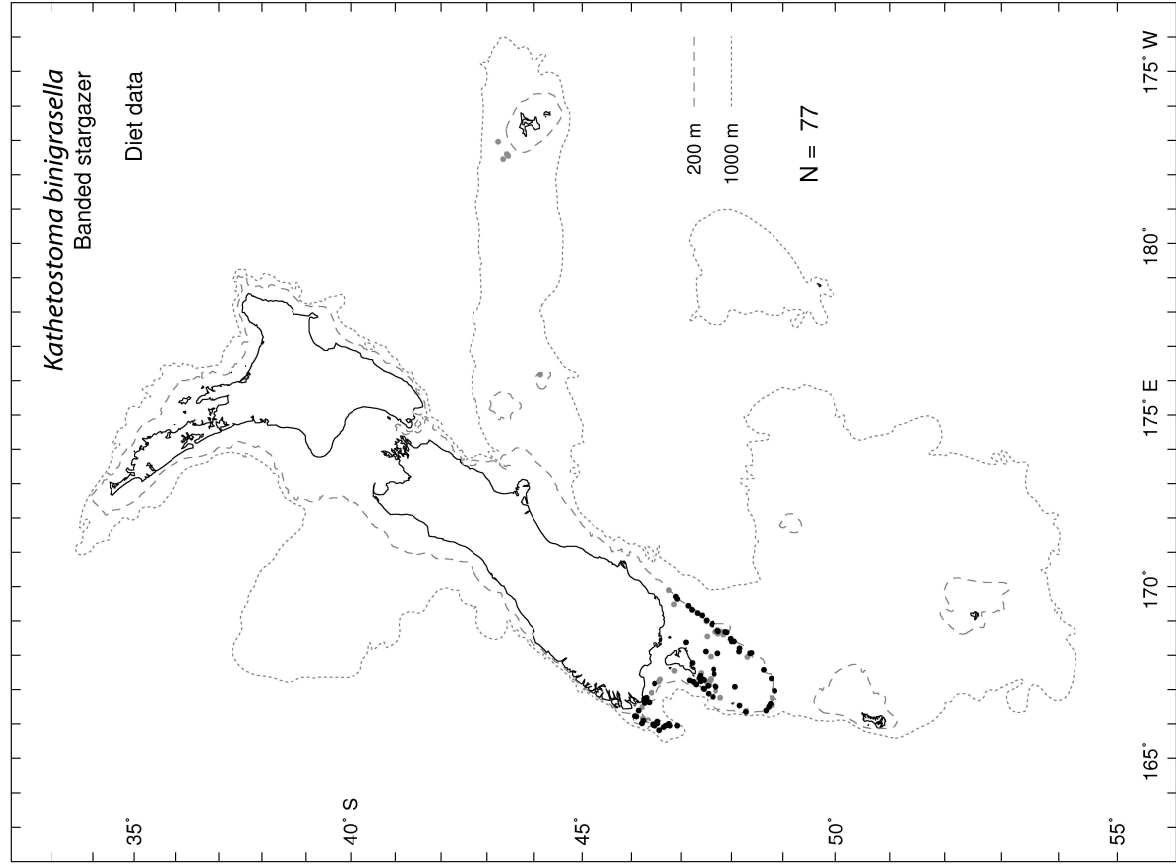


Figure B11a. The distribution of all banded stargazer (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

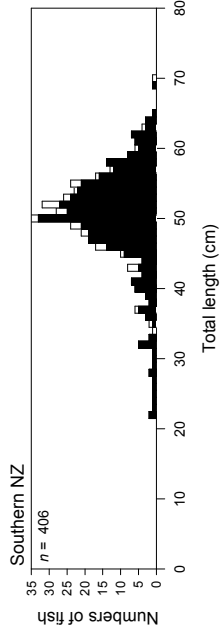


Figure B11b. The length frequency of banded stargazer where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

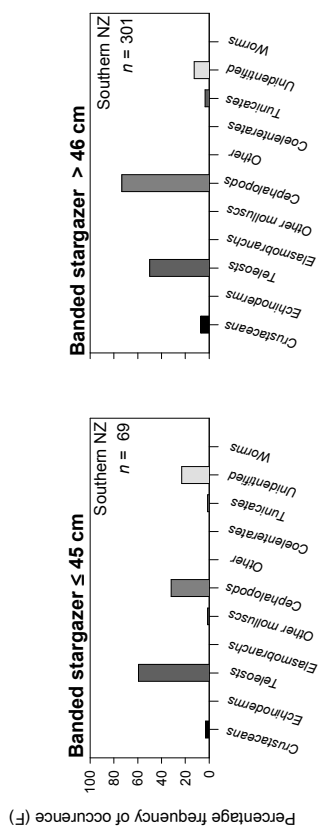


Figure B11c. The importance of major prey groups in the diet of banded stargazer examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.



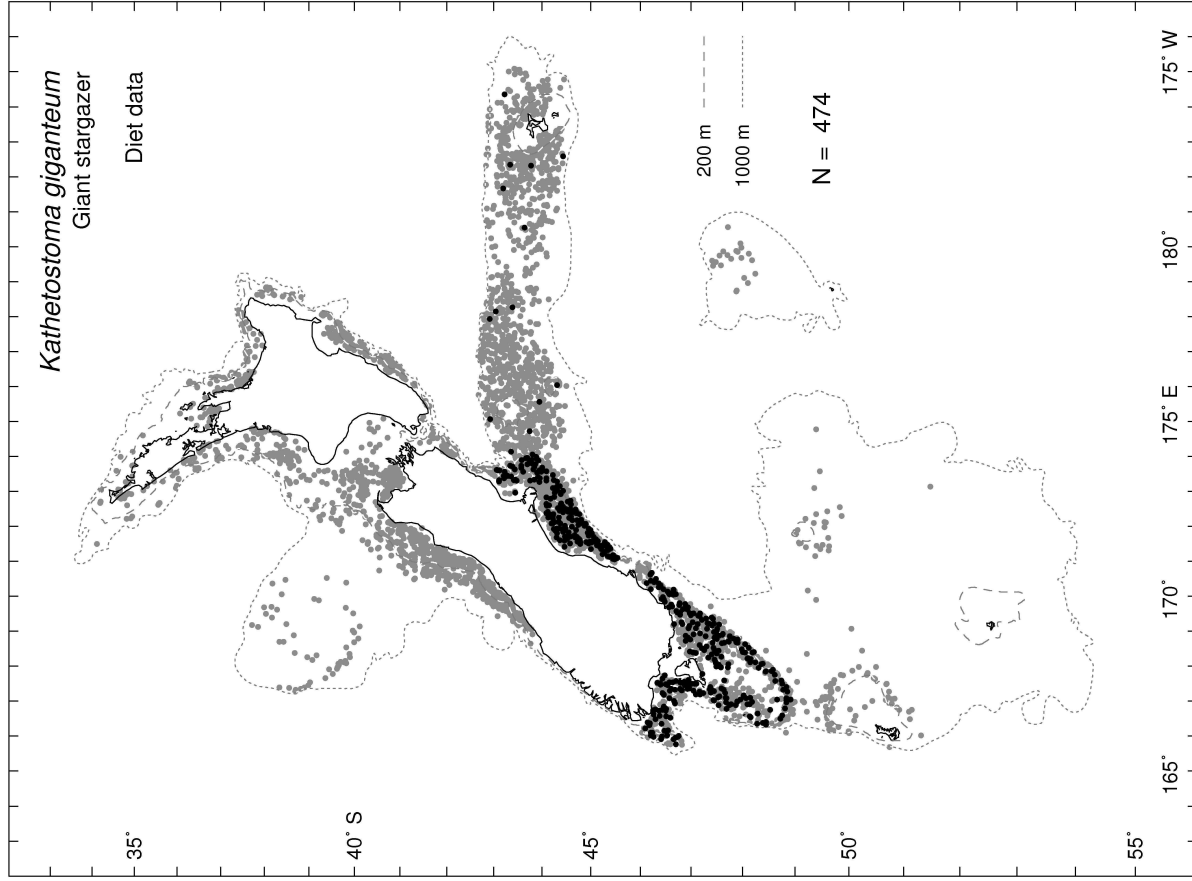


Figure B12a. The distribution of all giant stargazer (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

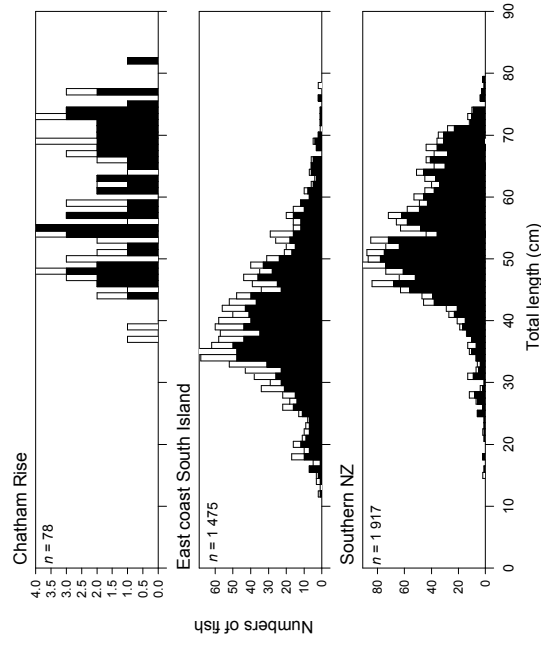


Figure B12b. The length frequency of giant stargazer where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.



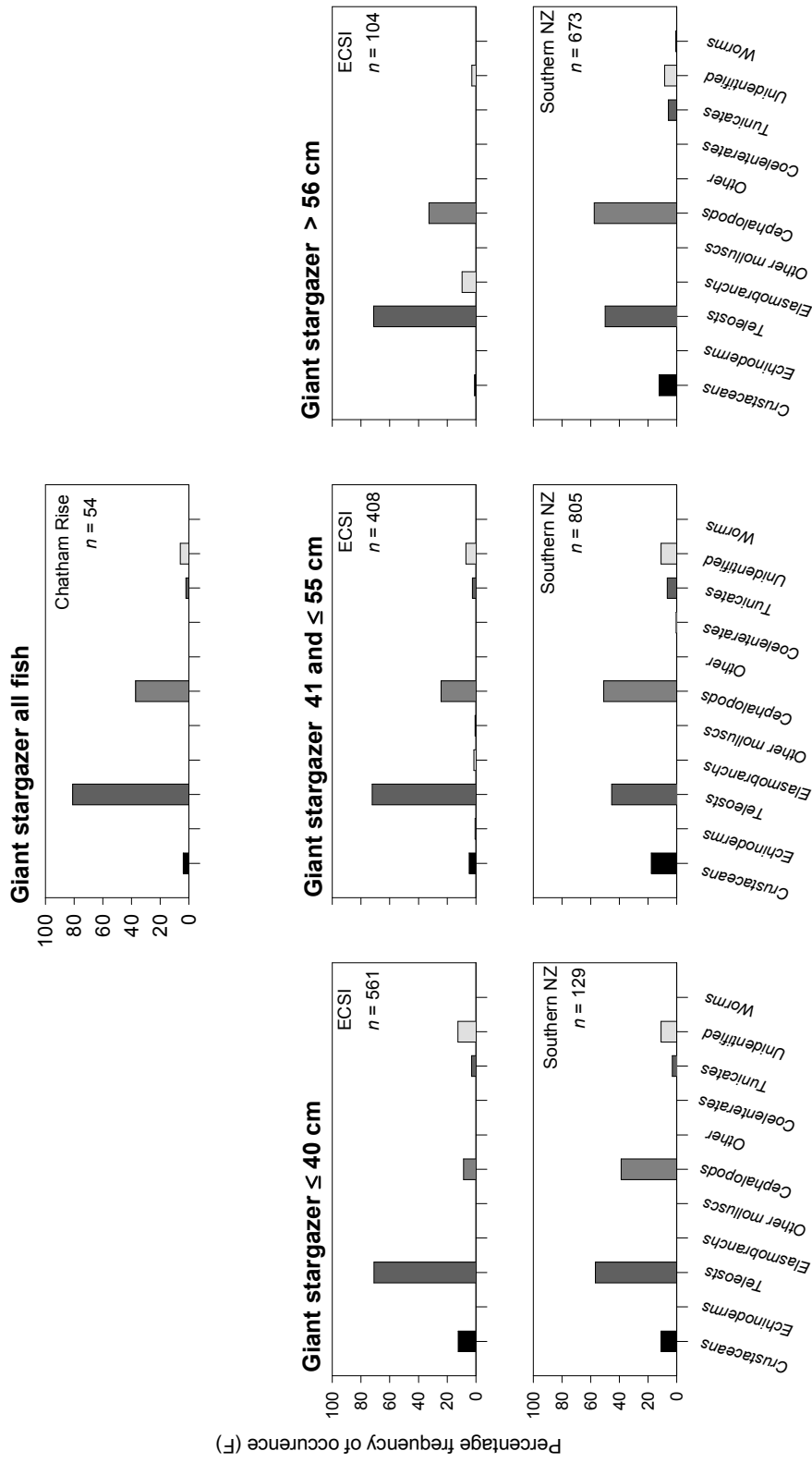


Figure B12c. The importance of major prey groups in the diet of giant stargazer examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

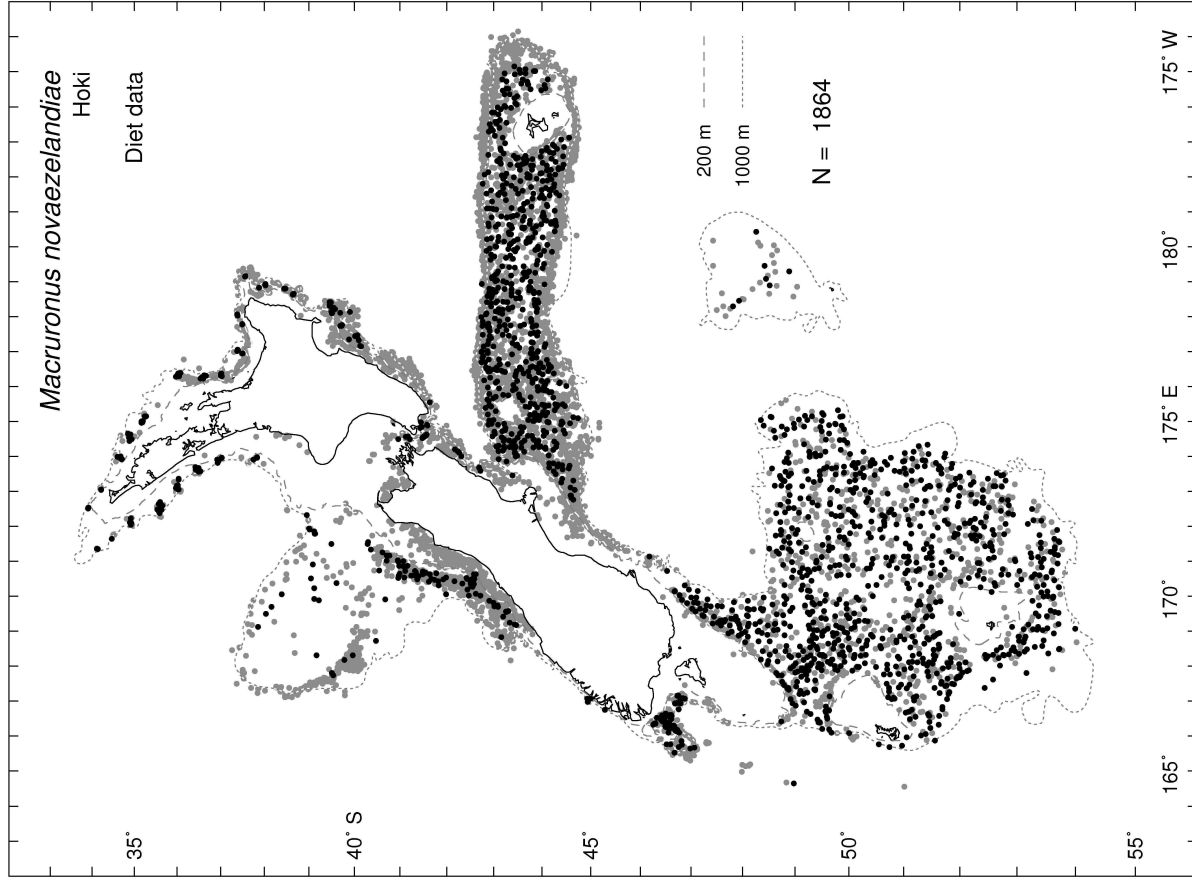


Figure B13a. The distribution of all hoki (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

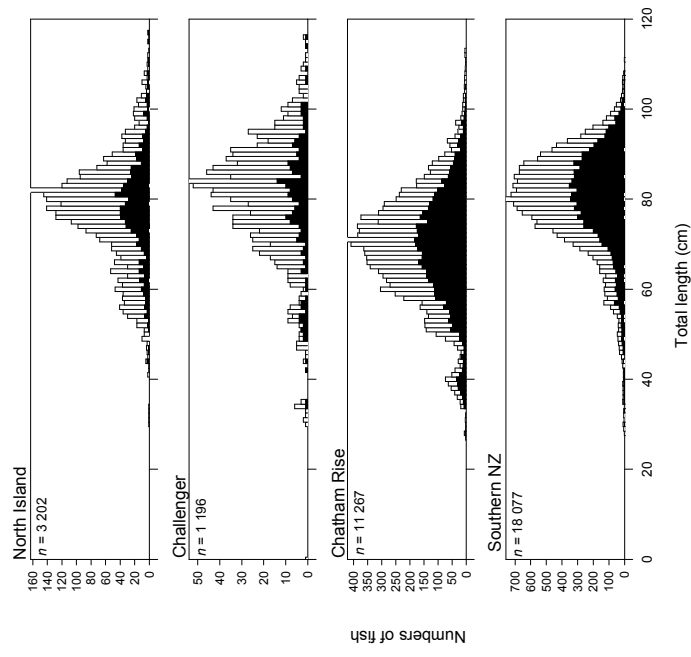


Figure B13b. The length frequency of hoki where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

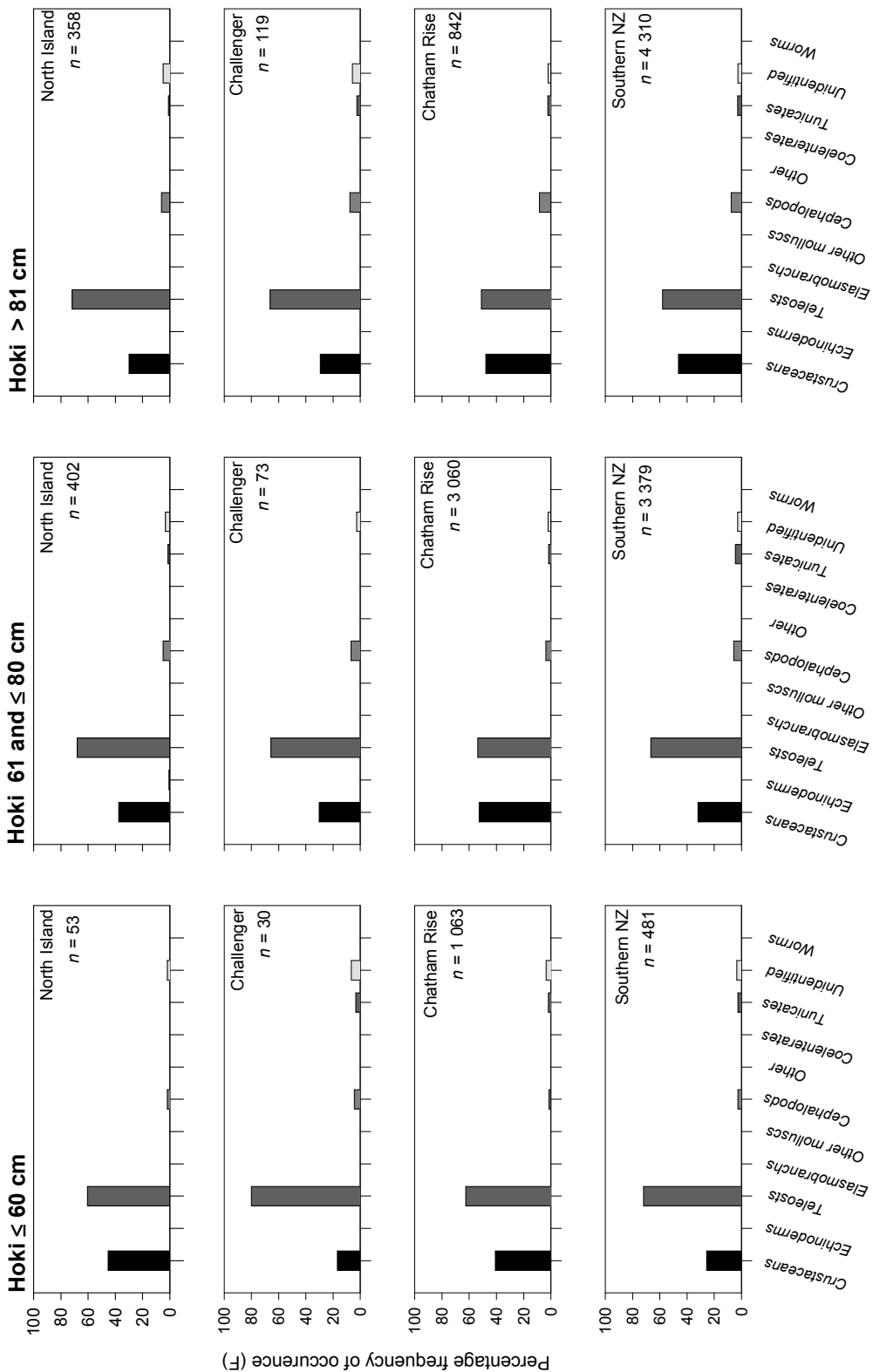


Figure B13c. The importance of major prey groups in the diet of hoki examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

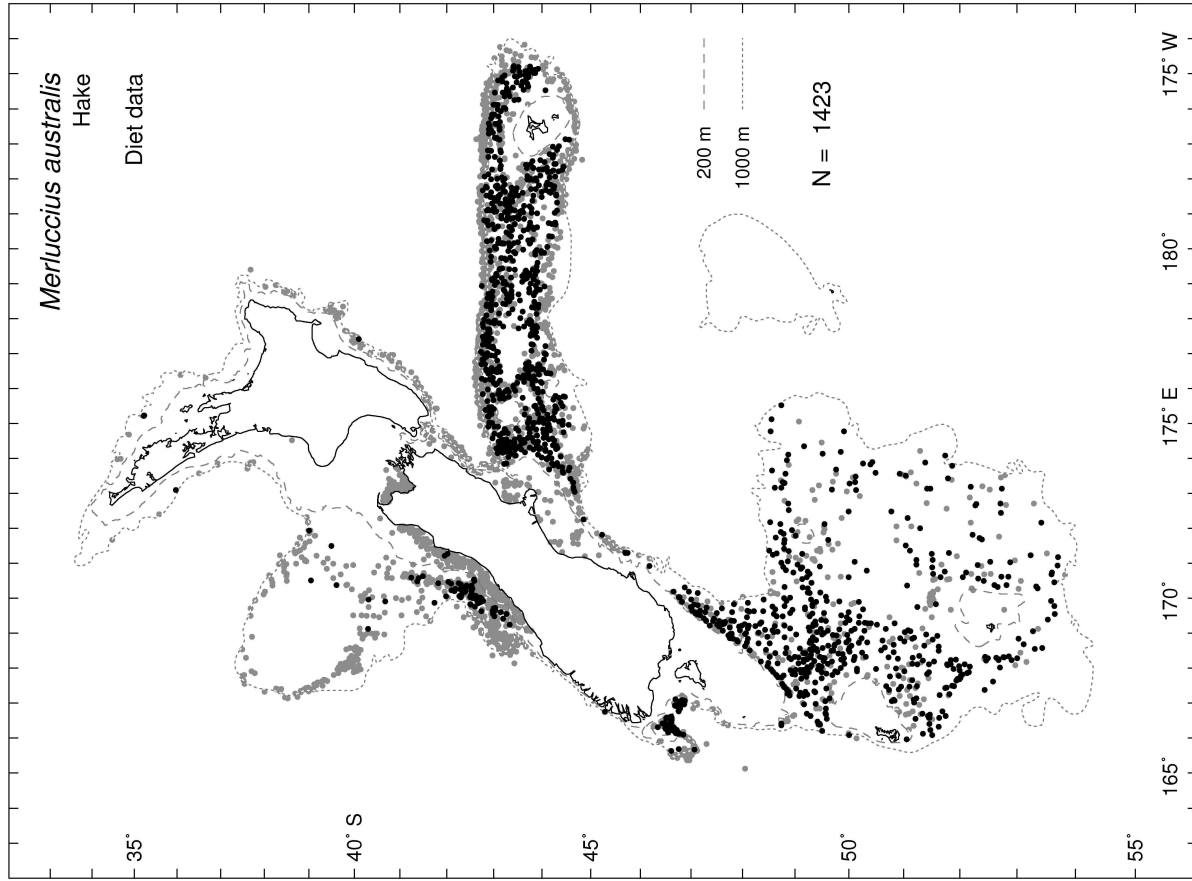


Figure B14a. The distribution of all hake (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

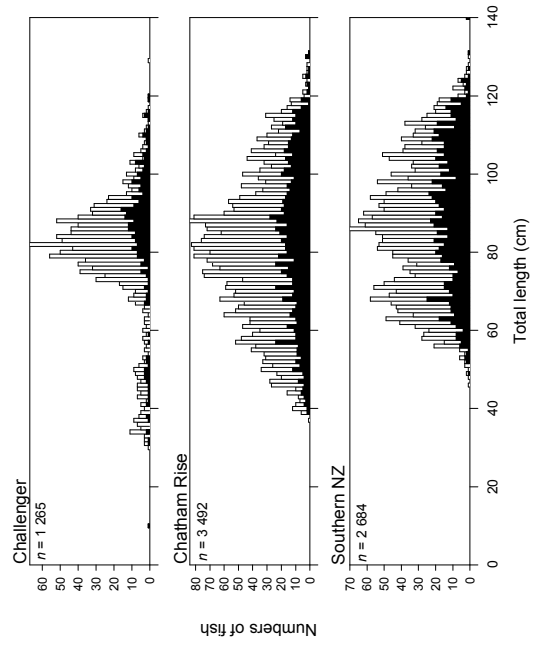


Figure B14b. The length frequency of hake where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. *n*, number of fish examined for diet.

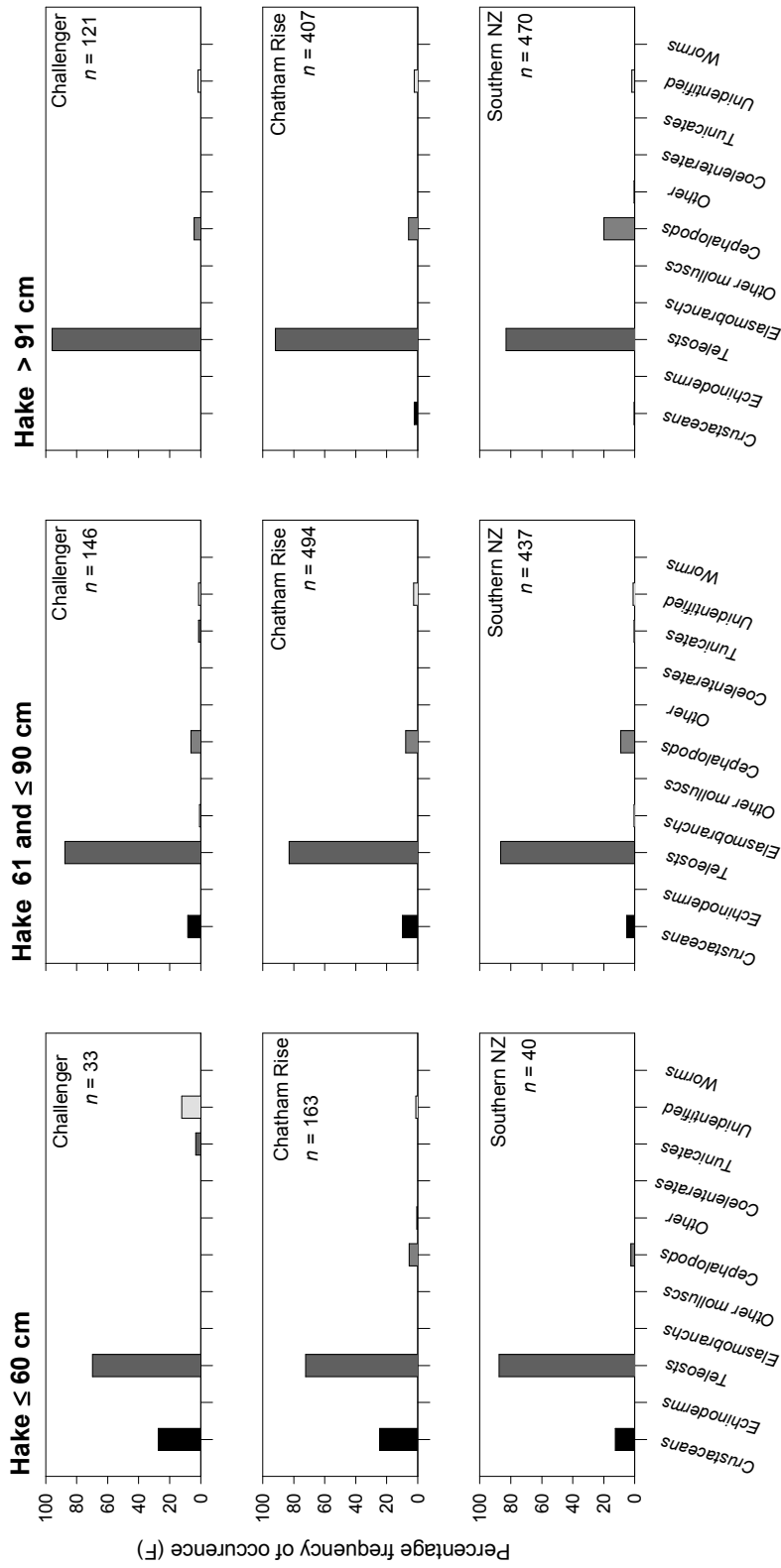


Figure B14c. The importance of major prey groups in the diet of hake examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

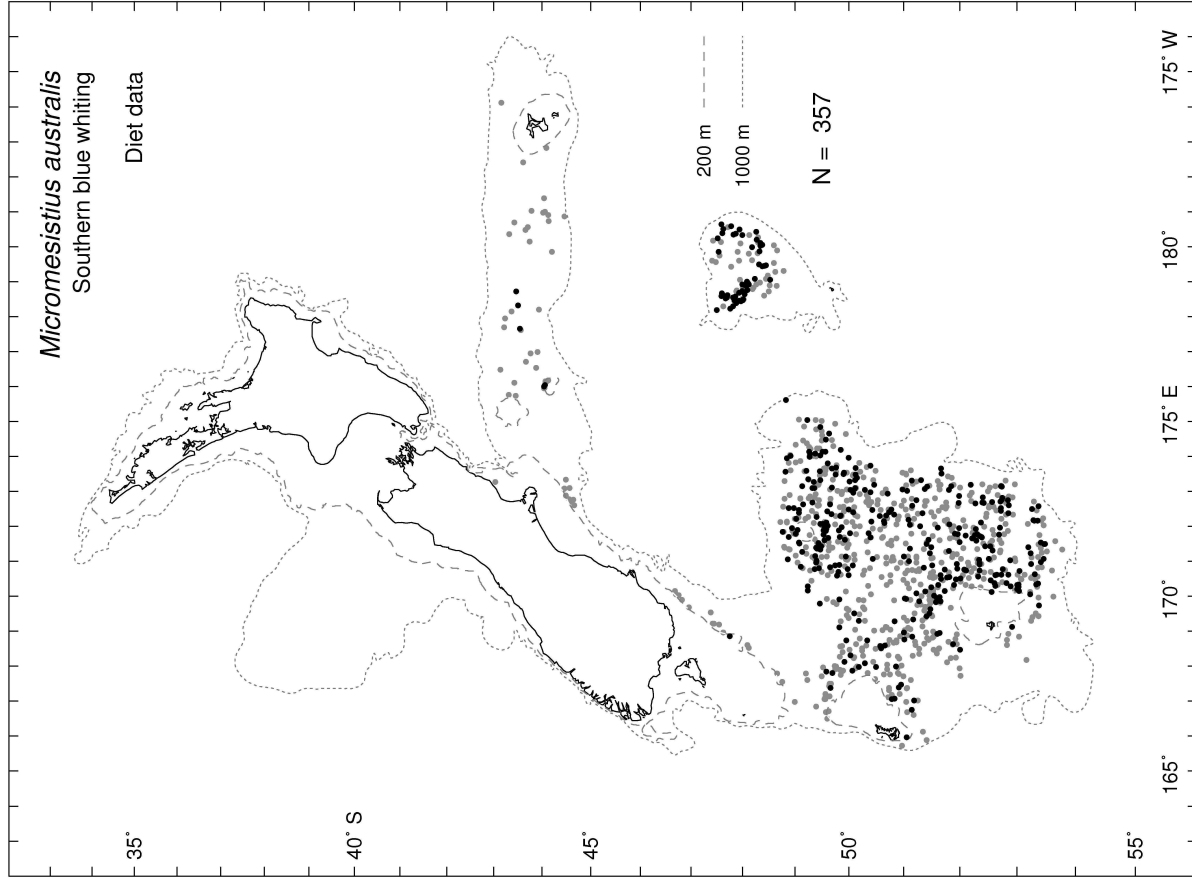


Figure B15a. The distribution of all southern blue whiting (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

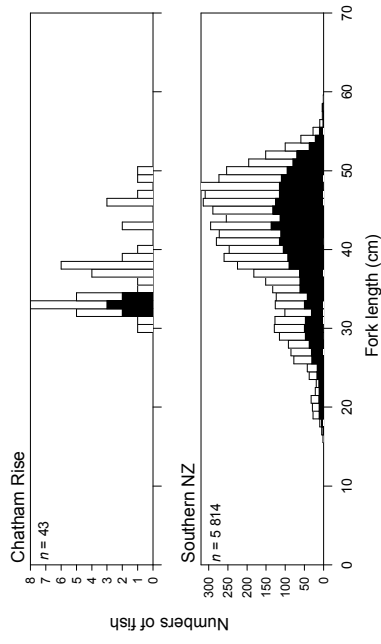


Figure B15b. The length frequency of southern blue whiting where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

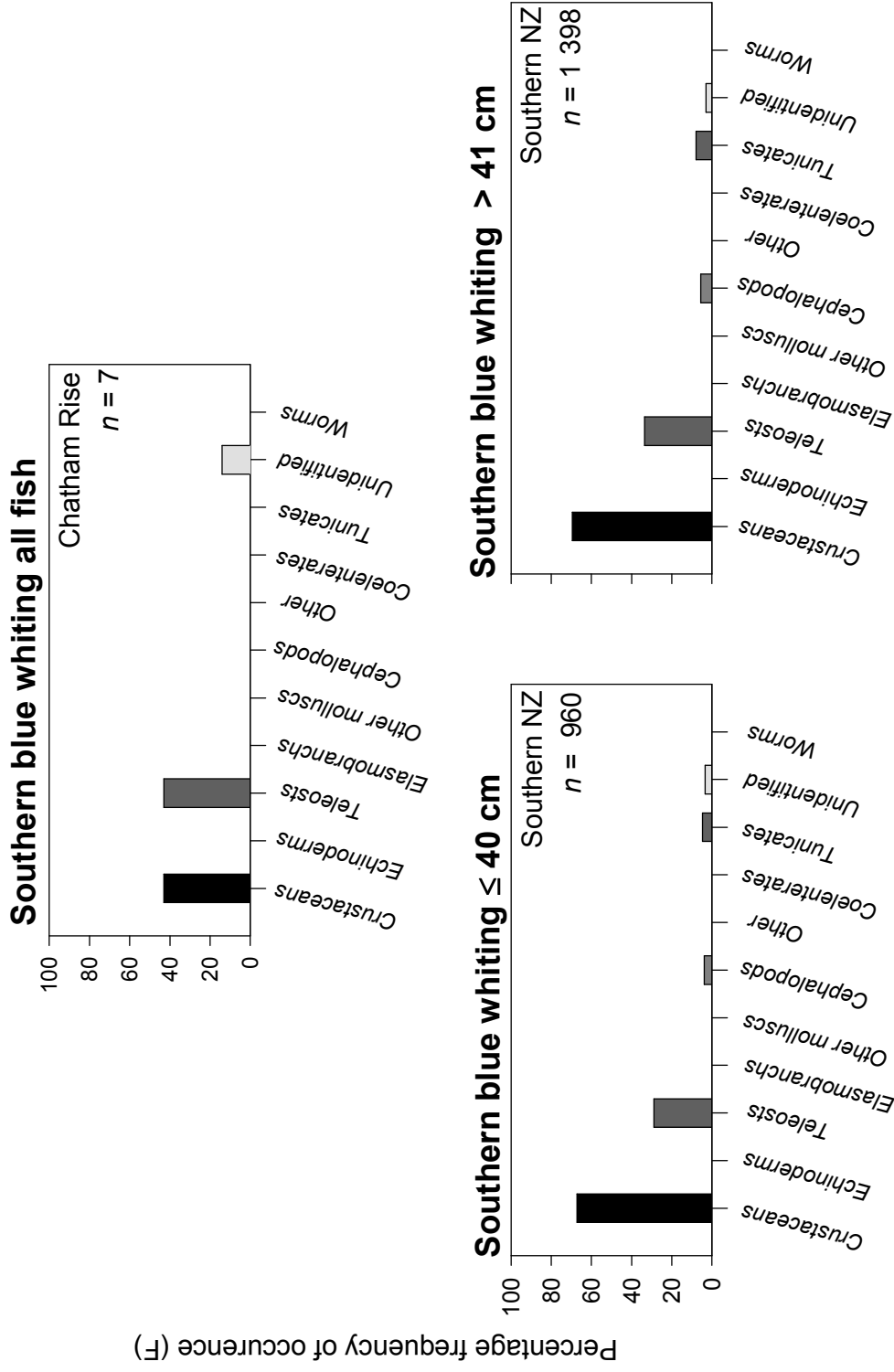


Figure B15c. The importance of major prey groups in the diet of southern blue whiting examined on research trawl surveys. Fish size groups are arbitrary designations. Areas are defined on p. 9. n, number of fish examined for diet.

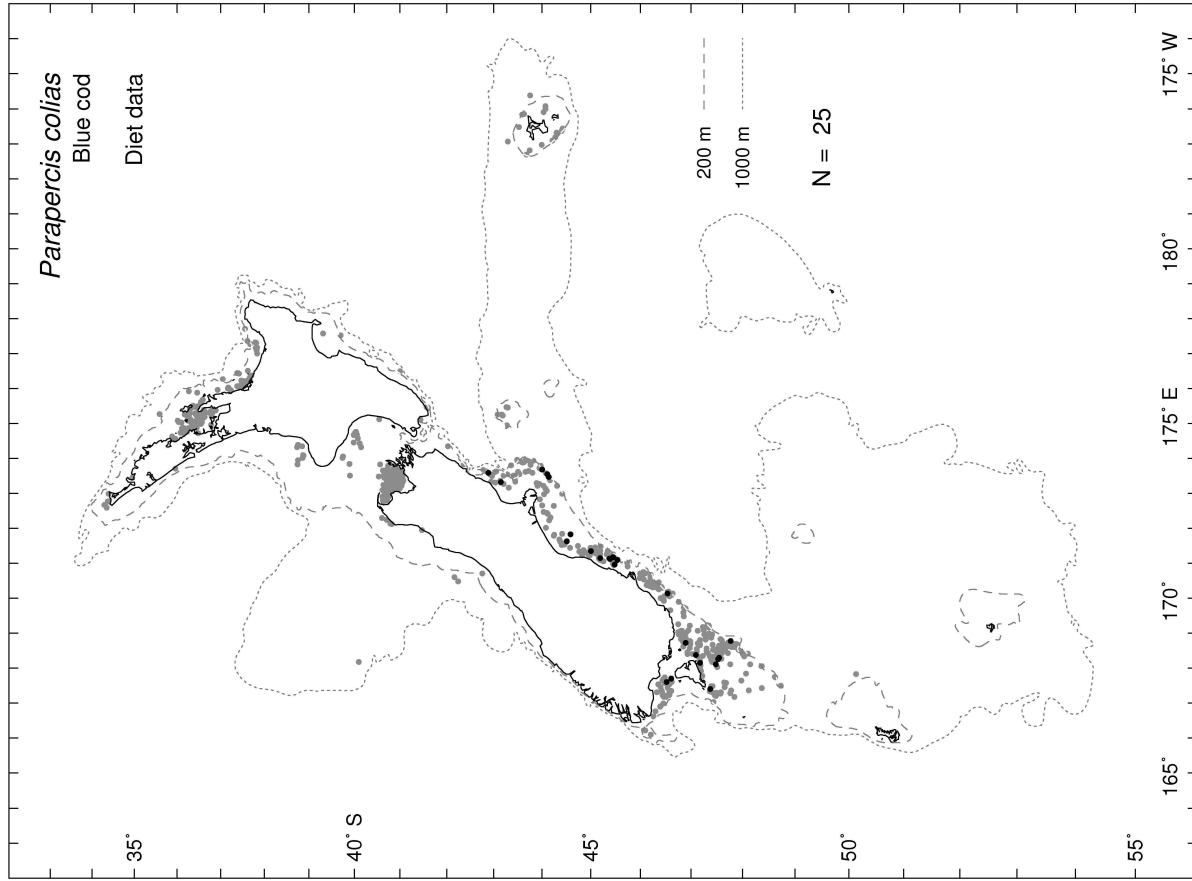


Figure B16a. The distribution of all blue cod (left panel) caught (grey dots) and those examined for diet (black dots) in research trawls 1960–2000. N, number of fish examined for diet.

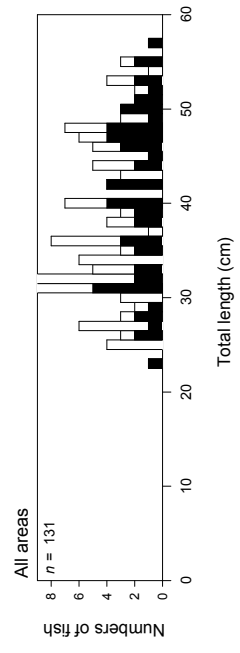


Figure B16b. The length frequency of blue cod where feeding data was recorded. Fish with empty stomachs are presented as white bars and fish containing prey items as black bars. Areas are defined on p. 9. n, number of fish examined for diet.

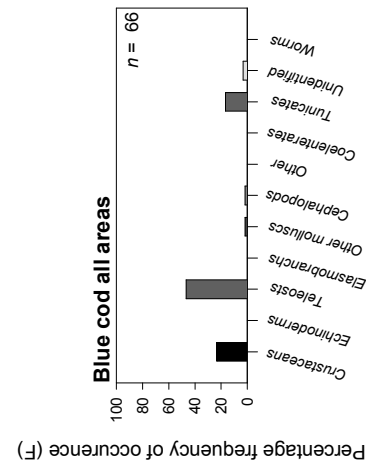


Figure B16c. The importance of major prey groups in the diet of blue cod examined on research trawl surveys. Fish size groups are arbitrary designations. n, number of fish examined for diet.



